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Effect of beech (Fagus sylvatica L.) rhizosphere on phosphorous availability in soils at different altitudes (Central Italy)

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1 Abstract

Phosphorus (P) is an important nutrient for plant growth but its availability in soil is limited. 2 Although plants are able to respond to the P shortage, climatic factors might modify the soil-plant-3 4 microorganisms system and reduce P availability. In this study we evaluated the rhizosphere effect of beech (Fagus sylvatica L.) in forest soils of Apennines mountains (central Italy) at two altitudes 5 (800 and 1000 m) and along 1° of latitudinal gradient, using latitude and altitude as proxies for 6 temperature change. Specifically, we tested if 1) soil organic C, total N, and organic and available P 7 8 decrease with increasing latitude and altitude, and 2) the rhizosphere effect on P availability becomes more pronounced when potential nutrient limitations are more severe, as it happens with 9 increasing latitude and altitude. The results showed that the small latitudinal gradient has no effect 10 on soil properties. Conversely, significant changes occurred between 800 and 1000 m above sea 11 level, as the soils at higher altitude showed greater total organic C (TOC) content, organic and 12 13 available P contents, and alkaline mono-phosphatases activity than the soils at lower altitude. Further, at the higher altitude, a marked rhizosphere effect was detected, as indicated by greater 14 15 concentration of TOC, water extractable organic C, and available P, and its fulfillment was mainly attributed to the release of labile organics through rhizodeposition. The availability of easy 16 degradable compounds in the rhizosphere should foster the mineralization of the organic matter 17 with a consequent increase of available P. Hence, we speculate that at high altitude the energy 18 supplied by the plants through rhizodeposition to the rhizosphere heterotrophic microbial 19 community is key for fuelling the rhizospheric processes and, in particular, P cycling. 20

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28 Abstract

Phosphorus (P) is an important nutrient for plant growth but its availability in soil is limited. 29 Although plants are able to respond to the P shortage, climatic factors might modify the soil-plant-30 microorganisms system and reduce P availability. In this study we evaluated the rhizosphere effect 31 of beech (Fagus sylvatica L.) in forest soils of Apennines mountains (central Italy) at two altitudes 32 (800 and 1000 m) and along 1° of latitudinal gradient, using latitude and altitude as proxies for 33 34 temperature change. Specifically, we tested if 1) soil organic C, total N, and organic and available P decrease with increasing latitude and altitude, and 2) the rhizosphere effect on P availability 35 becomes more pronounced when potential nutrient limitations are more severe, as it happens with 36 37 increasing latitude and altitude. The results showed that the small latitudinal gradient has no effect on soil properties. Conversely, significant changes occurred between 800 and 1000 m above sea 38 level, as the soils at higher altitude showed greater total organic C (TOC) content, organic and 39 40 available P contents, and alkaline mono-phosphatases activity than the soils at lower altitude. Further, at the higher altitude, a marked rhizosphere effect was detected, as indicated by greater 41 42 concentration of TOC, water extractable organic C, and available P, and its fulfillment was mainly attributed to the release of labile organics through rhizodeposition. The availability of easy 43 degradable compounds in the rhizosphere should foster the mineralization of the organic matter 44 with a consequent increase of available P. Hence, we speculate that at high altitude the energy 45 supplied by the plants through rhizodeposition to the rhizosphere heterotrophic microbial 46 community is key for fuelling the rhizospheric processes and, in particular, P cycling. 47

48

49 1. Introduction

50 Phosphorus (P) is one of the most critical nutrients for the growth of plants and microorganisms, 51 and is present in soil both in inorganic and organic forms. The inorganic P includes primary 52 minerals (e.g., apatites, strengite) and secondary minerals such as calcium-, iron-, and aluminum-53 phosphates (Shen et al., 2011), while the organic P is mainly made up of phospholipids,

orthophosphate monoesters, nucleic acids, teichoic acids and phosphonates (Rubæk et al., 1999; 54 55 Kruse et al., 2015). The content and distribution of these different P forms do not have a constant trend along the soil profile (Makarov et al., 2004; Chiu et al., 2005: Backnäs et al., 2012). In 56 grassland and beech forest soils located on north-facing slope of southern Swiss Alps, Beck and 57 Elsenbeer (1999) found that the variation with depth of the organic P concentration mainly depends 58 on type and age of vegetation, and soil characteristics. Then, Chiu et al. (2005) studied grassland 59 60 and coniferous forest soils and found that the inorganic orthophosphate concentration decreased, but orthophosphate monoesters increased with increasing soil depth. The generally low availability of P 61 in soil is due to the scarce solubility of P-bearing compounds, both for inorganic and organic forms 62 63 (Hinsinger, 2001). For example, in the soil solution P exists as orthophosphate anions and dissolved forms of organic P, with concentrations that range from 0.01 to 3.0 mg 1^{-1} (Frossard et al., 2000). 64 However, plants are able to respond to P deficiency by root-exudation of organic acids that increase 65 66 P availability in the soil close to the roots (Ström et al., 2002; Oburger et al., 2009; Zhao et al., 2010). Furthermore, the simple organic compounds comprising the plant exudates enhance the 67 activity of the microorganisms that, in turn, favours P availability by i) soil acidification due to the 68 69 CO₂ produced through respiration, and *ii*) releasing organic acids and phosphatase enzymes (Marschner et al., 2011). A great part of the complex interactions among P-bearing compounds, 70 plant and microorganisms occurs in the small soil volume between fine roots and earthy material, 71 the rhizosphere, where most of the chemical, biochemical and biological reactions take place (e.g., 72 Hinsinger et al., 2003; Richter et al., 2007; Lambers et al., 2009). Because of its high sensitivity to 73 the environmental conditions (Turpault et al., 2007), the biogeochemical processes occurring in the 74 75 rhizosphere can be strongly affected by climate. Indeed, climate (temperature, precipitation amount and pattern) is one of the key soil forming factors that inherently controls the soil profile 76 77 development (Darwish and Zurayk, 1997; Fernández Sanjurjo et al., 2003; Riebe et al., 2004), soil 78 microbial activity (Qiu et al., 2005; Devi and Yadava, 2006), soil organic matter dynamics (Jobbagy and Jackson, 2000; Brevik, 2013), and macro- and micro-nutrients cycles (Butler et al., 2012; 79

Vincent et al., 2014; Zhang et al., 2014). For example, in a phytotron experiment, Kumar et al. 80 (2011) found that higher temperatures increased the amount of available P, but decreased organic P 81 in the rhizosphere of wheat. Instead, as far as we are aware, in vivo studies on the P dynamics in the 82 rhizosphere of forest species at different temperature have never been achieved, although they could 83 help better understanding the resilience of forest ecosystems with respect to global (and more 84 regionally localized) warming. The objective of this study was to provide novel field and laboratory 85 information on this argument. To assess the influence of temperature on the rhizospheric P pool, we 86 selected one of the main diffused forest species in Europe, i.e. the European beech (Fagus sylvatica 87 L.). By using latitude and altitude as proxies for temperature change (Vincent et al., 2014), we 88 89 contrasted the associated changes in European beech rhizosphere and bulk soil collected from the different horizons of forest soils at two altitudes (800 and 1000 m) on three mountains located 90 within 1° of latitudinal gradient in central Italy. Specifically, we tested the hypotheses that: 1) soil 91 92 organic C, total N, and organic and available P decrease with increasing latitude and altitude, and 2) the rhizosphere effect on P availability becomes more pronounced when potential nutrient 93 94 limitations are more severe, as it happens with increasing latitude and altitude. The above two hypotheses were tested on rhizosphere and bulk soil from the three study areas through measuring 95 of organic C, total N, organic and available P, and phosphatase activities; additionally, ³¹P-NMR 96 97 analyses were performed to assess the different forms of soil P.

98

99 **2. Materials and methods**

100 *2.1. Study sites*

As study areas, three calcareous massifs were selected on the Apennines chain (central Italy): Mount Terminillo (42°28' N, 12°59' E), Mount San Vicino (43°19' N, 13°03' E), and Mount Acuto (43°28' N, 12°41' E) (Figure S1 of the Supplementary Materials). For each area, two European beech (*Fagus sylvatica* L.) forests were chosen on the north-facing slopes at about 800 and 1000 m above sea level (a.s.l.). A description of the environmental conditions of each site is reported in

Table 1. Here it suffices to summarize that the three areas have a similar mean annual air 106 temperature (MAAT) that varies from about 10°C at 800 m, to about 9°C at 1000 m a.s.l. 107 Following the latitudinal transect, coldest and warmest months showed a contrasting trend as, going 108 north, the mean of the coldest month (January) decreased of $1.6-2.0^{\circ}$ C, while that of the warmest 109 month (July) increased of 1.6-1.7°C. In all the areas, the mean temperature for both coldest and 110 warmest months is lower in the soils at 1000 m than in those at 800 m a.s.l. of 0.6-1.0 and 1.0-111 112 1.1°C, respectively. The mean annual precipitation, similar at both altitudes, do not follow a latitudinal transect, and is the highest at Mount Acuto, the lowest at Mount San Vicino. 113

All the forests were coppices in conversion, with the conversion that started from about 20 to about 40 years ago. As indicated by the diameter at breast height (Table 1), the most recent conversions occurred at Mount Terminillo and Mount San Vicino, at 800 and 1000 m a.s.l., respectively; the oldest conversions started at Mount Terminillo for the coppice at 1000 m a.s.l., and at Mount Acuto for the woods at both altitudes. At all sites beech was the dominant tree, with dominances ranging from 80 to 100%. While the soil cover due to litter was always complete, the coverage due to understory ranged from 5 to 50%, with scarce to null signs of erosion.

121 All the soils had developed from limestone rocks with small flintstone layers.

122

123 2.2. Soil sampling

During the winter 2014, at each altitudinal site two profiles were dug within a plot of about 100 m². 124 for a total of 12 profiles (3 latitudes x 2 altitudes x 2 profiles). The rational of the winter sampling 125 126 was that, in this season, root respiration and exudation, and root-associated microorganism activity are at their lowest intensity (Epron et al., 2001; Buée et al., 2005; Meinen et al., 2009; Ruehr and 127 Buchmann, 2010; Calvaruso et al., 2014); hence, more reliable and stable information on the 128 rhizosphere status can be obtained in winter rather than in more dynamic seasons like spring or 129 summer. Each profile was opened at 50-60 cm from the stem of the biggest beeches found in the 130 selected site. Approximately, the age of the trees ranged from about 40 years at Mount Terminillo 131

(800 m a.s.l.) to about 60-65 years at Mount Acuto. However, the age of the tree is of secondary importance as, for the protocol adopted to obtaining the rhizosphere samples (see below), we considered only the fine roots, which activity is little dependent on the age of adult plants as they are renewed every few year (Trumbore and Gaudinki, 2003; Agnelli et al., 2014).

The profiles were dug till the parent rock, and the soils were morphologically described according 136 to Schoeneberger et al. (2012). As a whole, the soils at 800 m a.s.l. showed a solum made of the 137 following sequence of horizons, with respective mean thicknesses (standard deviations in 138 parentheses): O = 7.2 cm (2.4), A = 7.0 cm (3.2), AB = 7.0 cm (1.5), Bw1 = 13.2 cm (3.7), Bw2 = 13.2 cm (3.7)139 12.8 cm (4.7), Bw3 = 26.5 cm (8.9), Bw4 = 18.3 cm (4.2). The soils at 1000 m a.s.l. showed the 140 141 following solum: O = 10.2 cm (6.2), A = 10.3 cm (3.2), AB = 14.2 cm (7.7), Bw1 = 19.8 cm (9.4), Bw2 = 10.0 cm (4.7). The mean thickness (excluding the O horizons) of the solum was 68.8 cm 142 (36.3) at 800 m, and 49.2 cm (19.9) at 1000 m. The underneath C horizons are not part of the solum 143 144 and were excluded from sampling. The litter was made by O horizons that were typical of the amphimus type of humus (Baize and Girard, 2008), which are present in soils with well-developed 145 146 O horizons rich of pedofauna, and resting on A horizons with well-developed crumb structure.

Roughly, in the soils at 800 m the fine earth content ranged from 85% in the A horizon to 50% in the Bw3 and Bw4 horizons, while in those at 1000 m it went from 80% in the A horizon to 35-40% in the Bw1 and Bw2 horizons. All the soils had a *mesic* soil temperature regime, and an *udic* soil moisture regime, and were classified as Mollisols or Inceptisols (Table 1) according to the Soil Survey Staff (2014).

For each profile, a large amount of sample (at least 3 kg) from each mineral horizon forming the*solum* was collected and stored in a portable refrigerator for the transport to the laboratory.

154

155 *2.3. Sample preparation*

Within one week from the sampling, the beech rhizosphere of each sample was isolated by pickingup the roots together with the adhering soil (Cocco et al., 2013; Massaccesi et al., 2015). The roots

with a diameter larger than 2 mm were discarded. After a light shaking to detach the weakly adhering soil particles, which were then added to the bulk soil (i.e., the soil not strictly adhering to the roots), the remaining soil material firmly adhering to the fine roots was considered as rhizosphere and was recovered by further shaking and gentle brushing. An aliquot of the field moist rhizosphere and bulk of each horizon was stored at 4°C for measuring the phosphatase activities, while the remaining material was air-dried and sieved through a 2 mm mesh to be used for the chemical and spectroscopic analyses.

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166 2.4. Chemical analysis

The soil pH was determined potentiometrically in water (pH_{H2O}) and in 1 M KCl solution (pH_{KCl}) 167 (solid:liquid ratio of 1:2.5) after 30 minutes of stirring by a combined glass-calomel electrode. Total 168 organic C content (TOC) was estimated by K-dichromate digestion, heating the suspension at 180 169 170 °C for 30 minutes (Nelson and Sommers, 1996), and total N content was determined by a Carlo Erba EA1110 dry combustion analyzer (Carlo Erba Instruments, Italy). To measure the water 171 172 extractable organic C (WEOC), 1 g of sample was placed into a plastic container, submerged with 173 distilled water (solid:liquid ratio 1:10) and shaken overnight with an orbital shaker (140 rpm). The mixture was left to rest for a while, centrifuged at 1400 g for 10 minutes, and then filtered through 174 175 Whatman 42 filter paper. The resulting solution was analyzed with a TOC-500A (Shimatzu, Japan) analyzer after the addition of a few drops of concentrated H₃PO₄ to eliminate carbonates. The total P 176 was determined by the ignition method, and the organic P was calculated by difference between the 177 total and inorganic P content (Kuo, 1996). Available P was estimated according to Olsen et al. 178 (1954). 179

Acid and alkaline mono-phosphatase activities were determined according to Tabatabai (1994). Briefly, in a 50-ml flask, 1 g of sample was added with 0.2 ml of toluene, 4 ml of modified universal buffer (at pH 6.5 and pH 11 for the acid and alkaline phosphatases, respectively), and 1 ml of 0.025 M *p*-nitrophenyl phosphate (*p*-NPP) solution. The mixture was incubated at 37 °C for 1 h so to induce the transformation of *p*-NPP into *p*-nitrophenol (*p*-NP) via phosphatase. After incubation, 1 mL of 0.5 M CaCl₂ and 4 mL of 0.5 M NaOH were added to the samples, mixed for few seconds, and then filtered through a Whatman 42 filter paper. The color intensity of the filtrate was measured against a blank at 420 nm by a Lambda EZ 150 UV/VIS Spectrometer (Perkin Elmer, USA). The results are expressed as μg of *p*-NP produced by 1 g of soil per h of incubation ($\mu g p$ -NP g⁻¹ h⁻¹).

For the ³¹P-NMR measurements, which were run only on the A horizons, the extracts were obtained 190 191 by shaking 5 g of sample with 100 ml of a solution containing 0.25 M NaOH and 0.05 M EDTA for 16 h at room temperature, in the dark. The mixture was centrifuged at 10,000 g for 30 min and the 192 liquid phase, once separated from the precipitated, was freeze-dried. The freeze-dried NaOH-EDTA 193 extracts were re-dissolved in 0.1 ml of 10 M NaOH solution and 0.5 ml of D₂O, and transferred to a 194 5-mm NMR tube for the analysis. The ³¹P-NMR spectra were obtained using an Arance 600 MHz 195 196 NMR spectrometer (Bruker, USA) operating at 243 MHz, with an acquisition time of 0.673 s, and a delay time of 0.5 s. For each sample 24,576 scans were run. Peak assignments were according to 197 198 Turner et al (2003), and the intensities of signals were determined by integration.

199 The accuracy of the run soil analyses follows: pH in water, 0.14; pH in KCl, 0.11; TOC, 2.05 g kg⁻¹; 200 total N, 0.19 g kg⁻¹; WEOC, 0.024 g kg⁻¹; total P, 35 mg kg⁻¹; organic P, 24 mg kg⁻¹; available P, 2.5 201 mg kg⁻¹; acid and alkaline mono-phosphatase activity, 5 μ g *p*-NP g⁻¹ h⁻¹.

202

203 2.5. Statistical analyses

To test the effect of each variable (latitude, altitude, soil horizons, and soil fractions) on the soil properties we performed canonical redundancy analyses (RDA). The RDA model was tested for significance using 999 random permutations. The variations of soil properties as a function of latitude, altitude, soil horizons, and soil fractions were assessed by a Principal Component Analysis (PCA). For each property, all the data were standardized prior the RDA and PCA by subtracting the mean and dividing by the standard deviation.

The RDA indicated a lack of significant effect for latitude (ANOVA, F= 2.39, P=0.057), whereas 210 211 the effects of altitude (ANOVA, F= 15.84, P=0.001), soil horizons (ANOVA, F=4.13, P=0.001) and soil fractions (ANOVA, F=62.86, P=0.001) on the soil properties were significant. Because of this, 212 only these three latter significant variables were considered in further detail in our study, and the 213 three latitudinal areas were therefore considered as replicates. Consequently, the analytical results 214 obtained from the two samples collected for each horizon at each latitude and altitude were 215 averaged, and these averages used as replicates so to have n=3. The data were checked for the 216 normality of the distribution and the homogeneity of the variances by Shapiro-Wilk and Levene 217 tests, respectively, and, if necessary, transformed by the Box and Cox (1964) procedure. To assess 218 219 significant differences among altitudes, horizons and soil fractions (rhizosphere and bulk soil), three-way ANOVA was performed, and the comparison of means was assessed by Fisher's LSD 220 post-hoc test (P < 0.05). Box plot diagrams were used to show the obtained data. The line inside each 221 222 box represents the median. The bottom and top of the box are the first and third quartiles, while the upper and lower whiskers indicate the minimum and maximum values, respectively; the + sign 223 224 within each box plot indicates the average.

225 The statistical analyses were performed using R software (R Core Team, 2014).

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227 **3. Results**

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228 3.1. PCA
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The PCA scoring plot (Figure 1) showed variations of the soil properties between rhizosphere and bulk and between the soils at 800 and 1000 m a.s.l., and identified the axes 1 and 2 that explained about 53 % and 24 % of the variation, respectively. All the soil parameters showed positive correlation with PC1 (Figure S2a of the Supplementary Materials), with TOC, alkaline and acid phosphatase activities, total N, and available and organic P contributing for more than 83 % to the variability (Table 2). While the pH_{H2O} and pH_{KCl} only slightly affected PC1, they explained 57 % of the variability along PC2 axis. The scoring plot (Figure 1) showed that the soil properties were affected mainly by altitude and soil fractions and to a lesser extent by soil horizons. Moreover, the PCA indicated that the differences between the rhizosphere and bulk were somewhat greater at 1000 m than at 800 m a.s.l..

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240 *3.2. Soil pH, organic C and total N*

In the soils at both altitudes, pH_{H2O} and pH_{KCI} remained relatively constant throughout the profile, and no significant difference was observed between rhizosphere and bulk soil (Figure 2a, b). However, both pH_{H2O} and pH_{KCI} from A to Bw2 horizons were higher at 1000 m than at 800 m altitude.

As expected, TOC content tended to decrease with depth throughout all the soils (Figure 3a). On a horizon by horizon comparison between the two altitudes, TOC content was greater at 1000 m than at 800 m. Then, TOC concentration was always higher in the rhizosphere than in the bulk soil of the same horizon. In contrast to TOC, WEOC concentration was similar at 800 and 1000 m a.s.l. (Figure 3b); however, whereas for the soils at 800 m no difference in WEOC content occurred between rhizosphere and bulk soil, at 1000 m the rhizosphere had consistently a greater WEOC content than the bulk soil.

The total N content decreased with depth in all the soils and, as for the TOC, on a horizon by horizon comparison it was higher at 1000 than at 800 m (Figure 3c). Rhizosphere and bulk showed similar total N contents at both altitudes, with the exceptions of the Bw1 horizon of the soils at 800 m, where the rhizosphere displayed a greater total N content than the bulk soil.

256

257 *3.3. Total, organic and available phosphorus contents, and phosphatase activities*

The total and organic P content was much higher in the soils at 1000 m than in those at 800 m (Figure 4a, b). At both altitudes, rhizosphere and bulk soil generally did not show significant differences, with the exceptions of the Bw1 horizon for total and organic P, and Bw3 horizon for total P, always for the soils at 800 m. The available P content (Figure 4c) was also generally higher in the soils at 1000 m than in those at 800 m; further, in the higher altitude soils the rhizosphere had a greater concentration of available P than the bulk soil in all the horizons, while in the soils at 800 m this occurred in four over six horizons (A, AB, Bw2, and Bw3). The TOC:organic P ratio (Table S1 of the Supplementary Materials) showed a decreasing trend for both rhizosphere and bulk soil at 800 m, but it was not significantly different between the two altitudes and between the fractions, with the exception of the Bw4 horizon at 800 m where the rhizosphere showed a greater value than that of the bulk soil.

In the soils at 800 m, alkaline and acid phosphatase activities (Figure 5a, b) decreased with 269 increasing depth from the A to the Bw1 horizons (Bw2 for acid phosphatase), to remain rather 270 271 constant more in depth. At 1000 m, both enzymatic activities decreased from the surface to the AB horizon, to remain rather constant in the horizon underneath. By contrasting horizon by horizon, the 272 alkaline phosphatase activity (Figure 5a) was greater in the soils at 1000 m, where small differences 273 between rhizosphere and bulk were observed only in the Bw1 horizon. The acid phosphatase 274 activity (Figure 5b) was similar in the A and AB horizons at the two altitudes, and greater at 1000 m 275 276 than at 800 m in the Bw1 and Bw2 horizons. The rhizosphere showed higher acid phosphatase activities than the bulk soil in the Bw1, Bw2 and Bw4 horizons at 800 m, and in the A and Bw1 277 horizons at 1000 m. 278

279

280 *3.4.* ³¹*P*-*NMR spectroscopy*

The ³¹P-NMR spectra (Figure S3 of the Supplementary Materials) indicated that the orthophosphate monoesters (from 3 to 6 ppm of the chemical shift) were the dominating P forms of the spectral area, followed by inorganic orthophosphates (from 5.9 to 6.5 ppm), orthophosphate diesters (from -2 to 0 ppm) and pyrophosphates (from -3.5 to -5.5 ppm). The spectra showed similar patterns for rhizosphere and bulk soil. Between the two altitudes, the orthophosphate diesters were more abundant at 1000 m than at 800 m for both rhizosphere and bulk soil (Figure 6).

287

288 **4. Discussion**

289 *4.1 Altitude and rhizosphere effect on pH, C and N*

The PCA performed on all the measured soil properties showed a marked difference between 290 rhizosphere and bulk of the studied soils, indicating that the beech roots induce a rhizosphere effect. 291 Similar results were reported by several other authors. For example, Wang et al. (2001) studied the 292 soil solution chemistry of beech and Norway spruce and found lower pH and nutrient 293 concentrations in the rhizosphere than in the bulk soil solutions. Esperschütz et al. (2009), studying 294 295 carbon fluxes from beech trees into the rhizosphere found that during the growing season the rhizosphere had a higher amount of dissolved organic matter than the bulk soil. Calvaruso et al. 296 297 (2011) found that the rhizosphere of several tree species including beech was enriched in organic C, N, Ca, Mg and K with respect to the bulk soil, and the extent of the rhizosphere effect depended on 298 the tree species. Finally, Cesarz et al. (2013) accomplished a rhizotron experiment and found that 299 beech induced a strong rhizosphere effect mostly because of roots exudates and associated 300 microbial community. Further, the PCA scoring plot (Figure 1) highlighted a clear distinction 301 302 between the soils at 800 m and those at 1000 m, confirming the effect of altitude on soil properties, 303 as commonly was reported by many authors (e.g., Tsui et al., 2004; Seibert et al., 2007; Cioci et al., 2008). 304

305 Even though the pH was the parameter that most strongly differentiated rhizosphere and bulk on the basis of PCA2, contrasting the two fractions for each horizon, no difference occurred for both 306 pH_{H2O} and pH_{KCI} . The weak rhizosphere effect on soil pH was ascribed, at least partly, to the 307 sampling period as the reduced winter root activity probably lessened the release of H⁺ at the soil-308 root interface in response to the lowered uptake of cations (Hinsinger et al., 2003). However, 309 Calvaruso et al. (2014), in a study conducted on soil solutions of an acidic beech forest soil, 310 measured lower pH values in the rhizosphere than in the bulk soil during winter, the reverse in 311 spring, and similar pH values in the two fractions in fall. In our case, the scarcity of pH differences 312 between rhizosphere and bulk could be also ascribed to the nature of the parent material, whose 313

carbonate content makes the rhizosphere little susceptible to acidification as *i*) the carbonate dissolution neutralizes the activity of the released protons (Agnelli et al., 2016), and *ii*) the rate of chemical weathering and decarbonation in mountain soils is minor than at lower altitudes (Riebe et al., 2004). However, these observations support the higher pH values that we found for both rhizosphere and bulk of the soils at the higher elevation.

319 The TOC and total N content followed a similar decreasing trend with depth for both rhizosphere 320 and bulk soil at the two altitudes. This result was expected for soils with well-developed A horizons characterized by a strong crumb structure whose cement is mostly made of organics. According to 321 several authors (e.g., Lemenih and Itanna, 2004; Dai and Huang, 2006; Follett et al., 2012), the 322 323 higher content of TOC and total N at 1000 than at 800 m was related to climatic conditions and, among those, mostly to the temperature, which drops with increasing altitude; in contrast, the 324 precipitation generally do not follow such a clear altitudinal trend (Körner, 2007; Griffiths et al., 325 326 2009). The accumulation of higher amounts of organic matter at 1000 m may have been fostered by the combination of the different effects caused by the lower temperature on the plant biomass 327 328 production and on the activity of the soil microbial community. Although the plant biomass 329 production decreases with increasing altitude (Zianis and Mencuccini, 2005), the higher amounts of TOC happens because of a lower microbial activity, which is due to colder soil temperatures 330 331 occurring at higher elevations (Blume et al., 2002; Xu et al., 2014). In our study areas, a larger TOC content was found in the rhizosphere than in the bulk in the soils at both altitudes, as it has generally 332 been observed in many different environments (e.g., Turpault et al., 2007; Zhao et al., 2010). As at 333 334 1000 m the rhizosphere had a larger WEOC content than the respective bulk soil in all the horizons, whereas this did not occur at 800 m, it suggested that beech was able to induce a stronger 335 rhizosphere effect at the higher altitude. The enrichment of WEOC in the rhizosphere is mainly 336 attributed to rhizodeposition processes (Chiang et al., 2006; Tuason and Arocena, 2009), which 337 supply most of the energetic substrates for the rhizosphere microbial community (Koranda et al., 338 2011; Cesarz et al., 2013). According to Kuzyakov (2002, 2010), the availability of easily 339

degradable compounds (and their consumption by the microflora) triggers a further mineralization 340 of stable organic matter through the so-called "priming effect". As a consequence, the rhizosphere 341 priming effect induced by root exudation boosts the organic matter cycling and the microbially-342 mediated release of nutrients (Kaiser et al., 2011). This process may play a key role in the 343 rhizosphere of the soils at higher altitude, where a scarcer soil development and a generally lowered 344 microbial activity due to climatic constraints limited nutrients availability. The allocation of plant 345 346 resources in the rhizosphere through rhizodeposition can therefore be seen as a strategy of the plants to overcome ecosystem (nutrient availability) restrictions (Boddy et al., 2008; Massaccesi et al., 347 2015). The fact that at 800 m the WEOC/TOC ratios for both rhizosphere and bulk were higher than 348 349 those at 1000 m (data not shown), indicated a more active organic matter cycling occurring at the lower altitude, where the microbial activity has less limitations because of a milder temperature 350 351 (Pietikäinen et al., 2005; Creamer et al., 2015).

352

4.2. Altitude and rhizosphere effect on P availability and related enzymatic activities

354 In agreement with many previous observations on forest soils, the total P content was mostly made up of organic P, which showed mean contents that are commonly reported for mountain soils (e.g., 355 Makarov et al., 2004; Talkner et al., 2009). According to Turner et al. (2002) and Stutter et al. 356 (2015), the greater concentration of organic P in the soils at 1000 m was attributed to the higher 357 abundance of organic matter in these soils. The dependence of the organic P concentration on soil 358 organic matter content was also confirmed by the TOC:organic P ratio, which showed no difference 359 360 between the two altitudes and between rhizosphere and bulk soil, although this ratio could be also affected by the amount of available P (Makarov et al., 2004) and P plant uptake (Saikh et al., 1998). 361 As the organic P is the main source of available P in soil (Turner et al., 2014), the larger 362 concentration of available P for both rhizosphere and bulk in the soils at 1000 m was ascribed to a 363 greater alkaline phosphatase activity all throughout these soils, which was probably induced by the 364 larger WEOC and TOC contents (Lemanowicz and Krzyzaniak, 2015; Stutter et al., 2015), and the 365

higher pH of these soils, all factors able to promote the alkaline phosphatase activities (Nannipieri 366 367 et al., 2011). For the Bw1 and Bw2 horizons, also the acid phosphatase activity was higher in the soils at 1000 m, but in this case the fostering factors were probably only the high contents of 368 WEOC and TOC. The question on whom, between plants roots and rhizosphere microbial 369 community, was the main responsible for the different production of phosphatase in the soils at 800 370 and 1000 m remains open. Previous studies found that the acid and alkaline phosphatase activities 371 372 are higher in the rhizosphere than in the bulk soil (Marschner et al., 2005; Zhao et al., 2007; Shi et al., 2011) because of rhizodepositions, which fuels the microbial activity and enhances the 373 production of extracellular enzymes in the rhizosphere (Brzostek et al., 2013). However, also the 374 375 plant roots produce enzymes (Nannipieri et al., 2011; Rejsek et al., 2012). To this regard, in a mesocosm experiment with young Fagus sylvatica L., Hofmann et al. (2016) found that plant 376 phosphatases contributed lesser than microbial ones to the total phosphatase activity in P-rich soil. 377 378 In our case, we hypothesized that the greater alkaline phosphatase activities found in the rhizosphere of the soils at 1000 m was due to the release from both beech roots and a specifically 379 380 adapted microbial biomass. In the soils at 1000 m, the larger phosphatase activities and the higher availability of easily degradable organics (WEOC) probably counterbalanced the minor microbial 381 activity caused by the lower temperature. 382

383 The mineralization of organic P compounds by hydrolysis of mononucleotides, sugar phosphates, phosphoproteins and inositol-phosphates via phosphatases is the process responsible for the release 384 of inorganic orthophosphates, which are part of the available P and can be taken up by living 385 386 organisms (Turner and Haygarth, 2005; Nannipieri et al., 2011). However, no substantial difference in phosphatase activities was found between rhizosphere and bulk soil at both altitudes. Because of 387 388 this, the larger available P content in the rhizosphere of all the horizons of the soils at 1000 m was 389 attributed to an intense P cycling that, in the soil close to the roots, was triggered by the exudation of labile organic compounds (Ström et al., 2002; Palomo et al., 2006); these would have promoted 390 the microbial activity and the consequent release of P and other nutrients through the organic matter 391

mineralization (Kuzyakov, 2010). Further, the higher content of available P in the rhizosphere compared to the bulk soil may be also favoured by the P uptake, which induces desorption of P from mineral surfaces (Gerke, 2015). However, a P solubilisation due to root release of protons following nutrient uptake and of organic acids cannot be excluded. This latter explanation would be valid even if no pH difference was detected between rhizosphere and bulk soil because of the buffering action of the calcareous parent material, and the complex spatial and temporal pattern of micro-niches occurring in the rhizosphere (Richter et al., 2007; Faget et al., 2013).

399 The hypothesis of a more intense P cycling occurring in the soil close to the roots was not supported by the results of the ³¹P NMR analysis, as no significantly different proportions of P forms between 400 401 rhizosphere and bulk soil were detected. This absence of differences between the fractions may be partly attributed to the soil variability (above and belowground) occurring even at the same altitude. 402 The most represented form of the P pools was that made of orthophosphate monoesters, either at 403 404 800 and 1000 m. This fact was rather expected as inositol-phosphates (which are the main component of the P monoesters) are strongly stabilized in soil by abiotic reactions with minerals, 405 406 which thereby hinder their biological degradation and favour their accumulation in soil (Turner et 407 al., 2002; Giaveno et al., 2010). The only difference between the sites showed by the NMR spectra was the larger proportion of orthophosphate diesters in the soils at 1000 m. As orthophosphate 408 409 diesters are considered indicators of microbial P cycling (Stutter et al., 2015), their greater amount at 1000 m than at 800 m supports the occurrence of a general stronger P turnover in the soils at the 410 higher altitude. This is consistent with the concept that high-altitude ecosystems, due to more 411 412 pronounced nutrient limitations when compared to lower altitude ones, are more dependent on mineralization of soil organic matter by microbial community (Parfitt et al., 2005). Indeed, when 413 the amount of available P in the soil is limited, P is largely immobilized in organic forms 414 415 (Bünemann et al., 2012).

416

In this work we evaluated the rhizosphere effect of beech in forest soils of central Italy, at two 418 altitudes (800 and 1000 m) and along 1° of latitudinal gradient. While the small latitudinal gradient 419 did not affect the rhizosphere and bulk soil properties, significant changes occurred between the 420 soils at the two altitudes, and a marked rhizosphere effect was detected in those at 1000 m. These 421 differences were observed in spite of the spatial and morphological heterogeneity of the forest soils, 422 which possibly affected the extent of the rhizosphere effect at the different altitudes. However, the 423 fact that we tested our hypotheses in natural soils, where above and belowground heterogeneities 424 are independent variables, may be considered as an added value to our research, and indicates that 425 sampling designs able to control the main climatic and physiographic variables may allow obtaining 426 427 significant results studying in vivo rhizosphere.

The clear rhizosphere effect that was found at the higher altitude and it was expressed by a greater 428 TOC, WEOC and available P concentrations, was attributed to rhizodepositions, which represent 429 430 the main source of energetic substrates for the rhizospheric microbial community-. The greater availability of easily degradable compounds (WEOC) in the rhizosphere should boost the 431 mineralization of organic matter, which in turn may favour the mineralization of the organic P 432 433 forms and increase the amount of available P (Figure 7). Therefore, we speculated that at high altitude the energy supplied by the plants through rhizodeposition to the rhizosphere heterotrophic 434 435 microbial community is key for fuelling the rhizospheric processes and, in particular, P cycling.

Our results suggested that an increase of the air temperature of about 1°C, which is expected globally for the year 2050 (IPCC, 2013), and that is equivalent to the temperature shift between our study sites at 800 and 1000 m a.s.l., might cause a shortage of available P in the high altitude beech forest soils.

440

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