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### **Developing a methodology for ecosystem assessment of Tam Giang-Cau Hai lagoon and estuaries of Thua Thien Hue and Quang Tri Provinces, Central Vietnam**

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I, Nguyen Vu Bao Chi, declare that this thesis and the work presented in it are my own and has been generated by me as the result of my own original research.

The title of the thesis is: “Developing a methodology for ecosystem assessment of Tam Giang-Cau Hai lagoon and estuaries of Thua Thien Hue and Quang Tri Provinces, Central Vietnam”

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Signed: Nguyen Vu Bao Chi

Date: September 17<sup>th</sup>, 2019

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## Table of Content

<b>Academic thesis: Declaration of Authorship.....</b>	<b>1</b>
<b>Acknowledgments .....</b>	<b>2</b>
<b>Table of Content.....</b>	<b>3</b>
<b>List of Figures.....</b>	<b>5</b>
<b>List of Tables .....</b>	<b>10</b>
<b>List of Acronyms .....</b>	<b>11</b>
<b>Abstract.....</b>	<b>12</b>
<b>Riassunto.....</b>	<b>17</b>
<b>Introduction.....</b>	<b>18</b>
<b>Objective of this thesis .....</b>	<b>20</b>
Why wetlands matter .....	20
Background .....	22
A framework linking environmental science to human welfare .....	24
Ecosystem functions and services.....	25
Classification of ecosystem functions and services .....	26
The pilot study area and project justification.....	27
Objectives of the study .....	28
Regional background and critical issues.....	28
<b>Presentation of data .....</b>	<b>31</b>
Materials and methods .....	31
<i>Methods of water and sediment samples collection.....</i>	<i>31</i>
<i>Methods of phytoplankton sample collection.....</i>	<i>31</i>
<i>Methods of sediment sample collection for foraminiferal studies .....</i>	<i>32</i>
<i>Water quality analyses.....</i>	<i>33</i>
<i>Grain-size analyses.....</i>	<i>35</i>
<i>Microalgae and phytoplankton analyses .....</i>	<i>35</i>
<i>Foraminifera analyses .....</i>	<i>35</i>
Results of analyses.....	36
<i>Foraminifera biology and ecology: proxies for environmental health.....</i>	<i>36</i>
<i>Analyses of foraminiferal assemblages from Cua Tung and Cua Viet estuaries,</i> <i>April 2017 .....</i>	<i>42</i>

<i>Analyses of samples from Cua Tung and Cua Viet estuaries, September 2017</i> .....	46
<i>Analysis of environmental data 2006-2018</i> .....	58
Discussion on characteristics of lagoon and estuarine environments of Central Vietnam.....	60
<i>Climate of Central Vietnam</i> .....	60
<i>Considerations on foraminiferal ecology</i> .....	61
<i>Tam Giang-Cau Hai lagoon physiography and ecosystems</i> .....	61
<i>The Tam Giang Cau Hai sub-basins and active river system</i> .....	62
<i>Tam Giang-Cau Hai lagoon hydrology</i> .....	63
<i>Tam Giang-Cau Hai lagoon water chemistry</i> .....	64
<i>Hydro-chemical characterization of the water masses and lagoon seasonality</i> ....	67
<i>Tam Giang-Cau Hai lagoon biocenotic mapping and distribution of main commercial species</i> .....	71
<i>Tam Giang-Cau Hai lagoon ecosystem zoning</i> .....	72
<i>Non-living resources (soil, groundwater, mineral resources)</i> .....	75
<b>Summary and conclusions</b> .....	<b>80</b>
Assessment of status of coastal environment .....	80
<i>Status of quality of coastal environment</i> .....	80
<i>The sources of environmental hazards, pollution and degradation in the coastal area</i> .....	81
<i>Critical issues relating to the quality of wetland habitats of Central Vietnam</i> .....	83
<i>Vulnerable ecosystems in Tam Giang-Cau Hai lagoon and coastal areas and risk assessment maps</i> .....	83
<i>Assessment of status of coastal biological resources</i> .....	85
<b>References</b> .....	<b>90</b>
<b>Annexes</b> .....	<b>96</b>
Annex 1 – Comprehensive datasets of surveys 2006-2013, secondary data considered in this synthesis .....	96
Annex 2 – Comprehensive datasets of surveys 2015-2018, primary data collected during the implementation of this PhD project.....	142

## List of Figures

Figure 1 – Geography and political boundaries (communes) of the Tam Giang-Cau Hai lagoon and its surrounding territory. The whole Tam Giang-Cau Hai lagoon belongs to the Thua Thien Hue Province; the 33 riparian communes belong to five districts. In the insert, the location of the Province and the Tam Giang-Cau Hai lagoon in the National context....	21
Figure 2: Location of Quang Tri Province and related river network .....	29
Figure 3 - Sampling stations in Quang Tri Province, April 2017.....	32
Figure 4 - Van Veen grab (left), Van Dorn sampler (center) and 50-mL-capacity Falcon conical tubes (right). .....	33
Figure 5 – Diagram showing a generalized foraminifer life cycle. Note the alternation between a haploid megalospheric form and a diploid microspheric form (redrawn after Goldstein, S.T., 1999).....	40
Figure 6 - Formation of new chamber and laminate structure in perforate foraminifera. (Erez, J., 2003, reprinted by Cusak, M., and Freer, A., 2008).....	40
Figure 7 - Model for temporal fusion of the seawater vacuoles (SWV) with the plasma membrane (pm) near the active calcification site. Seawater enters the cell through deep invaginations or semi-open vacuoles (sov). A vacuole is pinched off and undergoes alkalization by one of the cellular proton transport mechanisms. This SWV concentrates inorganic carbon by diffusion of CO <sub>2</sub> (aq) from the acidic cytosol into the alkaline SWV. This process is enhanced by adjacent mitochondria (m) and by the acidic vesicles (av) that release CO <sub>2</sub> . The [Ca <sup>2+</sup> ] and [CO <sub>3</sub> <sup>2-</sup> ] enriched vacuoles fuse with the cell membrane and supply the ions for calcification. The vacuoles are then resealed and release their content apically, away from the growing crystals (after Bentov, S., <i>et al.</i> 2009).....	41
Figure 8 - Relative abundances of the main living benthic foraminifera genus in Cua Tung and Cua Viet estuaries, Quang Tri Province, April 2017.....	43
Figure 9 - Percentage of foraminifera species in Cua Tung (QT5, QT6, QT7) and Cua Viet (QT1, QT2, QT3, QT4) estuaries, Quang Tri Province. Survey April 2017. TC, total calcareous; TMC, total Miliolids calcareous; TA, total agglutinated.....	44
Figure 10 - Proportion of tests of living benthic foraminifera in Cua Tung and Cua Viet estuaries, Quang Tri Province, April 2017 .....	45

Figure 11 - Correlation between salinity and benthic foraminiferal diversity (richness) in samples from Cua Viet and Cua Tung estuaries, Quang Tri Province, Survey April 2017 .....	46
Figure 12 - Sediment grain-size variations in 15 stations within 4 sampling areas.....	47
Figure 13 - Grain size fraction compositions in samples from the Cau Hai inlet (Thua Thien Hue Province).....	48
Figure 14 - Grain size fraction compositions in Tam Giang .....	48
Figure 15 - Grain size fraction compositions in Cua Viet .....	49
Figure 16 – Relative abundances of the main living benthic foraminifera genus in Cua Viet estuary, Quang Tri Province, September 2017 .....	53
Figure 17 – Relative abundances of the main living benthic foraminifera genus in Tam Giang lagoon, Thua Thien Hue Province, September 2017 .....	53
Figure 18 - Relative abundances of the main living benthic foraminifera genus in Cau Hai lagoon, Thua Thien Hue Province, September 2017 .....	54
Figure 19 - Percentage of foraminifera species from samples from Cua Tung and Cua Viet estuaries of Quang Tri Province, Tam Giang-Cau Hai lagoon, Thua Thien Hue Province. TC, Total Calcareous; TMC, Total Miliolids Calcareous; TA, Total Agglutinated.....	54
Figure 20 – Proportion of tests of living benthic foraminifera in Cua Viet estuary, Quang Tri Province, September 2017.....	55
Figure 21 – Proportion of tests of living benthic foraminifera in Tam Giang lagoon, Thua Thien Hue Province, September 2017 .....	56
Figure 22 - Proportion of tests of living benthic foraminifera in Cau Hai lagoon, Thua Thien Hue Province, September 2017 .....	56
Figure 23 - Percentage of foraminifera species from samples from Cua Tung and Cua Viet estuaries of Quang Tri Province, Tam Giang-Cau Hai lagoon, Thua Thien Hue Province. TC, Total Calcareous; TMC, Total Miliolids Calcareous; TA, Total Agglutinated.....	57

Figure 24 - Correlation between in salinity and benthic foraminiferal diversity (richness) of Cua Tung and Cua Viet estuaries of Quang Tri Province and of Tam Giang-Cau Hai lagoon, Thua Thien Hue Province.....	58
Figure 25 - The Tam Giang-Cau Hai lagoon sub-basins and their tributary river network. From northwest, the O Lau river discharging into the Tam Giang-Cau Hai lagoon; the Bo and Huong rivers, directly discharging into the Eastern Sea though the Thuan An inlet and the Truoi river, discharging into the Cau Hai sub-basin.....	62
Figure 26 – Diffusion and dispersion of the spots during the first 48 hours of simulation,forced by the astronomical tide of April 2011.....	65
Figure 27 - Results of cluster analysis of environmental parameter (dissolved oxygen, pH, salinity and temperature) for the four surveyed seasons of 2006 (April, spring; May, dry season; August, beginning of rainy season; November, rainy season but unusually dry in 2006). Cluster description is: 1, fresh water characterized by very low salinity (1 psu), lowest pH (except November), highest DO in early spring and fall and lowest, in summer (May and August); 2, low-salinity (average 7.8 psu) inner-lagoon water, as low as 3.5 psu during rainy months of April and August and as much as 13 psu in dry months of May and November; pH fluctuates between 7.5 during rainy months and 8.1 during dry month; 3, low-salinity brackish water, with intermediate salinity values ranging from 10 and 19 psu, lowest dissolved oxygen concentration and pH higher than 8; 4, low-salinity brackish waters, with stable salinity of 14 psu, temperature above 30 degree, pH average around 8 and concentration of dissolved oxygen systematically higher than cluster 3 (in excess of 7 mg/l). During the driest month of May, these intermediate waters are indistinguishable from those of cluster 5; 5, typical brackish water with salinity ranging between 16 and 18 psu, temperature normally higher than 30 degrees and average pH around 8. The concentration of dissolved oxygen is higher than 7 mg/l and salinity raised over 22 psu during driest month of May; 6, cooler marine waters (average 26.5 °C in April; lower than cluster 6); 7, marine waters, with salinity higher than 20 psu, as much as 30 psu during the dry months of May and November; temperature between 28,5 and 30,5 °C. Lowest concentration of dissolved oxygen in November, similarly to cluster 3.....	69



Figure 28 - Natural and anthropic habitat distribution of Tam Giang-Cau Hai. Legend: 1, hydrophytes prairies; 2, sea grass fields; 3, aquaculture ponds; 4, tidal flats; 5, sandy-mud lagoon floor; 6. muddy lagoon floor; 7, meadows flooded in the rainy season. .... 71

Figure 29 - Distribution map of characteristic Tam Giang-Cau Hai biota and selected commercial species (natural fish population, macrophytobenthos, macrozoobenthos). Legend: 1, *Meretrix sp. (Tria)*, fresh-water shrimps, fresh-water weeds; 2, *Meretrix sp. (Tria)*, sea crab, thorned black crab (*ram*), *Potunus sp.* (blue crab or *ghe*); 3, *Saccostrea sp.*, *Meretrix sp.*, Cerithidea; 4, *Meretrix sp.*, *Potunus sp.* (blue crab or *ghe*), fresh-water *Tilapia*, *Siganus*, sea fishes; 5, *Meretrix sp.*, *Potunus sp.* (blue crab or *ghe*), seagrass, Veneridae and others bivalves, *Corbicula*, *Siganus*, *Metapenaeus ensis*, shrimps (*tôm go mo*); 6, Cerithidae (*ôc*), seaweeds; 7, Cerithidae (*ôc*), *Corbicula*, seaweeds; 8, *Solen sp.*, *Metapenaeus sp.*, Cerithidae; 9, *Corbicula*, *Portunus* (blue crab or *ghe*), *Pteria*, *Saccostrea*, Penaeidae. .... 72

Figure 30 - Integrated ecosystem zoning map derived from the distribution of water body characteristics (cluster analysis), lagoon-bottom features, sediment types and selected biota compositions (legend for fields a-I, see Fig. 29). Ecosystem legend: 1, Upper Tam Giang, O' Lau eco-zone; 2, Central Tam Giang, distal O' Lau eco-zone; 3, Lower Tam Giang eco-zone; 4, Huong river inner delta eco-zone; 5, Sam Chuon eco-zone; 6, Upper Thuy Tu marine relic waters; 7, Lower Thuy Tu channel and western Cau Hai continental domain; 8, Eastern Cau Hai marine and submarine domain, including mixing zone. The lower Tam Giang, the Sam Chuon and eastern Cau Hai show affinities in the way of having low-dynamic, stagnant water masses, largely of continental origin ..... 73

Figure 31 - Simplified land-use map of the territory surrounding the Tam Giang-Cau Hai lagoon. Legend: 1, agricultural areas (rice paddies and cultivated fields) in alluvial plain lowland; 2, aquaculture areas, potential sites of effluent waste-water discharge and sources of organic pollutants (fish feeds, fish metabolism residues etc.); 3, urban (and industrial) areas; 4, semi-natural tropical and subtropical moist broadleaf forests; 5, semi-natural sandy area, with irregular and discontinuous bush coverage; 6, inland waters and sea. .... 76

Figure 32 - Land use/land cover categories and nomenclature defined for coastal Thua Thien Hue Province, modified after CORINE land types classifications..... 77

Figure 33 - Cartoon showing a typical example of coastal phreatic aquifer from sand dunes.....	79
Figure 34 - Hazards classification for the Tam Giang-Cau Hai and the coastal province of Thua Thien Hue. The time scale indicates the time frame and frequency at which hazard may turn into an emergency. ....	81
Figure 35 - Zoning map of the Tam Giang-Cau Hai lagoon based on risk-level towards fishery activities, as assessed in 2004, and range of proposed interventions in support of the sector (as per Decision No. 3677/QD-UB of 2004 of the Thua Thien Hue Provincial People’s Committee).....	84
Figure 36 - Risk maps of the Tam Giang-Cau Hai lagoon representing the risk level evaluated for each of the identified ecosystems (above) and communes (below). Risk levels have been ranked according to three categories: 1, <i>low</i> , whenever emergencies are mild and their effects are likely to be naturally mitigated or nullified in the short term; 2, <i>moderate</i> , whenever emergencies, once they manifest, are circumscribed or controllable and 3, <i>high</i> , whenever emergencies are persistent and/or multiple emergencies can manifest simultaneously. ....	85
Figure 37 - Percent of species distribution in Tam Giang-Cau Hai lagoon identified during the assessments 2006-2018. ....	86
Figure 38 - Distribution of number of aquatic species in the Tam Giang-Cau Hai lagoon sub-basins.....	89

## List of Tables

Table 1 – Parameter measuring and detection methods .....	34
Table 2 - Foraminifera species abundance each station in Cua Viet, Cua Tung estuaries, Quang Tri Province - Survey April 2017.....	42
Table 3 - Diversity of foraminiferal species from samples of Cua Tung and Cua Viet estuaries, Quang Tri Province, survey April 2017. H', Shannon-Weiner Diversity Index; D, Simpson Diversity Index.....	45
Table 4 - Grain-size fraction compositions.....	47
Table 5 - Foraminifera species abundance from samples from Cua Tung and Cua Viet estuaries of Quang Tri Province, Tam Giang-Cau Hai lagoon, Thua Thien Hue Province.....	52
Table 6 - Diversity of foraminiferal species of Cua Tung and Cua Viet estuaries of Quang Tri Province, Tam Giang-Cau Hai lagoon, Thua Thien Hue Province. H', Shannon-Weiner Diversity Index; D, Simpson Diversity Index .....	57
Table 7 - Production per catch session (in tons per hectare) and extrapolated bulk biomass (in tons), May 2006.....	89

## **List of Acronyms**

BOD	Biological Oxygen Demand
DANIDA	Danish International Development Agency
DO	Dissolved Oxygen
ES	Ecosystem Services
FAO	Food and Agricultural Organization
FOBIMO	Foraminiferal Biomonitoring
GoV	Government of Vietnam
GPS	Global Positioning System
ICZM	Integrated Coastal Zone Management
ISQG	Interim Sediment Quality Guidelines
IUCN	International Union for the Conservation of Nature
IUU	Illegal, Unregulated and Unreported
IWRM	Integrated Water Resources Management
MEA	Millennium Ecosystem Assessment
MSC	Monitoring, Surveillance and Control
NGO	Non-Governmental Organizations
OC	Organic Compound
PEL	Permissible Exposure Limit
TCVN	Technical Committee of Vietnam
TEEB	The Economics of Ecosystems and Biodiversity
TN	Total Nitrogen
TP	Total Phosphorus
US EPA	United States Environmental Protection Agency
UN	United Nations
WORMS	World Register of Marine Species

## **Abstract**

The objective of this thesis is to systematize the environmental datasets acquired during a period of nearly two decades at set locations in Tam Giang-Cau Hai lagoon of Thua Thien Hue and Quang Tri Provinces, integrating existing secondary datasets with newly acquired primary information during 2006-2018.

In addition to physico-chemical parameters, biological proxy indicators of ecosystem integrity were incorporated in the database, in attempt to develop a tool-box methodology to reveal the degree of effectiveness of lagoon and wetland ecosystem functions being, long-term and seasonal variations and the degree of environmental stress caused by human activities.

Coastal plains and wetlands have been privileged sites for human civilizations for millennia. Their habitats and resources have been critical to the development and survival of humanity. Wetland ecosystems are part of our natural assets, providing services worth trillions of US dollars every year and making a vital contribution to life. With increasing demographic pressure and modern society craving for water and food resources, not counting threats posed by climate changes, the need to maximize these benefits has never been greater or more urgent.

A fundamental issue in developing countries is to harmonize environmental conservation and social progress. Human conditions are tightly linked to the state of the environment where economic activities and social life are carried out. The Millennium Ecosystem Assessment stemmed from the assumption that a close relationship between welfare and prosperity of a community, its security and resilience is related in a multi-faceted way to the salubrity of the environment and its effectiveness in performing functions and providing services supportive to human life.

The following approach has been followed to set up a framework for the implementation of developmental projects or the drafting of ecosystem-based management policies and plans aiming at both environmental conservation and human well-being:

- Inventory of ecosystems and habitats in a specific locale
- Development of a set of objectively verifiable and quantifiable indicators to appraise the state of ecosystems, their effectiveness in performing functions and

delivering services.

- Establish a science-based framework for environmental monitoring and data acquisition for use of scientists and policy makers
- Establish a GIS-based environmental relational database to serve the purpose of planning, management and policy drafting.
- Develop thematic mapping for planning, tracking environmental changes, land use, soil consumption, illegality and abusive encroachment of urban settlements onto wetland.
- Use remote-sensing techniques to extrapolated point-source information to land and water surfaces

A methodology for each of these components is delivered as output of this thesis, then integrated into an operational tool-box for use of managers and decision makers.

The Tam Giang-Cau Hai lagoon with its 22,000 hectares of water surface and estuary wetlands, with associated deltas and forests, represent the largest lagoon system in Southeast Asia. It is a key area that, as a consequence of post-war rural repopulation, demographic increase, resource overuse, urbanization and industrialization, has undergone substantial changes in the past two decades.

As a consequence of human pressure and encroachment of the natural habitats, there is a perception that ecosystem functions and services have been impaired; however, a clear and science-grounded understanding to what extent the beneficial effects to the riparian communities has been compromised.

This doctoral thesis has been conceived with the objectives of developing a methodology for a science-based analysis of environmental effectiveness and ecosystem-service provision based on:

- available and routinely acquired data,
- standardization, spatialization and consistency assessment of available information,
- ecosystem and habitat identification,
- assessment of resource state and trends
- tool development for data dissemination

The environmental degradation of the Central Vietnam wetlands and increasing pressure on their aquatic resources, together with typhoons, floods and droughts that

yearly strike the province concurred in creating a situation of particular vulnerability for people living on and around it. This situation of persistent threat and creeping decline has been a matter of concern in the past two decades: the application of the methods of science in support of environmental monitoring and control will contribute to a more effective management of the locale in the long term.

Original datasets were collected, in the years 2015-2018, according to a protocol, laboratory procedures and sampling grid compatible with previous campaigns implemented in 2006-2013 whose data are available from the local literature and internal Provincial reports.

A substantial component of the study was dedicated to benthic foraminifera, specific taxonomic category considered of importance as indicators of environmental quality and stress.

In order to assess water quality, the following list of parameters were measured, both directly in the field during sample collection and in the laboratory.

- Water depth of station,
- Turbidity,
- Temperature,
- Flow velocity during tidal flood and ebb,
- Compositional and textural parameters of bottom sediments,
- Total solid and suspended solid present in the water,
- pH,
- Salinity,
- Dissolved oxygen,
- Nitrites and nitrates,
- Ammonia,
- Alkalinity,
- Total phosphorous,
- Total nitrogen,
- Chlorophyll A
- Zoobenthos biomass
- Phytoplankton

- Microalgae and toxic algae
- Total coliforms, E. coli and Vibrio cells

The focus of the thesis is hinged on the assumption/observation that benthic foraminiferal assemblages display an overly sensitivity to environmental changes. The rationale of the investigation is therefore based on a threefold approach:

- Comparative analyses of historical time series of environmental data over the period of a decade.
- Relationships between environment parameter changes and shifts in the composition of foraminiferal assemblages.
- Calibration of foraminifera assemblages as a predictive tool for upcoming environmental changes.

Findings of this dissertation are:

- The Tam Giang-Cau Hai and wetlands of Central Vietnam maintain their rather pristine state compared to other lagoon and deltaic settings of the world, despite the high population density, the strong demand for food resources and on going vigorous development. Historical events of the second half of the past century slowed down the process of industrialization and urbanization, maintaining agriculture and fishery into artisanal conditions and the aesthetic of the natural environment into its untouched beauty.
- Aquaculture has been another major element of perturbation of the natural system: the attempt to promote shrimp production in the nineties generated in cascade negative impacts whose repercussions are still felt today: widespread organic pollution, wiping out of endemic mangrove ecosystem, eliminating inter-subtidal spawning and nursery platform, loss of productivity, break of reproductive cycle of numerous species, loss of biodiversity, hindrance of circulation etc.

Critical issues relating to the quality of wetland habitats of Central Vietnam

- Parameters and variables of water quality, sediments, benthic associations in Tam Giang-Cau Hai lagoon and estuaries of Central Vietnam portray a rather stable situation and a persistent acceptable standard for aquaculture; however, high organic-matter input to open waters relating to biological excretion and excessive feed may create eutrophication and anoxia lethal for life.



- High-impact shrimp monoculture is unsustainable on the long term: alternative methods (e.g. clam farming) need to be studied and piloted to mitigate negative effects of effluents and aquaculture waste waters.
- Livelihood shift is necessary, having short-term impact on household welfare and economy, but unquestionably beneficial on the long term.
- Need of a viable community-based solid-waste collection and treatment plan, including the creation of a community-based collection network at commune level.
- Appropriateness of the future implementation of a wetland ecosystem carrying capacity exercise, in support of future planning and zoning for conservation and exclusion
- Need of a water-quality monitoring system for early warning of upcoming critical conditions and implementation of protection measures.

Biodiversity is an asset of Central Vietnam wetlands. In terms of species richness, these far outnumber any other lagoon in Vietnam and in the world. Count totals approximately 1000 species, most of them systematically identified: phytoplankton records the highest in number of representative species (287), fish's species are 215-230; birds, 73; zooplankton, 72; zoobenthos, 193 species, seaweeds 46, higher plant, 31; water grass 18 (of which 7 species of sea grass).

High biodiversity depends on physiographic complexities, diversity of habitats, and seasonal changes of the aquatic environment: salinity fluctuations between the dry and rainy season affect the composition of biota and species predominance.

## **Riassunto**

L'obiettivo di questa tesi è stato quello di creare e sistematizzare set di dati ambientali acquisiti durante quasi due decenni in siti costieri del Vietnam Centrale, (laguna di Tam Giang-Cau Hai) nelle Provincie di Thua Thien Hue e Quang Tri.

Oltre ai parametri fisico-chimici, nel database sono stati inclusi indicatori biologici di integrità dell'ecosistema, nel tentativo di sviluppare un prontuario e linee guida per rivelare il grado di efficacia delle funzioni e dei servizi ecosistemici offerti dagli ambienti lagunari e zone umide

Le pianure costiere e le zone umide offrono habitat e risorse naturali fondamentali per lo sviluppo e la sopravvivenza dell'umanità. Gli ecosistemi delle zone umide costiere offrono all' uomo risorse naturali e servizi per un valore di trilioni di dollari ogni anno e apportando un contributo fondamentale alla prosperità delle comunità rivierasche. Con l'aumento della pressione demografica e la società moderna che brama acqua e risorse alimentari, la necessità di massimizzare questi benefici non è mai stata così grande o urgente.

Nei paesi in via di sviluppo è fondamentale l'armonizzazione della conservazione ambientale e del progresso sociale. Le valutazioni del Millennium Ecosystem Assessment sono basate sul presupposto che esista una stretta relazione tra benessere e prosperità di una comunità e la sua sicurezza e resilienza: queste due condizioni sono strettamente legate alla salubrità dell'ambiente, alla sua efficacia nello svolgimento di funzioni indispensabili all'uomo e alla fornitura di servizi di supporto alla vita.

Il seguente approccio è stato seguito per istituire un protocollo di analisi ambientale

- Inventario degli ecosistemi e degli habitat.
- Sviluppo di indicatori oggettivamente verificabili e quantificabili per valutare lo stato di salute degli ecosistemi,
- Costruzione di un database relazionale ambientale a base GIS per scopi di pianificazione e gestione.
- Sviluppo di mappe tematiche utilizzando il telerilevamento.

## **Introduction**

Despite awareness and concern on environmental integrity has increased over the recent decades, wetlands are largely considered in both developing and developed countries as wasteland, unusable territory to be preferably drained and converted for development and other land uses.

Over the past century more than half the acreage of coastal wetlands and lagoons have been lost through reclamation and converted into arable land and/or urban areas. A fraction of natural wetland have undergone degradation because of human encroachment aiming to resource utilization and land occupation, such that ecosystems relying on freshwater availability are in major decline. Demographic increase and demand for water and food causes additional pressure on wetland ecosystems and as wetlands are degraded, communities struggle for subsistence, insecurity and poverty.

Since the early years of this century, Vietnam has undergone a vigorous process of economic development accompanied by aggressive land occupation and soil consumption, to accommodate infrastructures, urban expansion and industrialization. Given the geography of the country, whose population primarily concentrates on a narrow coastal stripe, embayments, coastal lagoons, wetlands and archipelagos were all to a various extent affected.

The Tam Giang-Cau Hai lagoon of Central Vietnam is a paradigm of such process of degradation, fueled by economic development, demographic increase after countryside re-population, resources overuse, deforestation and water-surface occupation by aquaculture.

Concern over the sustainability of resource use of the Tam Giang-Cau Hai lagoon raised in the late nineties when signs of depletion of fishery and loss of biodiversity became evident as a consequence of negative impact of wetland forest eradication, aquaculture development and population increase. In those years, a number of international agencies (FAO, DANIDA, World Bank, Asian Development Bank, IUCN) and a number of National cooperation agencies bilaterally intervened in support of sustainable resource use and food security in this area and elsewhere in the country.

In the years between the 2001 and 2005, the Millennium Ecosystem Assessment program was launched to appraise the consequences of the changes in the ecosystem's

structure and modifications of natural habitats affecting human well-being and to establish sound approaches and scientific methodologies for initiatives required to promote environmental conservation, sustainable use of resources and maintain the integrity of those functions that ecosystems perform ultimately and intimately contributing to societal welfare.

In the early 2000' IUCN supported the concept of Integrated Water Resource Management in the Huong River basin, whose the Tam Giang-Cau Hai lagoon is the seaward termination, in the attempt to balance 'environmental flows' in the whole catchment area subjected to diverse and contrasting needs (hydro-power, sand mining, water transportation, urbanization, agriculture, fishery and aquaculture in its downstream side. The concept of 'environmental flows' implies the application of appropriate policies and management measures to ensure that water resources are equitably shared in a catchment, in an appropriate way to maintain downstream ecosystems and services unaltered. Creating an environmental flow regime is an important foundation of Integrated Water Resources Management (IWRM).

Understanding the complexities of coastal marine and wetland ecosystems, in 2005, an FAO rural development project entered on duty in the Tam Giang-Cau Hai lagoon, applying the concept of integrated coastal zone management (ICZM) not just to water but to all resources being utilized by the riparian communities (fishery, forestry, agriculture), in the attempt to regulate their use and ensure a sustainable food supply in the long term. For the first time, the concept of ecosystem management was applied and a cross-sectorial, holistic approach was used to delineate a management system of the largest lagoon system in Southeast Asia, compatible to the multi-faceted needs of the communities of resource users.

In compliance with the spirit of Millennium Ecosystem Assessment, the development of scientific basis to evaluate the integrity of Tam Giang-Cau Hai lagoon ecosystems was piloted by the FAO project in the years 2005-2102, in the attempt to develop a relational database of environmental standardized parameters usable as a baseline by the Provincial technical departments for support to decision making.

## **Objective of this thesis**

The FAO-supported project carried out extensive monitoring in the Tam Giang-Cau Hai lagoon during the years 2006-2012. The scope of data collection and storage related to the concept of linking the ability of an ecosystem to effectively maintain its functions unaltered (and therefore to ensure services to the human communities living on it) to a set of objectively detectable and verifiable parameters, as reference values to be used in monitoring.

Given the socio-economic focus of the FAO-supported project, the science-based approach to ecosystem management remained at a piloting stage, yet providing substantial data series to be elaborated further and contributing to an assessment of ecosystem functionality.

The objective of this thesis is to systematize the environmental datasets acquired during a period of nearly two decades at set locations in Tam Giang-Cau Hai lagoon of Thua Thien Hue and Quang Tri Provinces, integrating existing secondary datasets with newly acquired primary information during 2006-2018.

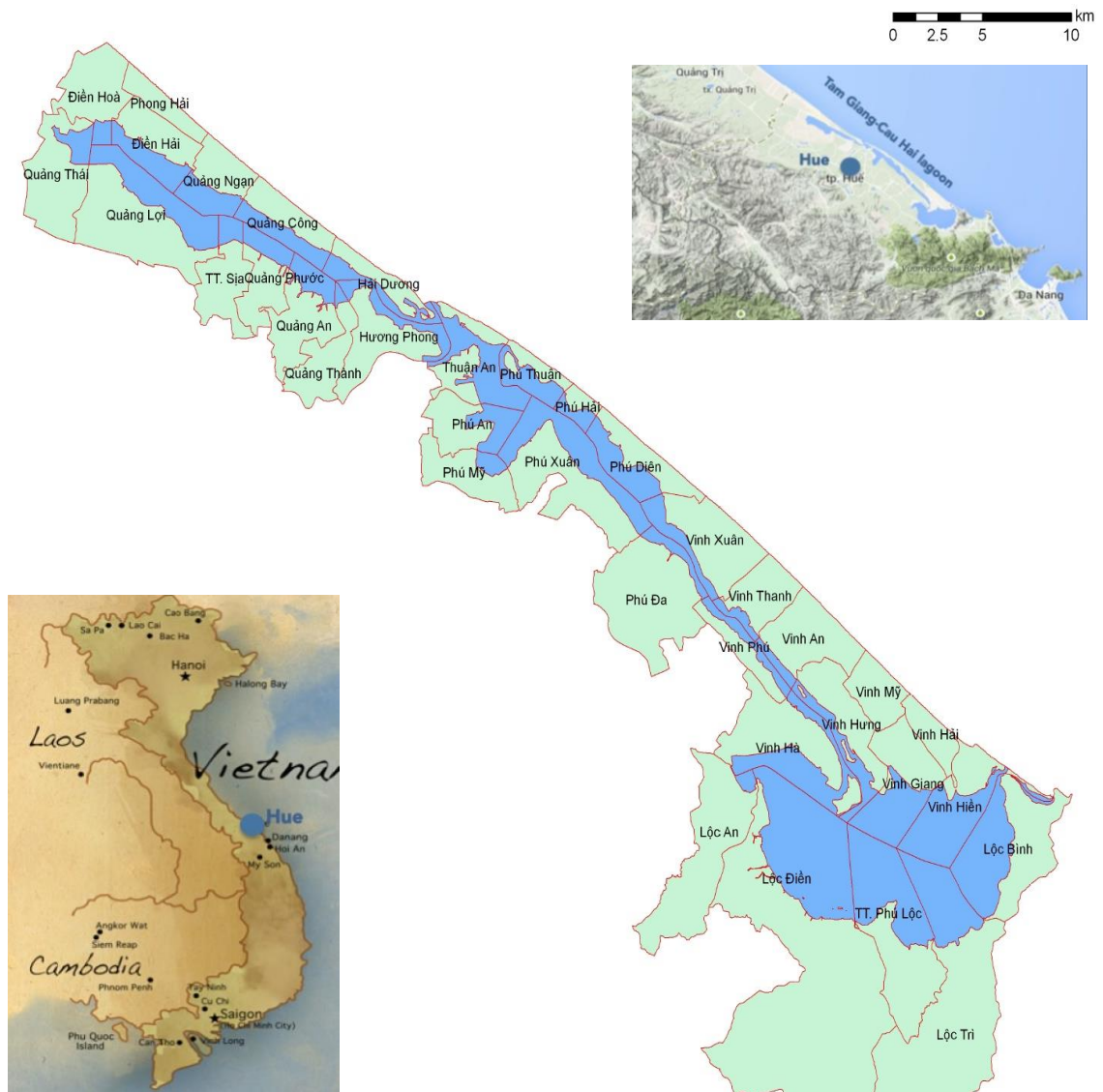
In addition to physico-chemical parameters, biological proxy indicators of ecosystem integrity were incorporated in the database, in attempt to develop a tool-box methodology to reveal the degree of effectiveness of lagoon and wetland ecosystem functions being, long-term and seasonal variations and the degree of environmental stress caused by human activities.

GIS-data spatialization will specifically locate sites/zones requiring attention, supporting a proactive approach to remediation, enhanced resilience, sustainable utilization of environmental resources and balanced development in a climate of aggressive urbanization and industrialization of the country.

### **Why wetlands matter**

Coastal plains and wetlands have been privileged sites for human civilizations for millennia. Their habitats and resources have been critical to the development and survival of humanity. Our progress in science and technological may support the misconception that nature can be ultimately controlled, but recurrent catastrophes rooted in an unsustainable use of land and water suggest that human society still depend on properly functioning ecosystems.

Wetland ecosystems are part of our natural assets, providing services worth trillions of US dollars every year and making a vital contribution to life. With increasing demographic pressure and modern society craving for water and food resources, not counting threats posed by climate changes, the need to maximize these benefits has never been greater or more urgent.



**Figure 1 – Geography and political boundaries (communes) of the Tam Giang-Cau Hai lagoon and its surrounding territory. The whole Tam Giang-Cau Hai lagoon belongs to the Thua Thien Hue Province; the 33 riparian communes belong to five districts. In the insert, the location of the Province and the Tam Giang-Cau Hai lagoon in the National context.**

Wetlands exist where superficial waters encroach over land; there mangroves forests, peatlands, salt marshes, streams, ponds and lakes, but also deltaic expanses, floodplains and flooded forests develop. Coastal wetlands occur where rivers meet the sea, water flow slows and expands into lagoons, embayments and shallow waters. Agriculture is practiced extensively in coastal wetlands, such as rice paddy fields and orchards, to a various extent altering the natural landscape.

Rivers convey fertile sediments to the sea, building-up floodplains that support agriculture. Rivers and estuaries are waterways facilitating maritime commerce and transportation; rivers are source of energy and drinking water for vast regions on Earth. River-related wetlands provide critical habitat for fishes and other aquatic animals vital for humans as food and proteins.

Deltaic swamps, lagoons and estuaries control and regulate water supply and quality to the most densely populated areas of the planet. Water risks to society, including scarcity, droughts and floods, are mitigated at wetlands, whose conservation and restoration is a cost-effective investment towards water security.

Wetland (mangrove) forests thrive in tropical and sub-tropical wetlands. These complex ecosystems develop under saline conditions unsuitable to most forest formations. Besides being nursery to a vast range of aquatic creatures, providing nutrients to nourish plankton, algae and fishes, mangrove forests offer physical protection against climatic extremes such as typhoons, storm winds and related floods. Dense mangrove forests may absorb and abate the destructive force of storm surge being an effective mitigation measure against weather deterioration due to climate changes. Moreover, mangroves are among the most effective carbon storage systems on Earth.

## **Background**

A fundamental issue in developing countries is to harmonize environmental conservation and social progress. Human conditions are tightly linked to the state of the environment where economic activities and social life are carried out. The Millennium Ecosystem Assessment stemmed from the assumption that a close relationship between welfare and prosperity of a community, its security and resilience is related in a multi-faceted way to the salubrity of the environment and its effectiveness in performing functions and providing services supportive to human life.

Since the early 2000', the concept of ecosystem-based management was reflected into policy papers, management regulations and plans, in such a way that the focus was shifted from human actions and economic activities solely, to broader spectrums of initiatives promoting conservation and rehabilitation of ecosystems as a whole.

The concept of ecosystem management pioneered by the FAO developmental project in Tam Giang-Cau Hai lagoon designed its implementation strategy to achieve both ecological and social progress, by blending environmental science, monitoring and community development, without detracting from its primary objective that was food security and poverty alleviation.

An analysis of World Bank projects with objectives of alleviating poverty and protecting biodiversity revealed that only 16% made major progress on both objectives.

The approach developed in this thesis relates to the delivery of a framework in which a detailed exploration of science data and social economic achievements in a specific area are conducive to balance biodiversity conservation and social progress as a result of how people manage their ecosystem services.

Scientific advances around ecosystem functions and ecosystem service production and the design of appropriate monitoring programs are pivotal for the implementation of conservation and developmental objectives that will successfully advance both environmental and social goals.

In the early days, the Millennium Ecosystem Assessment endeavor exposed huge gaps in what later became known as science of ecosystem services. Yet, many development projects of those days embodied both aspects, recognized as intimately related by both conservationists and developmental agencies, of stewardship of nature, biodiversity conservation and economic development.

The results were a series of developmental initiatives, especially in the Tam Giang-Cau Hai lagoon, with integrated conservation and development purpose, all justified by the assumption that nature provides human communities with benefits and poverty and environment degradation and the "two sides of the same coin".

Understanding the constraints that influence the outcomes of projects with dual conservation and economic development goals is crucial. One possible approach is to elaborate an investigation framework to be applied for assessing the connections



between ecosystem services and economic development, by adopting indicators and metrics that could increase the likelihood of concurrent positive outcomes (win-win) in both interconnected aspects of natural and social life.

### **A framework linking environmental science to human welfare.**

There are two possible approaches by which the science of ecosystem services can contribute to biodiversity and environmental integrity as well as to social welfare, economic progress and poverty alleviation.

One strategy is to promote economic viability of activities deriving from the effectiveness of services delivered by a specific ecosystem, in such a way that these activities may generate additional income and, in the best case, promote investments whose profits could be utilized to compensate eventual loss of economic opportunities suffered by communities engaged in environmental conservation.

In this case, the full accounting of ecosystem services available and a better insight on how and rater at which theses services are delivered, and economic value attached to them, might be an incentive to pay for nature conservation.

The second approach relates to the internalization of benefits deriving from the ecosystem management, by fostering a community-based economy through promotion of rural industries and access to the market of those goods (e.g. organic produce) and resources (e.g. touristic offer) whose availability is enhanced by the adoption of better environmental conservation practices.

When the economic value of natural ecosystems and resources are explicitly quantified, benefits are more valued both by the people who directly interact with the ecosystems and/or (government) agencies that would have to financially contribute from substitute sources, should ecosystems become impaired.

Whichever strategy is adopted, the following investigation steps should be undertaken to set up a framework for the implementation of developmental projects or the drafting of ecosystem-based management policies and plans aiming at both environmental conservation and human well-being:

- Inventory of ecosystems and habitats in a specific locale

- Development of a set of objectively verifiable and quantifiable indicators to appraise the state of ecosystems, their effectiveness in performing functions and delivering services.
- Establish a science-based framework for environmental monitoring and data acquisition for use of scientists and policy makers
- Establish a GIS-based environmental relational database to serve the purpose of planning, management and policy drafting.
- Develop thematic mapping for planning, tracking environmental changes, land use, soil consumption, illegality and abusive encroachment of urban settlements onto wetland.
- Use remote-sensing techniques to extrapolated point-source information to land and water surfaces

A methodology for each of these components is delivered as output of this thesis, then integrated into an operational tool-box for use of managers and decision makers.

Practical applications of the methodology and use of the toolbox is to adopt a scientific approach based on objectively verifiable indicators to ecosystem assessments and effectiveness of ecosystem functions and services supporting human life and community's well-being.

### **Ecosystem functions and services**

Ecosystem functions are natural process that take place in animal and plant communities of the different biomes through exchanges of energy and nutrients in the food chain which are vital to the sustenance of plant and animal life on the planet.

Ecosystem services are ecosystem functions that are directly beneficial to humans.

These concepts evolved from the vast literature flourished on the subject, dating back since the sixties and seventies (King, 1966; Neef, 1966; Van Der Maarel and Dauvillier, 1978) and exponentially grown in the nineties (De Groot, 2002) detailing with increasing precision the roles natural ecosystems have in supporting human life and improving capacity to cope with adverse (natural, economic, societal) events (i.e. “resilience” in modern terms).

The “ecosystem service” concept has evolved through time to attain its modern significance of a multi-layered approach at the interface between ecosystems and human well-being.

Daily (1997) defines ecosystem services as “the conditions and processes through which natural ecosystems, and the species that make them up sustain and fulfill human life”. Costanza et al., 1997 and Millennium Ecosystem Assessment (2005) provide further predications as “**benefits** human populations derives, directly or indirectly from ecosystem functions.

Boyd and Banzhaf (2007) put emphasis of “consumption or utilization” of ecological components (*i.e.* resources, tangible or intangible such as landscape) for the purpose of achieving ever increasing human well-being. TEEB (2009) and Fisher *et al.*, (2009) follow the same line of reasoning, whilst U.S. EPA (2008) focuses on the concept of sustainability defining ES as “products of ecological functions or processes that directly or indirectly contribute to human well-being, or have the potential to do so in the future”.

Ecosystem services are susceptible to human abuse when natural processes are impaired or excessive resources are withdrawn to sustain a growing population. Therefore, the nature of these ecosystem services and how these are sustained should be understood.

### **Classification of ecosystem functions and services**

The Millennium Ecosystem Assessment (MEA) document provides a framework for understanding and assessing the benefits that ecosystems deliver to communities. According to it, services are grouped into four categories:

- Supporting services. These include basic functions, such as nutrient recycling, primary production or soil formation, not directly beneficial to humans but indispensable to support ecosystem effectiveness and capacity to deliver the services of the other categories. TEEB replaced "supporting services" of the MEA document with "habitat services" defined as "ecosystem functions ... a subset of the interactions between ecosystem structure and processes that underpin the capacity of an ecosystem to provide goods and services"
- Provisioning services relate to the production of commodities usable by humans as food, raw materials (e.g. lumber, fuel, etc.), genetic resources, water and energy, but

also biogenic minerals, medicinal and ornamental resources.

- Regulating services relate to the role ecosystems play in altering/regulating natural and biological processes, or even planetary cycles such as climate, carbon cycle etc. Pollination is a basic regulating service, but also carbon sequestration, crucial in climate regulation, flood control, waste decomposition and detoxification, purification of water and air, pest and disease control.
- Cultural services pertain to the spheres of aesthetics, spiritual life, history, support to education (school excursions) and recreational activities, such as ecotourism and sports).

### **The pilot study area and project justification.**

The Tam Giang-Cau Hai lagoon with its 22,000 hectares of water surface and associated wetlands, deltas and forests, is the largest lagoon system in Southeast Asia. It is a key area that, as a consequence of post-war rural repopulation, demographic increase, resource overuse, urbanization and industrialization, has undergone substantial changes in the past two decades.

As a consequence of human pressure and encroachment of the natural habitats, there is a perception that ecosystem functions and services have been impaired; however, a clear and science-grounded understanding to what extent the beneficial effects to the riparian communities has been compromised.

Initiatives by international organizations (FAO, IUCN), NGOs, bilateral donors and the GoV have been promoted with the objectives of reverting trends of environmental degradation, primarily with a focus to poverty eradication, social and economic development but without a strategy and scientific information on how to restoring ecosystem functionality and protect ecosystem-service delivery to humans.

Environmental data have been collected extensively by governmental agencies and academic/research institutions, but databases hardly used for developing appropriate policies and undertaking management measures in support of environmental defense and resource conservation.

## **Objectives of the study**

This doctoral thesis has been conceived with the objectives of developing a methodology for a science-based analysis of environmental effectiveness and ecosystem-service provision based on:

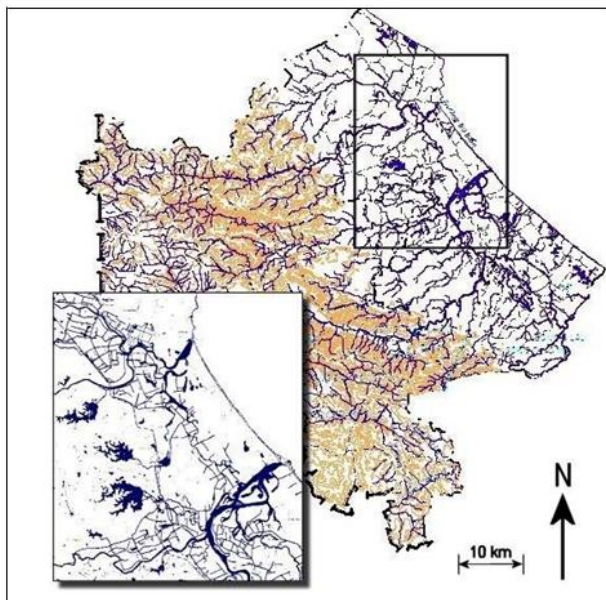
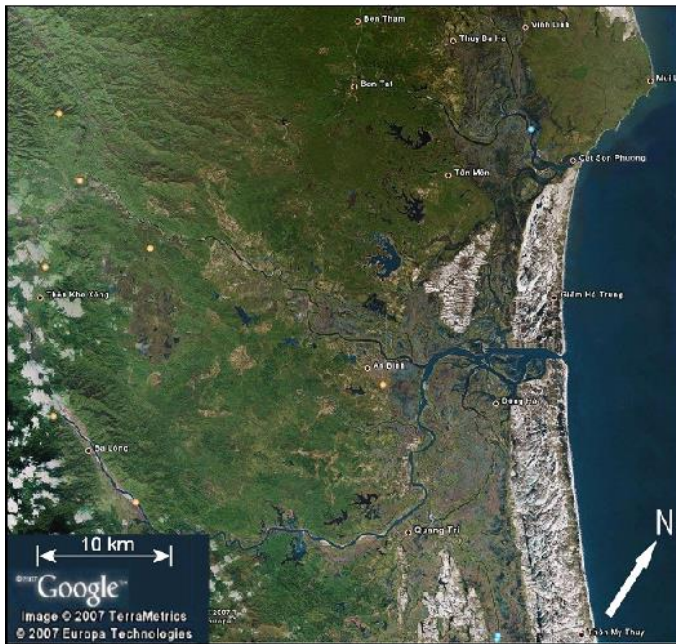
- available and routinely acquired data,
- standardization, spatialization and consistency assessment of available information,
- ecosystem and habitat identification,
- assessment of resource state and trends
- tool development for data dissemination

## **Regional background and critical issues**

The coastal zone of the Thua Thien Hue Province makes up 34% of its total area and about 81% of its population; it is where most economic activities are concentrated.

The Tam Giang-Cau Hai lagoon, with its 68 km in length and 22,000 hectares of water surface is the largest lagoon in South East Asia. A community of over 300,000 people, mainly farmers and fishers, live on the resources of the water basin and its surrounding land. Administratively, thirty-three communes face the waters of the lagoon, grouped into five lagoon coastal districts, all under the Provincial authority of Thua Thien Hue.

North of Thua Thien Hue lays Quang Tri Province, the natural continuation of this Central Vietnam wetland, lagoon and estuary complex spanning from Phu Yen to Quang Binh.



**Figure 2: Location of Quang Tri Province and related river network**

The Quang Tri province is located in a strategic area for transport and communication, at the junctions among the National Route 1A, the Ho Chi Minh Highway and the East-West Economic Corridor. Also the North-South Expressway and Coastal Highway. Located at the border-gate of Vietnam, Quang Tri is the focal node of the Indian Ocean and South China commercial traffic, terminal of the East-West Economic Corridor and the Mekong sub-region.

Quang Tri province has a total area of 4,592 km<sup>2</sup> and a large river network that provides a suitable setting for aquaculture. The Quang Tri province also has 55% area of the

mountainous territory and a 75-km-long coastline with a number of touristic destinations (Cửa Tùng, Cửa Việt, and Mỹ Thủy).

There are two major rivers: the Ben Hai river to north and the Thach Han river to the south, flowing into their own estuaries in Cua Tung and Cua Viet. The estuarine system in Quang Tri is complex because of the large number of tributaries draining the hinterland. In the estuaries, the salt water from the sea can invade up to 24 km inland creating a broad brackish zone of embayments, ox-bow lakes and wetlands.

The coastal belt is occupied by coastal dwellers; associated waterways and wetlands are being used for shrimp and fish farming (Trinh Q.T., 2010). Many local farmers shifted to shrimp culture because of high economic income from some successful models, but unplanned development of this industry brought up excessive pollution from untreated effluents, environmental deterioration and the prospect of unsustainable resource use.

There is a clear need of environmental management for this extensive wetland and lagoon system and a latent need for management of the coastal and inshore marine fisheries as a whole. According to existing data and feedback from fishers, commune leaders and local knowledge, the main reasons for the need to manage the coastal zone in an integrated manner are:

- The persisting degradation of the lagoon environment and ecosystems, because of overfishing, fish farming and deforestation.
- A significant decrease in stock abundance and diversity caused by illegal, unregulated and unreported (IUU) fishing and lack of monitoring, surveillance and control (MSC).
- A natural tendency to inlet siltation because of insufficient tidal draft.
- The need to include communities in management.
- The need to harmonize processes of urban expansion, water-surface and agricultural land tenure, industrialization and rural development.

The objective of this study is to provide decision makers scientific tools to revert the negative trends of the past decades that turned the lagoon into a livestock-depleted freshwater lake, polluted by untreated effluents, with most of its wetland habitats and wildlife endangered or vanished.

Two pilot area were selected in this stretch of Central Vietnam coastal zone: the Quang Tri estuaries and the Thua Thien Hue Tam Giang-Cau Hai lagoon Giang-Cau lagoon.

The environmental degradation of the Tam Giang-Cau Hai lagoon and increasing pressure on its aquatic resources, together with typhoons, floods and droughts that yearly strike the province concurred in creating a situation of particular vulnerability for people living on and around it. This situation of persistent threat and creeping decline has been a matter of concern in the past two decades: the application of the methods of science in support of environmental monitoring and control will contribute to a more effective management of the locale in the long term.

### **Presentation of data**

Original datasets were collected, in the years 2015-2018, according to a protocol, laboratory procedures and sampling grid compatible with previous campaigns implemented in 2006-2013 whose data are available from the local literature and internal Provincial reports.

Only selected datasets are discussed in the text, but the whole archive of information considered in this synthesis is given as Annex 1.

A substantial component of the study was dedicated to benthic foraminifera, specific taxonomic category considered of importance as indicators of environmental quality and stress.

### **Materials and methods**

#### Methods of water and sediment samples collection

Water samples were collected using a bathometer at 3 depths: surface, mid-water and bottom layer. Sediment samples were collected using a Van Veen grab, wrapped in zinc foil. The water and sediment samples were stored at  $< 4^{\circ}\text{C}$  in the laboratory, before being analyzed.

#### Methods of phytoplankton sample collection

Phytoplankton samples were collected using a phytoplankton net with a 15 cm diameter and 45 cm length, and a mesh with a dimension of 20  $\mu\text{m}$ , in an area of 15-m radius at each station. Upon collection, phytoplankton samples were fixed with formol 4%,



transported to the laboratory and preserved at  $< 4^{\circ}\text{C}$ . Laboratory identification relied on morphological comparison method. Samples were analyzed with Olympus CH40 microscope with contrast phase.

#### Methods of sediment sample collection for foraminiferal studies

Samples were collected from the Tam Giang – Cau Hai lagoon in Thua Thien Hue Province and Cua Tung, Cua Viet estuaries in Quang Tri Province. Sediment and water sampling were collected using a Van Veen grab and Van Dorn sampler, respectively.



**Figure 3 - Sampling stations in Quang Tri Province, April 2017.**



**Figure 4 - Van Veen grab (left), Van Dorn sampler (center) and 50-mL-capacity Falcon conical tubes (right).**

Sediment and water samples from each location were stored together in 50-mL-capacity BD Falcon Conical Tubes and transferred to the laboratory at the end of the day for storage in refrigerated chambers.

The positions of the sampling stations were determined and recorded as WGS84 latitude and longitude, using the Global Position System.

#### Water quality analyses

In order to assess water quality, the following list of parameters were measured, both directly in the field during sample collection and in the laboratory.

- Water depth of station,
- Turbidity,
- Temperature,
- Flow velocity during tidal flood and ebb,
- Compositional and textural parameters of bottom sediments,
- Total solid and suspended solid present in the water,
- pH,
- Salinity,
- Dissolved oxygen,
- Nitrites and nitrates,
- Ammonia,
- Alkalinity,
- Total phosphorous,

- Total nitrogen,
- Chlorophyll A
- Zoobenthos biomass
- Phytoplankton
- Microalgae and toxic algae
- Total coliforms, E. coli and Vibrio cells
- Measuring methods are listed in the following Table 1.
- 

Type of analysis	Measuring method
Sediment	Visual description and grain size analysis
Depth	Measuring stick and ruler
Turbidity	Secchi disk
Temperature	Digital thermometer
Dissolved Oxygen (DO)	Sera test kit
pH	Sera test ki
kH	Sera test kit
NH <sub>4</sub> <sup>+</sup> /NH <sub>3</sub>	Sera test kit
NO <sub>3</sub>	Sera test kit
NO <sub>2</sub>	Sera test kit
Salinity	Optical salinometer
Total Solid (TS)	Mass balance method (dried at 103–105°C)
Suspended solids (SS)	Mass balance method (dried at 105°C).
Chlorophyll-a	Spectrophotometric determination method (SMEWW 10200 H)
Detection of E. coli and	TCVN 6187-1996 (Standard method ISO 9308 – 1 – 1990) – filter membrane method.

**Table 1 – Parameter measuring and detection methods**

### Grain-size analyses

Grain-size analyses were performed using the principle of diffraction. Three grain-size fractions were considered according to Wentworth, C (1922): silts ( $<63\ \mu\text{m}$ ), very fine sands (63 to  $125\ \mu\text{m}$ ) and sands ( $>125\ \mu\text{m}$ ). After being dried at  $40^\circ\text{C}$ , the sediment samples were weighed on a high-precision scale, then sieved through  $125\text{-}\mu\text{m}$ -mesh and  $63\text{-}\mu\text{m}$ -mesh sieves and weighed again separately.

### Microalgae and phytoplankton analyses

Qualitative algae determinations were based on morphology following classification by Ngoc An Truong (1993), Larsen and Ngoc Lam Nguyen (2004), and Tomas (1997) used in light microscopy. Quantitative counts were made using Sedgewick Rafter chamber (volume 1ml) under a microscope with a magnification of 100x.

### Foraminifera analyses

Samples were analyzed for foraminiferal content following the technique described by Walton, W.R., (1952) and Lutze, G.F., and Altenbach, A., (1991). After sampling, sediments were preserved in the laboratory adding 10% concentrated formalin solution buffered with sodium borate along with Rose Bengal stain (1g/L) for a minimum time of 14 days for discriminating living specimens and dead tests, following the FOBIMO protocol (Schönfeld, J., *et al.*, 2012).

Samples were washed through sieves  $63\text{-}\mu\text{m}$ -mesh sieves, then dried at  $40^\circ\text{C}$ , weighed on high-precision scale and sieved again with  $125\text{-}\mu\text{m}$ -mesh and  $63\text{-}\mu\text{m}$ -mesh sieves, corresponding to the coarser and finer grain-size limits of fine sand. Each residual sample was then weighed separately and the results expressed as percentage of each size fraction against the total weight of the sample.

The counted living (stained) benthic foraminifera were hand-picked by dry picking and stored in micropaleontological slides.

For species identification, the World Register of Marine Species (WORMS, <http://www.marinespecies.org/>) classification was used.

Species evenness, richness, and diversity indexes as the Shannon-Weiner (Shannon C. E., and Weaver, W., 1949) and Simpson Index (Simpson, 1949) were used to evaluate

foraminiferal species diversity, in order to compare these data to the environmental standards of estuaries and coastal wetlands.

### **Results of analyses**

The results of analysis of sediment and water samples collected during surveys in both Tam Giang-Cau Hai lagoon, Cua Tung and Cua Viet estuaries are herewith presented followed by a discussion and conclusions on ecosystem identification and mapping.

The focus of the thesis is hinged on the assumption/observation that benthic foraminiferal assemblages display an overly sensitivity to environmental changes. The rationale of the investigation is therefore based on a threefold approach:

- Comparative analyses of historical time series of environmental data over the period of a decade.
- Relationships between environment parameter changes and shifts in the composition of foraminiferal assemblages.
- Calibration of foraminifera assemblages as a predictive tool for upcoming environmental changes.

### Foraminifera biology and ecology: proxies for environmental health

Foraminifera are single celled protozoans that consist of successive chambers and widely distributed in marine environment from the intertidal zone to the deep ocean floor, range from the polar region to the Equator, in marginal to deep basin. In the fifth century B.C., Herodotus reported that large benthic foraminifera of the genus *Nummulites* are abundant in the limestone used to build the Egyptian pyramids at Gizah.

In the 16<sup>th</sup> and 17<sup>th</sup> centuries, smaller foraminifera were for the first time described, illustrated, and variously classified. The French naturalist Alcide d'Orbigny (1826) revolutionized the foraminifers' taxonomy describing them as tiny cephalopods. Felix Dujardin (1835) first determined their true protozoan nature and promoted their recognition as a separate Class (d'Orbigny, 1839).

The famous world cruise of H.M.S Challenger in 1880's collected many samples and Henry Bowman Brady (1884) illustrated the foraminifera dredged from the sea floor.

Throughout the 20<sup>th</sup> century, there were several prominent scientists working on in foraminiferal studies. Joseph Augustine Cushman and his team published 556 papers

and particularly the first journal on Contributions from the Cushman Laboratory for foraminiferal research (Cushman, 1927) and one of the most influential texts in the field is: "Foraminifera: Their Classification and Economic Use" (Cushman J.A., 1928). Oil industry, after World War I, drove an impulse on the development of benthic foraminiferal biostratigraphy. In the decade prior to World War II, the Paleogene zonation was developed for the Caucasus Mountains of the southwest Soviet Union (Subbotina, 1953) and the Caribbean (Bolli, H., 1957a, 1957b, 1957c, 1966).

Furthermore, the postwar search for petroleum in Trinidad in the southernmost Caribbean region led to a revolution in biostratigraphy. The standard provided by Hans Bolli, who published Cretaceous and Tertiary zonation (Bolli, H. M., 1957a, 1957b, 1957c, 1966), was applied and expanded in the later biostratigraphic research (Blow, W.H., 1969, 1979; Berggren, W. A., 1969; Stainforth, R. M., *et al.* 1975).

Beside the development of foraminiferal biostratigraphy, the micropaleontological techniques expanded, to be applied to the ocean and deep-sea cores (Schott, D. W., 1935; Cushman, J. A., & Henbest, L.G., 1940; Emiliani, C., & Edwards, G., 1953; Ericson, D. B., & Wollin, G., 1956). The Deep Sea Drilling Project (1968 - 1983) contributed successfully to the expansion of the micropalaeontological investigations on deep-sea. Further studies on living foraminifera have contributed to the understanding of their fossil counterparts (Reiss, Z., & Hottinger, L., 1984, Hemleben, C. *et al.* 1989) and through foraminiferal ecology, this groups has become a tool for environmental analysis, providing bio-indicators for environmental stress and impacts from natural and anthropic hazards (Jorissen, F. J., 1988; Hottinger, L., & Pecheux, J. F., 1991).

According to the World Register of Marine Species (2018), at present there are 8,983 recognized recent species and 34,766 recognized fossil species recorded.

The classifications of foraminifera are based on the characteristics of the shell or test, including morphology, apertural structure, shell components, and types of chamber arrangements.

The tests of foraminiferal skeleton is secreted or agglutinated shell and consists of one or more chambers. The chambers are separated by septa, but with a continuous connection through a foramen in each subsequent septum. Through the opening on the last chamber, the protoplasm extends outside the test and the pseudopodia extrude to capture prey and aid in movement. The chambers may arrange as a single row

(uniserial), a double row (biserial), or a triple row (triserial). Some tests coil in a single plane (planispiral) or coil up in a spire like the shell of a snail (trochospiral). The emergent part in center of the spire is called umbo and the indentation part in the other side is called umbilicus. If the earlier chambers enveloped by the later ones, the test is called involute; if they are visible, the test is termed evolute. The external line of junction between the chambers is known as suture.

The organelles and the cytoplasm of foraminifera include: the Golgi bodied, ribosomes, vacuoles, microbodies, microtubules, xanthosomes and endoplasmic reticulum. The protoplasm consists of small amounts of organic pigments, iron compounds, brown and red deposits of fatty material, brown excremental particles and green splotches (Boersma, A., 1998).

Foraminifera are roughly divided into two major groups: planktonic and benthic. The planktonic species float passively, moved only by currents but capable of vertical migration. Their assemblages provide information about the conditions in surface water where they live. The benthic species live on the sea floor, either at the surface or buried in the sediment, or are attached to plant stems, rocks or particles. Benthic foraminifers constitute the most diverse group, small compared to the other shelled micro-organisms in modern seas (Gupta, S., 1999) and play an important role in the economy and balance of the biosphere (Cimerman, F. and Langer, M.R., 1991).

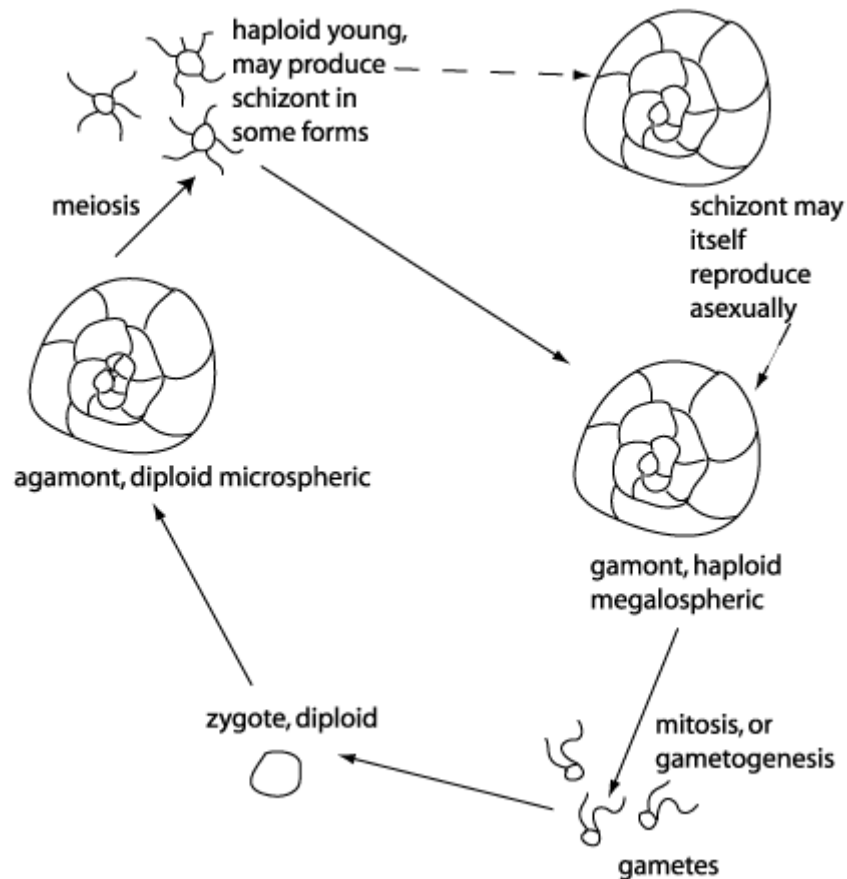
From the 1960s, benthic foraminifera were used to describe the state of the environments in which they live or lived (Resig, J.M., 1960, Watkins, J.G. 1961 and Boltovskoy, E., 1965) and the methodology has increasingly developed until the most recent decades. Because of their short life cycles (Boltovskoy, E., 1965, and Murray, J.W., 1991 and 2006), high biodiversity and specific ecological requirements, benthic foraminifera are particular sensitive to respond with environmental changes and can be successfully used as bio-indicators of environmental status in a wide range of marine and marginal marine environments (Alve, E., 1991 and 1995; Hayward, B.W., 1993; Frontalini, F., *et al.*, 2009; Romano, E., *et al.*, 2015). In coastal and estuarine environments, under natural conditions, foraminifera tend to reveal the relative inflow of salt water with fresh water, which affects salinity (Nichols, M. M., 1974 and Odum, H.T., 1974) and other variables.

The foraminiferal life cycle is termed heterophasic, which is characterized by the alternation of sexual and asexual generations (Fig. 5). The asexual phase, *schizogony*, produce the gamonts have a larger initial chamber, the *proloculus*, and a smaller test size termed the macrospheric generation. Meanwhile, the sexually-produced, *gamogony*, a gamont, which is characterized by a smaller proloculus and an overall larger test size, called the microspheric generation (Goldstein, S. T., 1999). Repetitive asexual reproduction, also accomplished through schizogony (Fig. 5), seems to be the main reproductive mechanism for benthic species (Alve, E., and Goldstein, S. T., 2003).

Benthic foraminifera are generally taxonomically classified on the basis of morphological parameters and primarily discerned on the basis of test chemistry. The most important groups include species with calcareous perforate tests, the Miliolids which have a calcareous imperforate test, agglutinated species and organic-walled (or soft-walled) ones. This distinction is particularly important also for the study of biological processes and their linkage to bio-mineralization, because the involved processes change on the basis of the test structure.

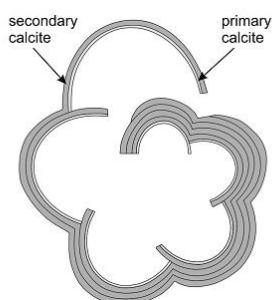
The most studied models for bio-mineralization relate to the calcareous perforate species, generally from the order Rotaliida. The calcite test consists of a series of chambers that are constructed sequentially. In life, the protoplasm of the cell pervades the chambers and extrudes to the exterior of the test, functioning in excretion, food capture, and chamber construction. Assembly of a new chamber wall begins with the protoplasm extending outward to define an





**Figure 5 – Diagram showing a generalized foraminifer life cycle. Note the alternation between a haploid megalospheric form and a diploid microspheric form (redrawn after Goldstein, S.T., 1999).**

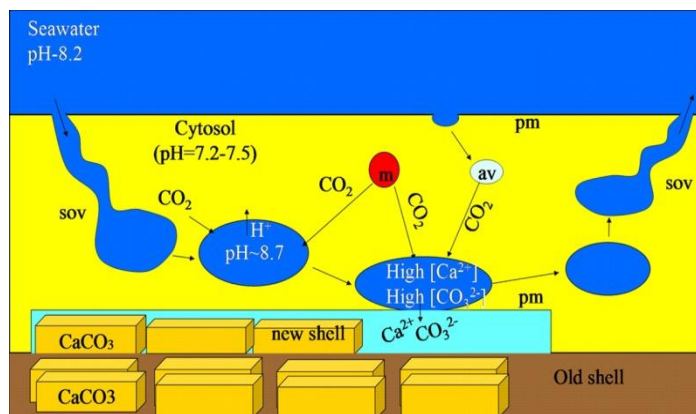
outline that extends beyond the future positioning of the chamber being constructed (Cusak, M., and Freer, A., 2008). Calcification then takes place within this defined space with the formation of an organic layer or membrane that forms the shape of the new chamber. Nucleation occurs on both sides of this organic template, forming a new layer of primary calcite in the form of a new chamber wall that is then overlain by another layer of (secondary) calcite that extends over the entire test (Figure 3). Thus, the perforate calcite tests become increasingly laminated (Cusak, M., and Freer, A., 2008).



**Figure 6 - Formation of new chamber and laminate structure in perforate foraminifera. (Erez, J., 2003, reprinted by Cusak, M., and Freer, A., 2008).**

The mechanism of bio-mineralization of Miliolids and thus the final structure of the test, is different. In this case calcite needles and associated organic matter are preformed within the cytoplasm and transported to the forming chamber. Then, calcification occurs as cytoplasm is being extruded from the last complete chamber, proceeding from the base of the new chamber to the aperture; that is, the extruded cytoplasm does not first form the shape of new chamber and then the wall as in the previous case (Goldstein, S.T, 1999). Therefore, Miliolid tests do not have an inner organic layer as for lamellar tests of the calcareous perforated specimens.

Bio-mineralization is the ability to precipitate minerals from soft tissue, calcite in the case of foraminifera: some recent studies have deepened the understanding of mechanisms at the basis of these processes.  $\text{CaCO}_3$  precipitation, both in calcareous species and Miliolids, occurs after the incorporation of seawater in the cytoplasmic vacuoles (as illustrated in Figure 4, after Bentov, S., *et al.*, 2009). In these vacuoles, precipitation is enhanced by local increasing of pH (up to  $>9$ ), as described by de Nooijer, L.J., *et al.* 2008.



**Figure 7 - Model for temporal fusion of the seawater vacuoles (SWV) with the plasma membrane (pm) near the active calcification site. Seawater enters the cell through deep invaginations or semi-open vacuoles (sov). A vacuole is pinched off and undergoes alkalization by one of the cellular**

**proton transport mechanisms. This SWV concentrates inorganic carbon by diffusion of  $\text{CO}_2(\text{aq})$  from the acidic cytosol into the alkaline SWV. This process is enhanced by adjacent mitochondria (m) and by the acidic vesicles (av) that release  $\text{CO}_2$ . The  $[\text{Ca}^{2+}]$  and  $[\text{CO}_3^{2-}]$  enriched vacuoles fuse with the cell membrane and supply the ions for calcification. The vacuoles are then resealed and release their content apically, away from the growing crystals (after Bentov, S., *et al.* 2009).**

Concerning agglutinated species, the mechanism of test formation is totally different. In fact, generally these foraminifera gather sediment grains into a mound near their aperture. Once cavity is formed within this mound of sediment, the foraminifer begins constructing a chamber by coating grains with a thin organic envelope, and then cementing them together with a small amount of organic cement. Some agglutinated

species (e.g. *Textularia* species) are also able to precipitate a calcitic cement, that further binds the agglutinated materials together (Goldstein, S.T., 1999).

Species/Stations	QT1	QT2	QT3	QT4	QT5	QT6	QT7
<i>Ammonia sp.</i>	0	0	0	0	2	6	5
<i>Dentalina sp.</i>	0	0	0	0	0	1	0
<i>Elphidium sp.</i>	0	0	0	0	8	20	0
<i>Fissurina sp.</i>	0	0	0	0	0	1	0
<i>Nonionella sp.</i>	0	0	0	0	0	2	0
<i>Planulina sp.</i>	0	0	0	0	0	2	0
<i>Reussella sp.</i>	0	0	0	0	0	3	0
<i>Sigmoilina sp.</i>	0	0	0	0	0	4	0
<i>Miliammina sp.</i>	7	32	2	3	0	0	0
<i>Miliolinella sp.</i>	0	0	0	0	1	0	0
<i>Quinqueloculina sp.</i>	0	0	0	0	28	15	0
<i>Spiroloculina sp.</i>	0	0	0	0	1	0	0
<i>Triloculina sp.</i>	0	0	0	0	5	2	0
<i>Ammobaculites sp.</i>	18	215	62	31	0	0	0
<i>Haplophragmoides sp.</i>	0	0	3	0	0	0	0
<i>Psammophaga sp.</i>	0	8	0	0	0	0	0
<i>Psammosphaera sp.</i>	0	0	9	1	0	0	0
<i>Trochammina sp.</i>	5	0	10	8	0	0	0
<b>Total</b>	<b>30</b>	<b>255</b>	<b>86</b>	<b>43</b>	<b>45</b>	<b>56</b>	<b>5</b>

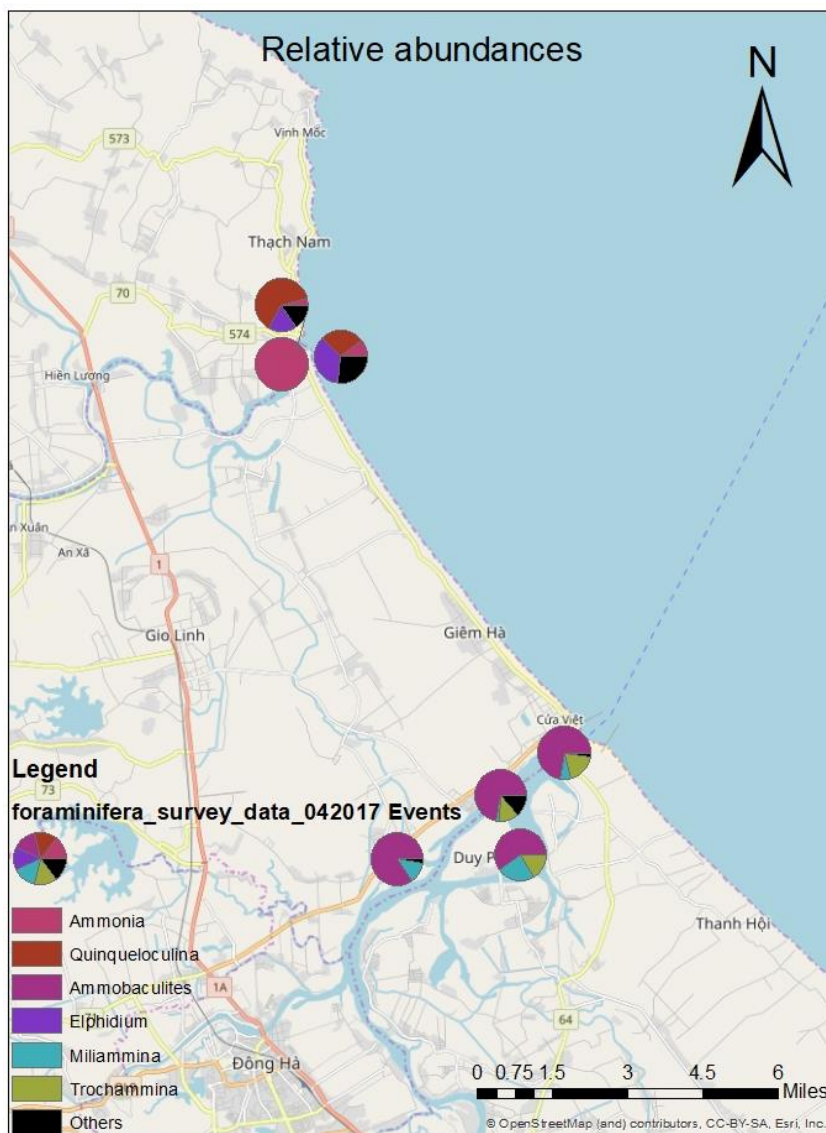
Table 2 - Foraminifera species abundance each station in Cua Viet, Cua Tung estuaries, Quang Tri Province - Survey April 2017

At last, soft-shelled foraminifera have generally an organic wall, without calcite. This group includes simple organic shelled species and micro-agglutinated species that are able to weakly incorporate fine sediment particles in the organic matrix (Gooday, A.J., 2002).

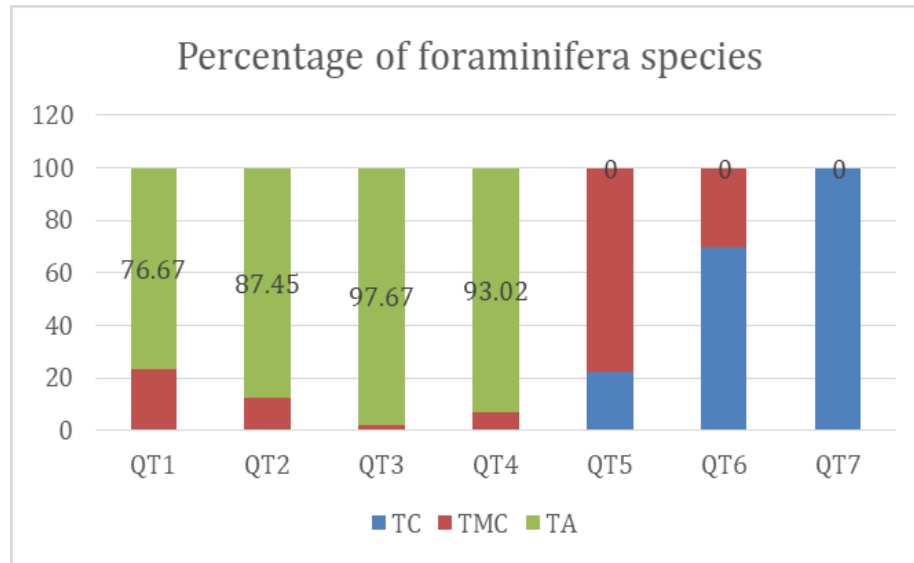
Analyses of foraminiferal assemblages from Cua Tung and Cua Viet estuaries, April 2017

Table 2 show that different locations have different foraminiferal assemblages. QT2 has the highest foraminiferal abundance with 255 individuals; QT6 is the station that has the highest species diversity. In QT7, only *Ammonia tepida*, a species which normally is found in brackish water, occurs in the studied sediment sample.

The assemblages are dominated by *Ammobaculites* sp. in Cua Viet area (QT1, QT2, QT3, and QT4) and by *Elphidium* sp. and *Ammonia* sp. in Cua Tung area (QT5, QT6, and QT7) (Figure 7). Subordinate species include *Dentalina* sp., *Fissurina* sp., *Miliolinella* sp., and *Spiroloculina* sp. with only 1 individual for each specie (Table 1). From inlet along to river branches in Cua Viet estuary, the percentage of *Ammobaculites* sp. increase compare to the total taxa composition in each station.



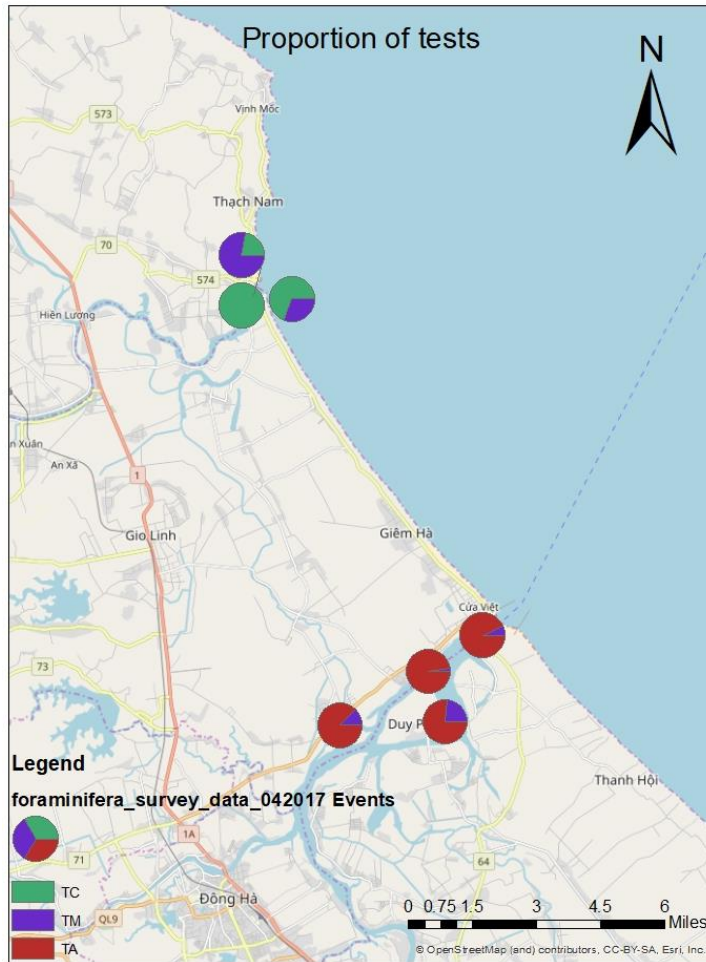
**Figure 8 - Relative abundances of the main living benthic foraminifera genus in Cua Tung and Cua Viet estuaries, Quang Tri Province, April 2017**



**Figure 9 - Percentage of foraminifera species in Cua Tung (QT5, QT6, QT7) and Cua Viet (QT1, QT2, QT3, QT4) estuaries, Quang Tri Province. Survey April 2017. TC, total calcareous; TMC, total Miliolids calcareous; TA, total agglutinated**

Taxa composition, as percentage of each foraminiferal group (calcareous perforated, miliolids calcareous, agglutinated) is shown in Figure 8. Comparison among samples from Cua Tung and Cua Viet estuaries, shows that agglutinated foraminifera are absent from samples of the former site (QT5, QT6, QT7) whilst calcareous foraminifera are absent from the latter (QT1, QT2, QT3, QT4), where salinity ranges vary from 10‰ ~ 12‰.

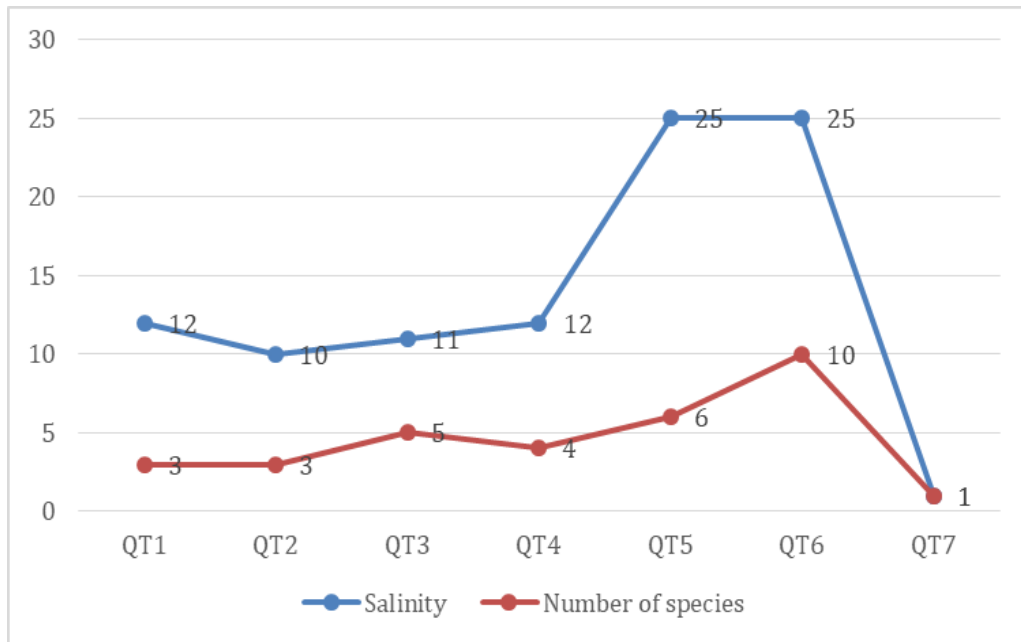
The abundance and diversity of living benthic foraminifera give clues about the environmental status of the estuaries in Quang Tri. Table 2 suggests that the diversity of foraminiferal species reflects salinity of water. Shannon-Weiner index value obtained for foraminifera was estimated to be 0 (lowest) at QT7 with lowest salinity is 1‰ and 2.011 (highest) at QT6 with highest salinity is 25‰. Similar to Simpson index, value was estimated lowest to be 1.000 at QT7 and highest to be 5.124 at QT6. Diverse of QT5 and QT6 increasing to 6 and 10 species meanwhile diverse of QT1 is 3, QT2 is 3, QT3 is 5, QT4 is 4 and QT7 is only 1.



**Figure 10 - Proportion of tests of living benthic foraminifera in Cua Tung and Cua Viet estuaries, Quang Tri Province, April 2017**

Station	Abundance	Species	H'	D	Salinity(‰)
QT1	30	3	0.945	2.261	12
QT2	255	3	0.513	1.374	10
QT3	86	5	0.927	1.832	11
QT4	43	4	0.822	1.786	12
QT5	45	6	1.21	2.325	25
QT6	56	10	2.011	5.124	25
QT7	5	1	0	1	1

**Table 3 - Diversity of foraminiferal species from samples of Cua Tung and Cua Viet estuaries, Quang Tri Province, survey April 2017. H', Shannon-Weiner Diversity Index; D, Simpson Diversity Index**



**Figure 11 - Correlation between salinity and benthic foraminiferal diversity (richness) in samples from Cua Viet and Cua Tung estuaries, Quang Tri Province, Survey April 2017**

Analyses of samples from Cua Tung and Cua Viet estuaries, September 2017

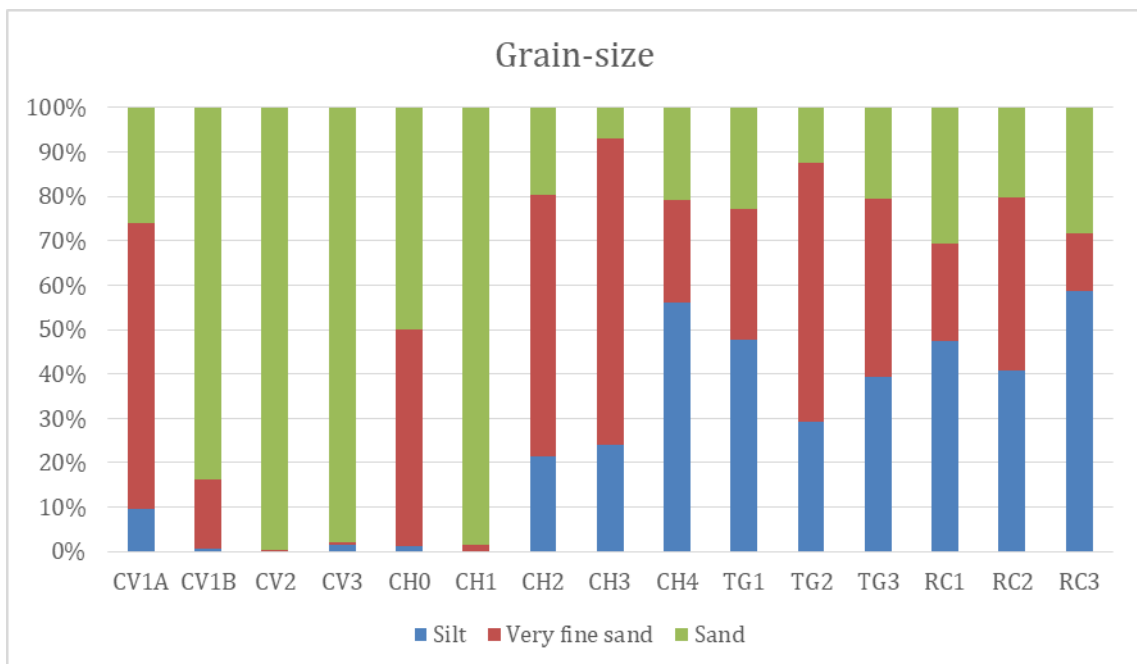
*Sediment characteristics*

Sand is the most abundant fraction in the stations of Cua Viet estuary (77.75% of total Cua Viet sediment samples) and it contributes 26.10% to 99.96% of sediment samples in Cua Viet estuary with the highest percentage is 99.96% in CV2. In this area, very fine sand is the second abundant fraction with 19.54%. The highest contents of very fine sand are found in CV1A (64.27%). Silt is rarely found in this area and contributes average on 2.93%

In Cau Hai lagoon sampling area, sand and very fine sand are dominant fractions of sediment samples with 44.09% and 39.56%, meanwhile, silt contributes 16.35% in total sediment samples.

Station	<63 $\mu$ m	63 $\mu$ m < sediment < 125 $\mu$ m	>125 $\mu$ m
CV1A	1.77	11.82	4.8
CV1B	0.14	3.76	20.16
CV2	0.02	0.04	19.59
CV3	0.26	0.15	18.2
CH0	0.26	9.87	10.09
CH1	0.03	0.29	23.04
CH2	3.27	9.03	3.03
CH3	4.89	14.06	1.44
CH4	6.38	2.62	2.38
TG1	4.97	3.07	2.38
TG2	2.57	5.12	1.08
TG3	3.34	3.38	1.75
RC1	3.21	1.47	2.08
RC2	5.38	5.11	2.66
RC3	3.64	0.81	1.76

**Table 4 - Grain-size fraction compositions**

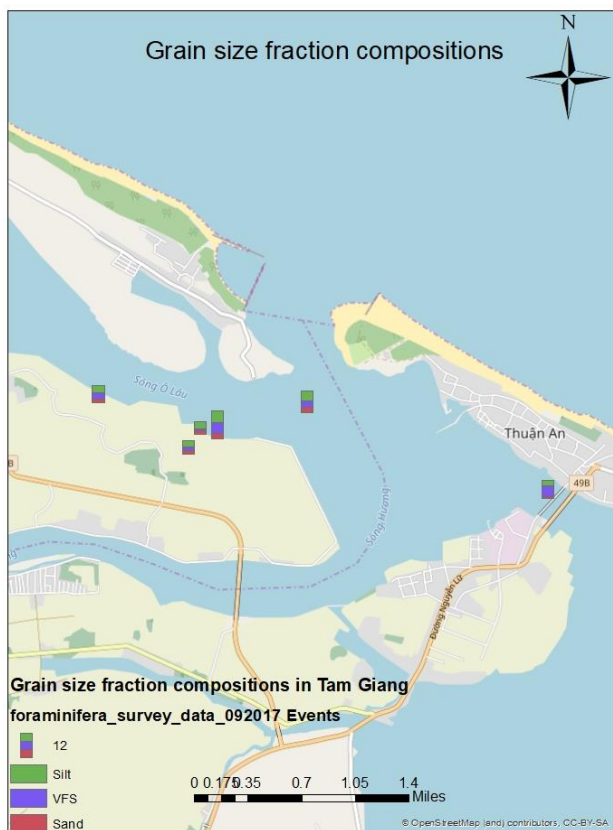


**Figure 12 - Sediment grain-size variations in 15 stations within 4 sampling areas**





**Figure 13 - Grain size fraction compositions in samples from the Cau Hai inlet (Thua Thien Hue Province)**



**Figure 14 - Grain size fraction compositions in Tam Giang**



**Figure 15 - Grain size fraction compositions in Cua Viet**

### *Foraminifera assemblages*

Taking into account sample grain size distributions, foraminifera assemblages have been analyzed and statistically evaluated, for each of the sampled stations, and compared against physico-chemical water parameters, in order to draw conclusions on impacts affecting urbanized and industrialized sites.

Table 4 shows the diversity and abundances of the foraminifera genus in Tam Giang – Cau Hai lagoon. *Ammonia* sp., *Elphidium* sp., *Miliammina* sp., *Haplophragmoides* sp., *Trochammina* sp., *Textularia* sp., *Ammobaculites* sp., *Ammoscalaria* sp. occur frequently in the sampling area (>50% stations). The most abundant genus is *Ammobaculites* sp. with 513 specimens distribute in almost every station except RC3 and CH0. Its abundance varies significantly among the sampling areas and the highest density is in RC2 with 133 specimens. *Ammonia* sp. is the second most abundant specimens (271 specimens), which occurs in TG2, RC2, and all the stations in Cau Hai lagoon. Its abundance varies as well significantly among the sampling areas and highest abundances are found in CH2 station.

Table 4A

Station/Species names	CV1A	CV1B	CV2	CV3	CH0	CH1	CH2	CH3	CH4
<i>Ammonia</i> sp.	192	16	3	27	7	3	131	52	26
<i>Nonion</i> sp.	2	1	0	0	0	0	0	0	1
<i>Nonionella</i> sp.	0	0	0	0	0	2	0	0	0
<i>Eggerella</i> sp.	2	0	0	0	0	0	0	0	0
<i>Reussella</i> sp.	12	1	0	0	2	1	6	2	0
<i>Fijiella</i> sp.	2	0	0	0	0	0	0	0	0
<i>Neocassidulina</i> sp.	4	1	0	0	1	0	0	0	0
<i>Hanzawaia</i> sp.	5	21	1	0	27	0	7	0	0
<i>Cibicides</i> sp.	4	0	0	0	0	0	0	0	0
<i>Elphidium</i> sp.	7	50	3	2	18	11	12	13	2
<i>Bolivina</i> sp.	9	0	0	0	4	0	2	0	0
<i>Bolivinellina</i> sp.	0	5	1	0	0	0	0	0	0
<i>Pararotalia</i> sp.	0	1	0	0	0	0	0	0	0
<i>Asterigerinata</i> sp.	0	3	0	0	4	0	0	0	0
<i>Guttulina</i> sp.	0	2	0	0	1	1	0	0	0
<i>Calcarina</i> sp.	0	0	0	0	1	0	0	0	0
<i>Hyalinea</i> sp.	0	0	0	0	0	3	0	0	0
<i>Rosalina</i> sp.	0	0	0	0	0	1	1	3	0
<i>Glandulina</i> sp.	0	0	0	0	0	0	1	0	0
<i>Quinqueloculina</i> sp.	6	21	2	0	17	4	7	1	0
<i>Triloculina</i> sp.	2	4	0	0	6	4	0	0	0
<i>Miliammina</i> sp.	12	15	1	0	12	9	3	2	0
<i>Pyrgo</i> sp.	1	4	0	0	4	9	0	0	0
<i>Articulina</i> sp.	4	0	0	0	3	0	0	0	0
<i>Spiroloculina</i> sp.	1	1	0	0	0	1	0	0	0
<i>Planispirinella</i> sp.	0	8	0	0	3	0	0	1	0
<i>Massilina</i> sp.	0	10	0	0	0	0	0	0	0
<i>Edentostomina</i> sp.	0	1	0	0	0	0	0	0	0
<i>Miliolinella</i> sp.	0	0	0	0	8	0	0	0	0
<i>Haplophragmoides</i> sp.	34	0	0	2	0	0	4	0	5
<i>Trochammina</i> sp.	4	3	0	0	0	0	12	3	17
<i>Textularia</i> sp.	26	1	0	0	1	1	55	23	19
<i>Ammobaculites</i> sp.	70	9	0	4	0	1	82	31	33
<i>Ammoscalaria</i>	4	0	0	0	0	0	21	0	14
<i>Nouria</i> sp.	1	0	0	0	0	0	0	0	0
<i>Reophax</i> sp.	1	0	0	0	0	0	28	19	21
<i>Sahulia</i> sp.	3	0	0	0	0	0	0	0	0

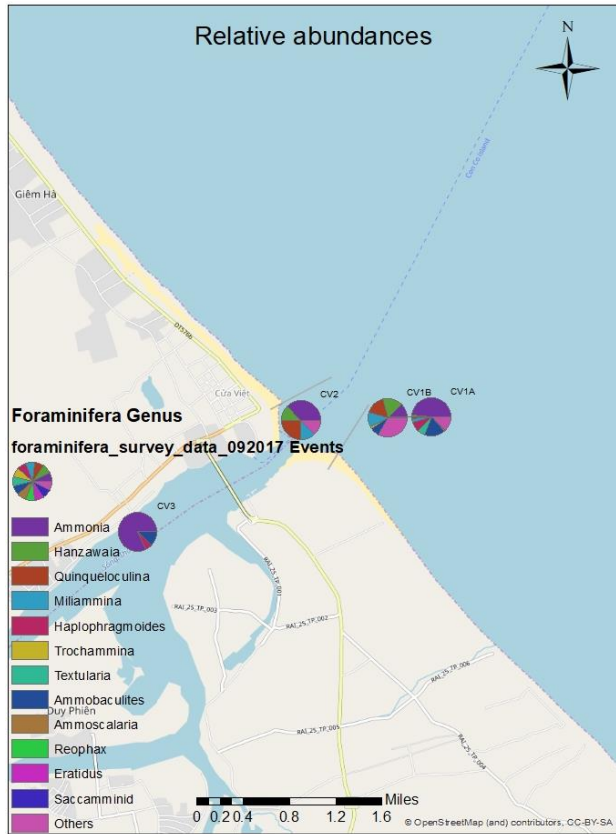
<i>Technitella</i> sp.	1	0	0	0	0	0	0	0	0
<i>Ammotium</i> sp.	0	0	0	0	0	0	0	0	0
<i>Eratidus</i> sp.	0	0	0	0	0	0	0	0	0
<i>Psammosphaera</i> sp.	0	0	0	0	0	0	0	0	0
<i>Arenoparrella</i> sp.	0	0	0	0	0	0	0	0	0
<i>Jadammina</i> sp.	0	0	0	0	0	0	0	0	0
<i>Saccamminid</i> sp.	0	0	0	0	0	0	0	0	0
<b>Total</b>	<b>409</b>	<b>178</b>	<b>11</b>	<b>35</b>	<b>119</b>	<b>51</b>	<b>372</b>	<b>150</b>	<b>138</b>

<b>Table 4B</b>						
<b>Station/Species names</b>	<b>TG1</b>	<b>TG2</b>	<b>TG3</b>	<b>RC1</b>	<b>RC2</b>	<b>RC3</b>
<i>Ammonia</i> sp.	0	46	0	0	6	0
<i>Nonion</i> sp.	0	1	0	0	0	0
<i>Nonionella</i> sp.	0	0	0	0	0	0
<i>Eggerella</i> sp.	0	0	0	1	0	0
<i>Reussella</i> sp.	0	2	0	0	0	0
<i>Fijiella</i> sp.	0	0	0	0	0	0
<i>Neocassidulina</i> sp.	0	0	0	0	0	0
<i>Hanzawaia</i> sp.	0	1	0	0	0	0
<i>Cibicides</i> sp.	0	1	0	0	0	0
<i>Elphidium</i> sp.	0	1	0	0	0	0
<i>Bolivina</i> sp.	0	0	0	0	0	0
<i>Bolivinellina</i> sp.	0	0	0	0	0	0
<i>Pararotalia</i> sp.	0	0	0	0	0	0
<i>Asterigerinata</i> sp.	0	0	0	0	0	0
<i>Guttulina</i> sp.	0	0	0	0	0	0
<i>Calcarina</i> sp.	0	0	0	0	0	0
<i>Hyalinea</i> sp.	0	0	0	0	0	0
<i>Rosalina</i> sp.	0	1	0	0	0	0
<i>Glandulina</i> sp.	0	0	0	0	0	0
<i>Quinqueloculina</i> sp.	0	1	0	0	0	0
<i>Triloculina</i> sp.	0	0	0	0	0	0
<i>Miliammina</i> sp.	21	1	41	3	56	1
<i>Pyrgo</i> sp.	0	0	0	0	0	0
<i>Articulina</i> sp.	0	0	0	0	0	0
<i>Spiroloculina</i> sp.	0	0	0	0	0	0
<i>Planispirinella</i> sp.	0	0	0	0	0	0
<i>Massilina</i> sp.	0	0	0	0	0	0
<i>Edentostomina</i> sp.	0	0	0	0	0	0
<i>Miliolinella</i> sp.	0	0	0	0	0	0

<i>Haplophragmoides</i> sp.	35	3	24	0	2	0
<i>Trochammina</i> sp.	6	0	23	3	3	0
<i>Textularia</i> sp.	3	21	3	0	0	0
<i>Ammobaculites</i> sp.	98	23	98	14	133	0
<i>Ammoscalaria</i>	12	17	11	0	17	0
<i>Nouria</i> sp.	0	0	0	0	4	0
<i>Reophax</i> sp.	47	0	5	0	0	0
<i>Sahulia</i> sp.	0	0	0	0	0	0
<i>Techinitella</i> sp.	0	0	0	0	0	0
<i>Ammotium</i> sp.	1	0	0	0	4	1
<i>Eratidus</i> sp.	54	0	18	0	11	0
<i>Psammosphaera</i> sp.	0	1	0	0	0	0
<i>Arenoparrella</i> sp.	0	0	0	0	0	3
<i>Jadammina</i> sp.	0	0	0	0	1	0
<i>Saccamminid</i> sp.	0	0	0	0	58	0
<b>Total</b>	<b>277</b>	<b>120</b>	<b>223</b>	<b>21</b>	<b>295</b>	<b>5</b>

**Table 5 - Foraminifera species abundance from samples from Cua Tung and Cua Viet estuaries of Quang Tri Province, Tam Giang-Cau Hai lagoon, Thua Thien Hue Province**

CH0 is the station with the highest diversity, with 17 genera existing at the sampling time. The second diversity station is CH2 with 15 genera present, then TG2 and CH1 station with 14 genera occur. Although that, CH2 is the most abundant station with 372 foraminifera specimens. The second abundant station is RC2 (295 specimens) with 11 genera exist in sediment sample.



**Figure 16 – Relative abundances of the main living benthic foraminifera genus in Cua Viet estuary, Quang Tri Province, September 2017**



**Figure 17 – Relative abundances of the main living benthic foraminifera genus in Tam Giang lagoon, Thua Thien Hue Province, September 2017**

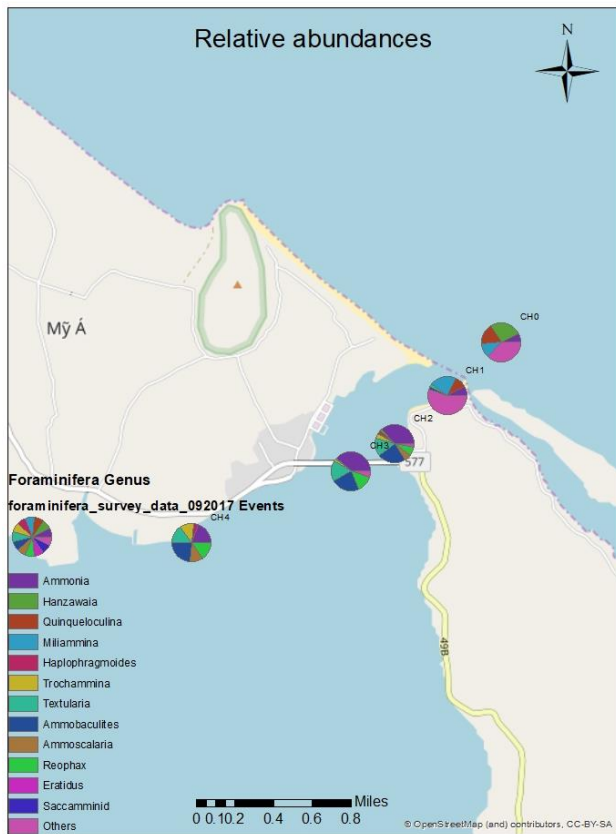


Figure 18 - Relative abundances of the main living benthic foraminifera genus in Cau Hai lagoon, Thua Thien Hue Province, September 2017

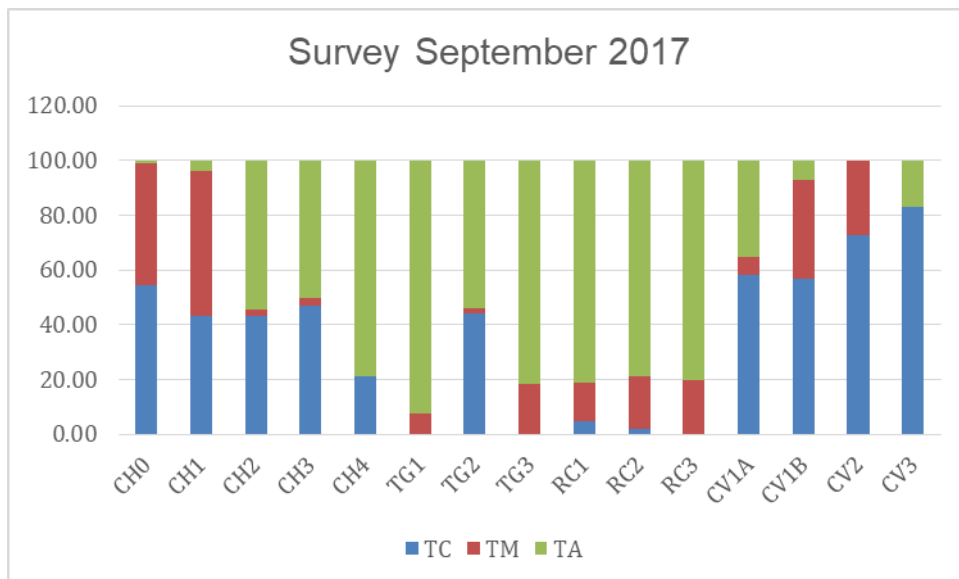


Figure 19 - Percentage of foraminifera species from samples from Cua Tung and Cua Viet estuaries of Quang Tri Province, Tam Giang-Cau Hai lagoon, Thua Thien Hue Province. TC, Total Calcareous; TMC, Total Miliolids Calcareous; TA, Total Agglutinated.

Figure 19 shows that, in different locations, the abundance of foraminifera group by shell component are different. Agglutinated species are rarely existing in station CH0, CH1, CV1A (0.84%, 3.92% and 7.30%) and absent in CV2 but occupy majority the living foraminifera assemblage in the other stations (>50%). The percentage of total calcareous species are high in stations CH0, CH1, CH3, TG2, all of Cua Viet estuary stations (> 43%) but low in stations CH4, RC1, RC2 (<22%) and absent in TG1, TG3 and RC3. Meanwhile, the percentage of total miliolids calcareous species are high in stations CH0, CH1, CV1B and CV2 (>35%), low in stations TG1, TG3, RC1, RC2, RC3 and CV1A (6%~27%), very low in stations CH2, CH3, TG2 (1% ~ 3%) and absent in station CH4 and CV3.

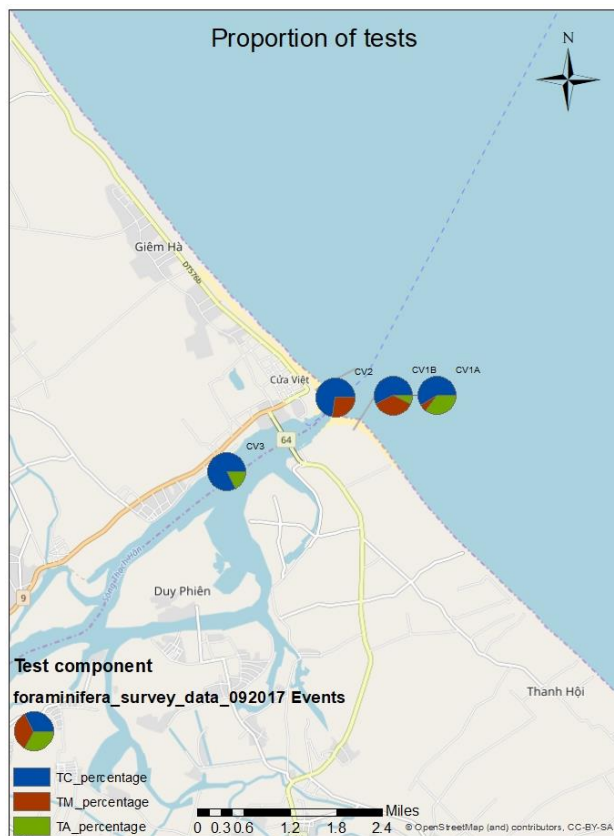
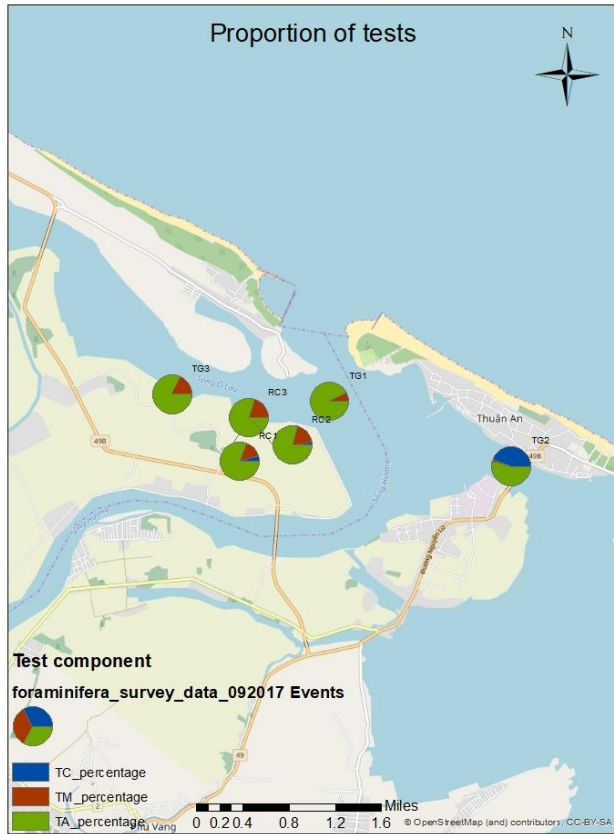


Figure 20 – Proportion of tests of living benthic foraminifera in Cua Viet estuary, Quang Tri Province, September 2017

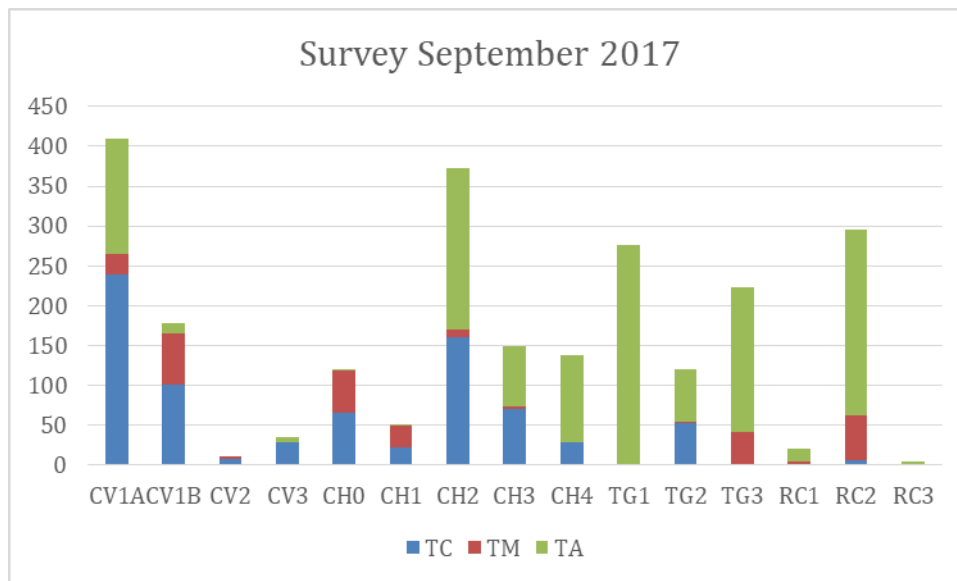




**Figure 21 – Proportion of tests of living benthic foraminifera in Tam Giang lagoon, Thua Thien Hue Province, September 2017**



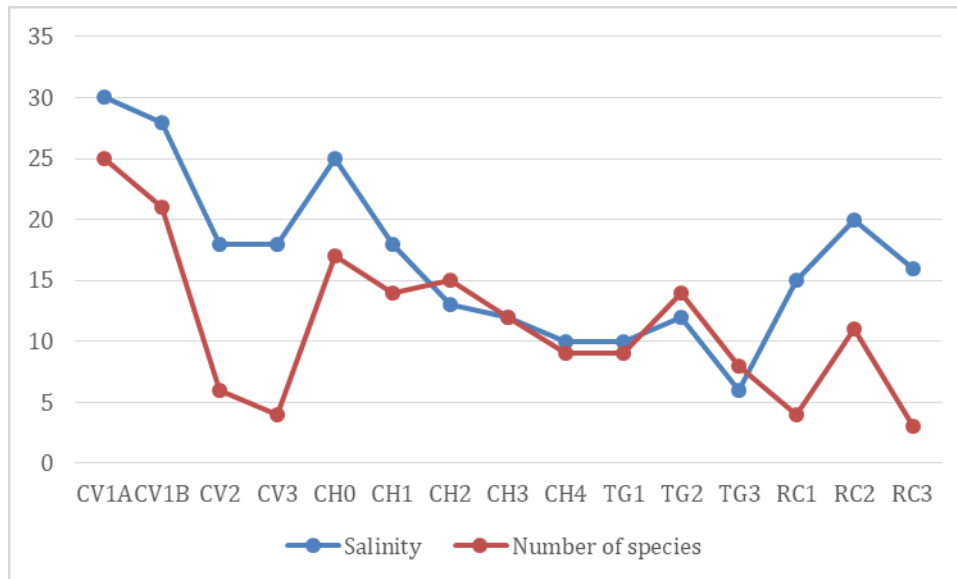
**Figure 22 - Proportion of tests of living benthic foraminifera in Cau Hai lagoon, Thua Thien Hue Province, September 2017**



**Figure 23 - Percentage of foraminifera species from samples from Cua Tung and Cua Viet estuaries of Quang Tri Province, Tam Giang-Cau Hai lagoon, Thua Thien Hue Province. TC, Total Calcareous; TMC, Total Miliolids Calcareous; TA, Total Agglutinated.**

Station	Abundance	Salinity	H	D	Number of species
CV1A	409	30	1.955428	3.785923	25
CV1B	178	28	2.400856	7.554261	21
CV2	11	18	1.672625	4.780845	6
CV3	35	18	0.775195	1.626826	4
CH0	119	25	2.387369	8.286132	17
CH1	51	18	2.265253	7.58309	14
CH2	372	13	1.925614	4.840632	15
CH3	150	12	1.820673	4.797181	12
CH4	138	10	1.923489	6.179104	9
TG1	277	10	1.732587	4.626409	9
TG2	120	12	1.745695	6.805293	14
TG3	223	6	1.641614	3.864247	8
RC1	21	15	0.971262	2.051163	4
RC2	295	20	1.577123	3.525992	11
RC3	5	16	0.950271	2.272727	3

**Table 6 - Diversity of foraminiferal species of Cua Tung and Cua Viet estuaries of Quang Tri Province, Tam Giang-Cau Hai lagoon, Thua Thien Hue Province. H', Shannon-Weiner Diversity Index; D, Simpson Diversity Index**



**Figure 24 - Correlation between in salinity and benthic foraminiferal diversity (richness) of Cua Tung and Cua Viet estuaries of Quang Tri Province and of Tam Giang-Cau Hai lagoon, Thua Thien Hue Province.**

Analysis of environmental data 2006-2018

Parameters indicating water quality such as bacteria counts and chlorophyll-a, both in the water and in the sediment in the period 2006-2012 showed values within the ranges of acceptability set up by the Government; that meaning that pollution from sea into lagoon through the inlets (closed to maritime port facilities) and from aquaculture is still modest. In particular, through the estuary of Tu Hien organic matter and toxic substances as  $\text{NO}_3$  and  $\text{NH}_3$  are reduced at times of enhanced water exchange with the sea. These observations are in harmony with findings of Le Cong Tuan (2008) who noted in his paper that also for the Sam Chuon lagoon (yet distant from the Thuan An inlet and hydrologically rather restricted) more than 70% of toxic substances were washed out after one days of water exchanges between sea and lagoon.

Most of water quality parameters of the Tam Giang-Cau Hai lagoon such as temperature, pH, DO, BOD5, ammonia, toxic metals (Cu, Pb, Zn and Cd) and OCs (DDTs and HCHs) met the requirement of Vietnam standard TCVN 2006 applied to the coastal water used for aquaculture. Although  $\text{PO}_4$  and  $\text{NO}_3$  concentration in the lagoon water was low ( $\text{N-NO}_3 < 0.26 \text{ mg/L}$  and  $\text{P-PO}_4 < 0.10 \text{ mg/l}$ ), total N (TN) concentrations were higher 1 mg/l higher than standards at several sites. The TN concentrations are one of the potential causes of eutrophication, especially in the dry

season. The results of chlorophyll-a in the lagoon water (higher than 5 µg/L at most sites in the dry season) were concurrent evidence of eutrophication, whilst <sup>45</sup>P, based on the ratio TN/TP in water column, was identified as a main limiting factor.

In Thuy Tu and Cau Hai area, water quality parameters such as temperature, pH and especially salinity are quite suitable for black tiger shrimp culture, therefore widely practiced and cause of constant pollution hazards in the dry season. Uncontrolled aquaculture caused marked environmental disadvantages to the lagoon environment such as increase in organic matter content, high bacterial counts and persistent eutrophication. In this study, the organic matter content in estuaries and lagoon sediments were considered for their effects on benthic foraminiferal assemblages and use of this taxonomic group as an indicator of abnormal trophic states.

Both proxies of primary production and the measurement of the detrital fractions of sediment organic matter allows to characterize eutrophic and even hypertrophic habitats.

Although the concentration of toxic chemicals (OCs and toxic metals) in water of most of the lagoon habitats remote from urban settlements is fairly low, HCHs level in the sediment is rather high at several sites (*e.g.* near Thuan An industrial hub), causing adverse effects on lagoon aquatic organism.

Toxic-metal concentration in lagoon sediments was in general lower than the levels of ISQG and PEL applied to the marine sediment. However, Cd and Cu content at several sites was higher than the ISQG set standards.

Most of the water quality parameters in the canals adjacent to the aquaculture ponds met the requirement of the Vietnam Standard TCVN 2006 applied to coastal waters used for aquaculture and 28 TCVN 171/2001 applied to intensive culture of black tiger shrimp. However, aquaculture activities need to be somehow regulated in order to mitigate risk of habitat deterioration.

## **Discussion on characteristics of lagoon and estuarine environments of Central Vietnam**

### Climate of Central Vietnam

Central Vietnam seasonality relates to regional climate which is transitional between the monsoonal regime prevailing in the south of the country, with rainy summers and dry winters, and the tropical four-season regime characteristic of the north.

Located at the southern end of the North Vietnam's continental climate regime and protected by the Hai Van Pass, the Thua Thien Hue and Quang Tri provinces suffer the a worst and most harsh weather regime than other areas in the country.

Central Vietnam is affected by the monsoon: their direction is from northeast, in winter, and southwest, in summer. Due to its proximity to the area of maximum rainfall in absolute values (Bach Ma range, with average rainfall in excess of 3,000 mm/year), the lagoon belongs to the most intense precipitations area, whose annual average is approximately 2,744 mm/year, higher than the average country rainfall of 1,900 mm/year.

Typical climatic characteristic of this region is that the rainy season often comes late compared to other monsoon-dominate areas, usually beginning in September until December, with a total rainfall of 2,000 mm approximately (72.8%). Rainfall is highest in October (740 mm, 26.96%), persisting through November. Rainfall of the remaining months is less important.

Highest rainfall concentrated during the few fall months easily lead to floods. On the other hand, spring and early summers are definitely dryer because of limited precipitations.

Storms annually batter the coast, with 0 to 4 storms yearly with wind speed from 20 to 40 m/sec. During the last century, there were 0 to 8 storm/year. Storms are accompanied by heavy rains, persisting from 2-3 to 5-6 days, resulting in generalized inundation of the whole coastal lowland. The storm season is from June to November, most frequently from July to October.

Sunlight radiation, air temperature, evaporation, and drought. The annual total days of sunlight is high, from 1,900 to 2,000 hours, resulting in rather high annual average temperature (25.2 °C). In spring and summer, hours of sunlight concentrate from May to

August, which correlate with highest recorded temperatures (as much as 39-40 °C). These will lead to excess evaporation, causing drought. In winter, conversely, sunlight is less intense (100-110 hours/month): the lowest sunlight radiation occurs in December, coincident with lowest average and minimum temperatures (11.4 °C and 8.8°C, respectively). In the winter months, water evaporation is about 37-74 mm/month, far exceeded by intense rainfall (2,000mm).

#### Considerations on foraminiferal ecology

The living benthic foraminifera of Cua Tung and Cua Viet estuaries and of Tam Giang – Cau Hai lagoon are abundant and moderately diversified, but their abundance is closely related to salinity. General trends with increasing salinity include: decreasing abundance of agglutinated species, increasing abundance of porcellanaceous species, calcareous perforate species and increasing diversity. This preliminary research has shown that benthic foraminiferal abundance correlates to the environmental status and encourage exploiting their use as bio-indicators also in relationship with a eutrophic gradient in the estuary.

#### Tam Giang-Cau Hai lagoon physiography and ecosystems.

The Tam Giang-Cau Hai lagoon is a complex physiographic wetland system, consisting of a number of interconnected, yet hydrologically independent sub-basins, each one having its own morphological features, hydro-chemical characteristics and ecology.

The physiographical complexity of the lagoon system relates on the processes that shaped the landscape in the Holocene, following the post-glacial rise of the sea level, and on the interplay of fluvial and marine processes.

The ecological complexities depend on i) the compartmented nature of the lagoon sub-basins, ii) their hydrology (i.e. the circulation of the water masses within the lagoon and exchange with the sea; iii) the chemical state of the waters, including pollutant load from aquaculture, agriculture and urban sources and iv) the climate and its seasonality.

The Tam Giang-Cau Hai lagoon sub-basins morphology originated from the inundation of a relic morphology consisting of beach ridges, deltas and incised valleys, following the Holocene rise in sea level.

## The Tam Giang Cau Hai sub-basins and active river system



Figure 25 - The Tam Giang-Cau Hai lagoon sub-basins and their tributary river network. From northwest, the O Lau river discharging into the Tam Giang-Cau Hai lagoon; the Bo and Huong rivers, directly discharging into the Eastern Sea though the Thuan An inlet and the Truoi river, discharging into the Cau Hai sub-basin.

Figure 25 shows the location and nomenclature of the main Tam Giang-Cau Hai lagoon sub-basins.

The upper Tam Giang is an elongated broad basin, natural extension of the often-submerged O' Lau river mouth, flanked on both side by a agricultural land.

The lower Tam Giang lagoon is a narrow corridor flanked by multiple rows of earthen aquaculture ponds encroaching the water surface, on the shores of the communes of Quang Cong, Hai Duong (to the east), Quang An, Quang Phuoc and Quang Dien (to the west).

The Huong river active delta extends from the Thuan An strait to the Huong Phong coastline of Ru Cha, comprising the Huong river mouth and sub-deltaic lobes of the Bo river

The Sam Chuon lagoon is an embayment that opens up between the corner of Thuy Dien and Thuan An, with three tributaries and one relic delta (An Truyen) nowadays all inactive (part of the sub-modern Huong River delta system),

The Thuy Tu channel is a waterway confined between modern and sub-modern beach-ridges, extending from the Thuan An strait to Cau Hai and building an inner flood-tidal delta lobe (Ba Con) at its debouch into this basin.

The Cau Hai sub-basin, semi-circular in shape, is surrounded by forested steep slopes, subject to intense runoff during the rainy season, with three main tributaries: Dai Giang,

Thieu-Hoa and Truoi rivers. The Thuy Tu channel ends in Cau Hai and a tidal inlet (Tu Hien) opens towards the East Sea (Bien Dong).

#### Tam Giang-Cau Hai lagoon hydrology.

The lagoon physiography has influence on the overall circulation and the exchange of water masses with the sea, restricted by the longshore transport of sediments silting the inlets and the micro-tidal regime that hinders the penetration inland of marine waters during the flood-ebb cycle. Lacking tidal forcing, the lagoon circulation is driven by riverine flow except than in proximity of the inlets, with a local effect by wind fetch.

These hydrologic conditions have negative consequences on inlet stability, lagoon siltation, ventilation and flushing out of pollutants.

The hydrology of the lagoon has different characteristics in the different sub-basins: that of a normal river outflow in the northern Tam Giang; dominated by the interplay of the Huong river discharge and tides, in the central sector of Huong Tra and Thuan An; restricted in the whole southern basins, because of adverse conditions created by nature and man. The Cau Hai basin is an enclosed pond with intermittent sea connection and seasonal runoff that make salinity low and to fluctuate erratically. The central Thuy Tu is a relic depression between successive beach-ridge systems, nowadays hydrologically inactive because of human interventions (aquaculture and river diversion). Sam Chuon is a restricted embayment, enclosed within the southern pre-modern Huong River delta. Moreover, localized hinterland uplift (Phu Bai) contributed to disrupt the river pattern with consequences on the whole hydrological balance of the coast.

Hydrology and water masses exchange is a critical factor in Tam Giang-Cau Hai, as these control ventilation, oxygen distribution or depletion, pollutant and nutrient dispersal, sedimentation; not to mention the role on biological life cycle, productivity, biota and vegetation that contribute to characterize ecosystems. Human interventions, particularly the construction of aquaculture ponds and deployment fishing implements (bottom nets, stake traps and net enclosures) aggravated the situation, contributing to anoxia, eutrophication and posing additional stress onto an impoverished biological stock.



The longshore-current sediment transport at the inlets and estuaries of Central Vietnam show patterns related to the winter monsoonal regime, when dominant winds are from north and north-west.

The general pattern of sediment transport is longshore, along the beaches laying on both sides seaward of the inlets. Waves, on the other hand, transport the sediment landward, from the ebb tidal delta and offshore bars towards the lagoon interior contributing to the inlet siltation.

Due to the shallow inlet channel, coarse sediments transported longshore sediments enter the narrow passages and estuaries facilitated by the entering tidal-flood waters.

Due to prevailing north-easterly winds, inlets and estuaries are sheltered from any south-easterly transport.

The Tu Hien inlet, in particular, is protected by the Cape of Chan May Tay. Due to the lack of sediment supply from its southern coast, sediments entering the Tu Hien inlet mainly follow the northern marginal flood currents and part of it is flushed out by the ebb currents. The seasonal monsoon regime has a strong influence on the wave climate and sediment transport pattern in the inlet areas. The seasonal variation of sediment transport induced by waves are extracted from results of hydrological modelling (Fig. 26 Matticchio and Stefanon, 2011).

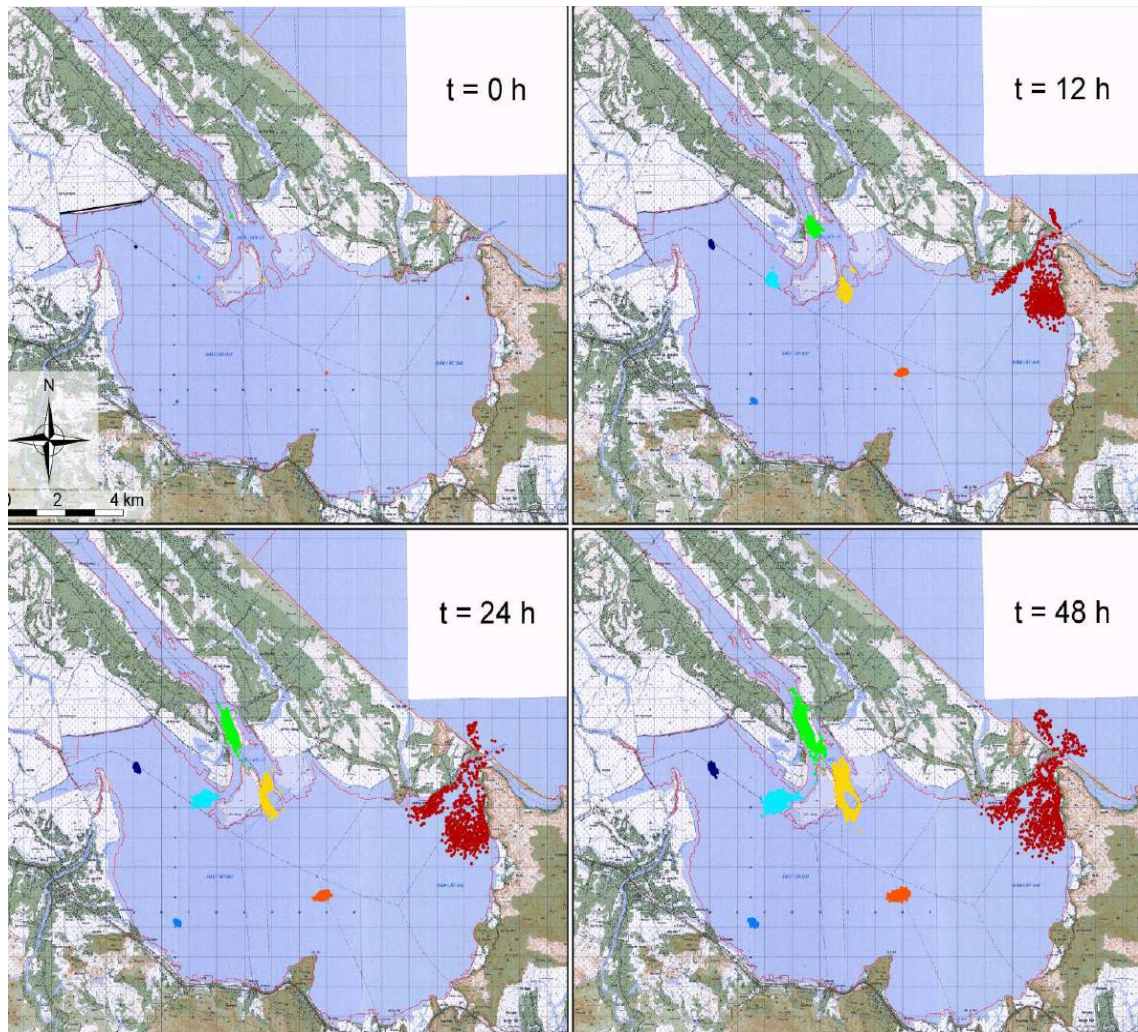
#### Tam Giang-Cau Hai lagoon water chemistry

Hydro-chemical parameter and their variation through time offer indirect clues to water circulation, pollution levels and a method to characterize lagoon ecosystems and habitats.

Salinity is an indicator of water flows and exchange of water masses between rivers, lagoon and the sea and function of climate (evaporation over precipitations). It is one of the parameters to defines ecosystems. Salinity and its fluctuations also affect the aquaculture.

Salinity fluctuates seasonally: on average, salinity builds up in the lagoon during the spring and early summer; it is generally high up to normal marine values close to two

inlets (Thuan An in the center and Tu Hien, in the south) during the summer (May), when excess evaporation and limited river discharge drives marine waters inland.



**Figure 26 – Diffusion and dispersion of the spots during the first 48 hours of simulation,forced by the astronomical tide of April 2011.**

Conversely, salinity is on average low in the northern lagoon of Tam Giang because of the steady outflow of the O’ Lau river, causing a regular gradient from nearly fresh to saline waters along the channel, from the river mouth to the tidal inlet (e.g. Fig. 27, blue arrow on the map for May).

Salinity begins to decrease from August, with marked drops in the event of exceptional precipitations. During these times, for example in Cau Hai, fresh water drained from the Dai Giang-Thieu-Hoa river plain and the Truoi river is evacuated through the Tu Hien inlet and partly channeled into the Thuy Tu, whenever excess outflow through the inlet is hindered (Fig. 27, blue arrows in the August map).

BOD (Biological Oxygen Demand)<sup>1</sup> and COD (Chemical Oxygen Demand)<sup>2</sup> are laboratory tests used to infer the level of organic pollutants present in riverine and lake waters and an indicator of water quality. Data from Tam Giang-Cau Hai display a situation which remains within the limits of standards set up by governments for water quality throughout the year, with an overall increase during the spring and summer months in coincidence with the aquaculture season. The correlation between fish farming and the dispersal of organic pollutants is evident: in the area comprised between the Thuan An narrow and the northern Thuy Tu for example, BOD and COD values reach maxima in the summer as a result of organics build-up, in the lack of efficient flushing. Elsewhere, the overall level of organic pollution is moderate and drops in the winter months, as a consequence of typhoons and rains.

The pH maintains within the normal range throughout the year and in virtually all lagoon sectors, except for the northern Tam Giang lagoon at the mouth of the O' Lau river. There, the influx of river waters seems to promote a marked shift towards extreme acidic values during summer, when agricultural activity is at its climax. The most likely reason for this pH excursion could be sought in the discharge of agricultural wastewaters in the lagoon, but other mechanisms such as decomposition of organic matter in the presence of dissolved oxygen (*e.g.* by oxygen reduction) or erosion of tropical coastal soils containing iron-sulphides (release of acid sulphate) could be possible.

The total phosphorous is on average low, in most lagoon sectors and in all seasons. The highest values are recorded in the Sam Chuon basin and in the northern Tam Giang-O' Lau river mouth, most evident during an episode of increased runoff (August 2006), associated to a drop in pH – a sign of contribution from agricultural effluents. Nitrogen does not strictly echo the pattern of phosphorous, but equally suggest a more diffused dispersal in the aquatic environment, possibly from undigested fish feed or manure from agricultural activities around the lagoon, with episodes of build-up in the less ventilated parts of the basin (Sam Chuon). N and P bespeak of a pronounced tendency of the

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<sup>1</sup> BOD test indicates how fast biological organisms use up oxygen in a body of water. It is used in water quality management and assessment, ecology and environmental science to crudely indicate the quality of a water source.

<sup>2</sup> COD test is commonly used to indirectly measure the amount of organic compounds pollutants in water (*e.g.* lakes and rivers) and represent a useful measure of water quality.

lagoon to eutrophication, eventually promoting algal blooms (see peak of chlorophyll in Sam Chuon) and anomalous fluctuation in the dissolved oxygen.

Coliform bacterial load in the lagoon waters is generally high, with peaks at places (*e.g.* Tu Hien inlet, in August) three orders of magnitude higher than the national standard for aquatic cultivation areas (1,000 NPM/100ml). Despite the lack of large urban area along the lagoon shores, fecal-coliforms component indicates relevant human and/or animal livestock contributions, hence a diffuse release of wastewaters from communities.

The presence of pesticides and organochlorine compounds<sup>3</sup> used as agricultural insecticides in the sediment of the lagoon is noteworthy, although they do not represent an immediate hazard as they are stored in the sediment. These chemicals are largely banned in most countries nowadays, but the use in Vietnam is still continued and is a potential hazard in those few areas of enhanced riverine discharge. However, high water turbidity because of suspended clay particles causes chemical absorption of POPs and thus their rapid removal by burial.

Overall, the contaminant load of the lagoon (phosphorous, nitrogen, ammonia) and indicators of eutrophication (chlorophyll) are not severe, if one considers the critical of some of the basins. The dissolved oxygen level never falls below reference minimum standard for Vietnam, that means that oxygen depletion never develops to an extent to become harmful to aquatic animal life.

The lagoon system has a pronounced seasonality, with progressive water-quality deterioration during spring and summer, in coincidence with the farming activities, and return to more normal conditions during autumnal typhoons, when toxic waste and contaminants are flushed out.

#### Hydro-chemical characterization of the water masses and lagoon seasonality

In order to define ecologic zones, a statistical cluster analysis was applied to datasets acquired from the monitoring stations of 2006, using the four fundamental variables

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<sup>3</sup> An organochloride, organochlorine, chlorocarbon, or chlorinated solvent is an organic compound containing at least one covalently bonded chlorine atom. Their wide structural variety and divergent chemical properties lead to a broad range of uses. These chemicals are typically nonaqueous and are usually denser than water due to the presence of heavy chlorine atoms. Many pesticides widely used in agriculture are organochlorides. These include DDT, dicofol, heptachlor, endosulfan, chlordane, mirex, and pentachlorophenol. These can be either hydrophilic or hydrophobic depending on their molecular structure.

characterizing the water body: temperature, salinity, dissolved oxygen concentration and pH<sup>4</sup>. Based on these parameters, the stations were grouped into clusters with at least 5% dissimilarity, thus displaying water masses affinities, seasonality and to some extent exchange among basins. Water-mass affinities, in particular, provided hints on lagoon circulation in different seasons, an aspect often overlooked and insufficiently investigated.

*O'Lao river mouth and Tam Giang basin.*

In northern Tam Giang basin, including the O' Lau river mouth and upstream from the Huong Phong narrows, water masses display, in the month of April, a gradient from riverine type (see 1 in the legend of Fig. 27) to brackish (type 2), partially restricted (type 3). In May, a month of limited continental runoff, the fresh fluvial water of the O' Lau river remained confined in the vicinities of the mouth, while the lower Tam Giang stagnant water (type 2) expanded to the whole basin.

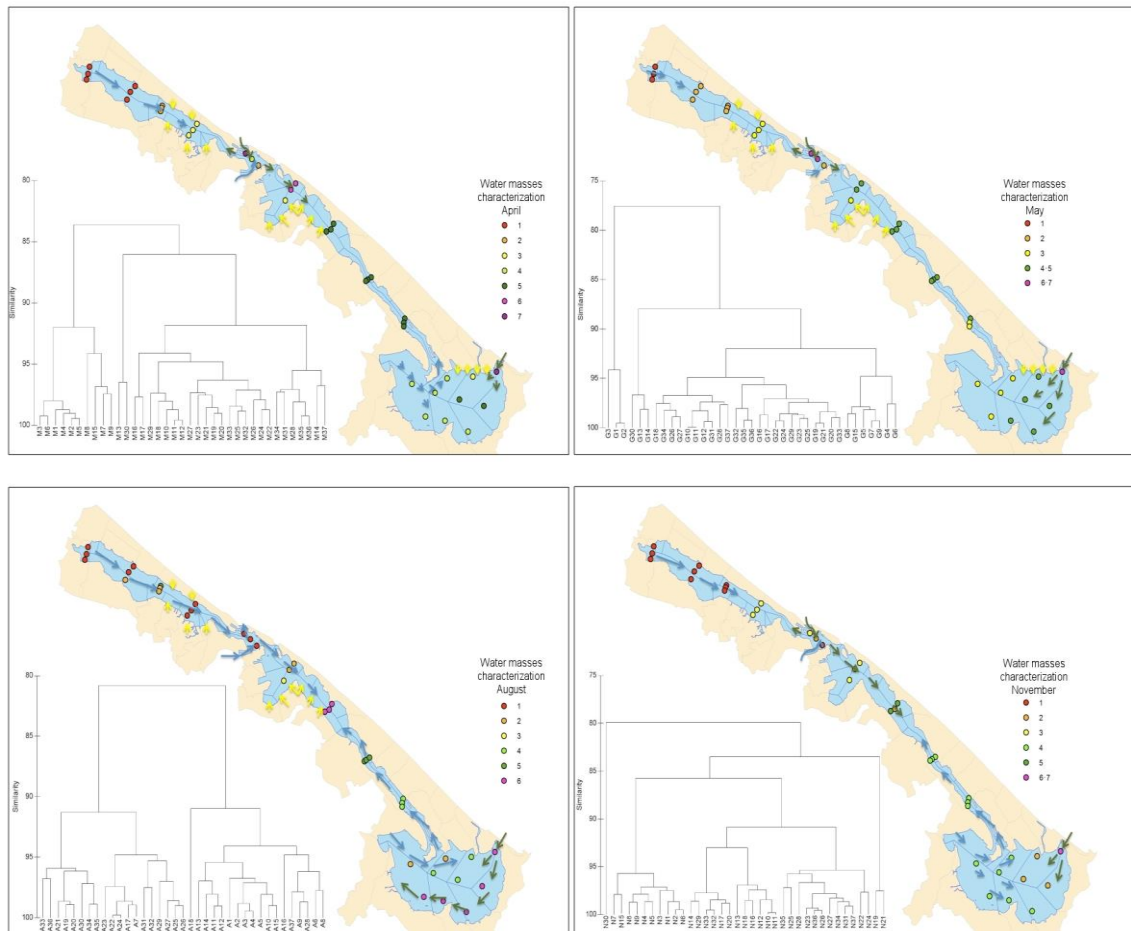
August 2006 was a month with enhanced precipitation (approximately 500 mm, three times the average of the month) and in that month fluvial waters (type 1) prevailed throughout the basin, with pockets of relic waters trapped locally. In November instead, three times dryer than average, signs of circulation restriction (type 3 waters) and reduced ventilation developed upstream from the Huong Phong narrows.

*Huong river mouth and Thuan An inlet.*

This area shows high variability of water characteristics, as it should be expected from mixing of riverine and marine flows. Marine saline waters (type 6 and 7) are detected in the vicinity of the inlet in the fair-weather months of April and May, diverted north towards Tam Giang, while riverine waters (type 1) predominate in the delta and expand, in the rainy season.

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<sup>4</sup> Cluster analysis is an exploratory data analysis tool for solving classification problems. Its object is to sort cases (people, things, events, etc) into groups, or clusters, so that the degree of association is strong between members of the same cluster and weak between members of different clusters.



**Figure 27 - Results of cluster analysis of environmental parameter (dissolved oxygen, pH, salinity and temperature) for the four surveyed seasons of 2006 (April, spring; May, dry season; August, beginning of rainy season; November, rainy season but unusually dry in 2006). Cluster description is: 1, fresh water characterized by very low salinity (1 psu), lowest pH (except November), highest DO in early spring and fall and lowest, in summer (May and August); 2, low-salinity (average 7.8 psu) inner-lagoon water, as low as 3.5 psu during rainy months of April and August and as much as 13 psu in dry months of May and November; pH fluctuates between 7.5 during rainy months and 8.1 during dry month; 3, low-salinity brackish water, with intermediate salinity values ranging from 10 and 19 psu, lowest dissolved oxygen concentration and pH higher than 8; 4, low-salinity brackish waters, with stable salinity of 14 psu, temperature above 30 degree, pH average around 8 and concentration of dissolved oxygen systematically higher than cluster 3 (in excess of 7 mg/l). During the driest month of May, these intermediate waters are indistinguishable from those of cluster 5; 5, typical brackish water with salinity ranging between 16 and 18 psu, temperature normally higher than 30 degrees and average pH around 8. The concentration of dissolved oxygen is higher than 7 mg/l and salinity raised over 22 psu during driest month of May; 6, cooler marine waters (average 26.5 °C in April; lower than cluster 6); 7, marine waters, with salinity higher than 20 psu, as much as 30 psu during the dry months of May and November; temperature between 28,5 and 30,5 °C. Lowest concentration of dissolved oxygen in November, similarly to cluster 3.**

*Thuy Tu channel and Cau Hai.*

These two sub-basins have somehow affine water characteristics through all seasons, indicating limited mixing with southward-flowing waters from the north. The influence of marine waters from Thuan An (type 6), in April and of riverine waters from the Huong river (type 2), in August and November, is recognizable in the Thuy Tu channel as far south as the Phu Xuan-Vinh Xuan narrows. Characteristic Cau Hai restricted waters (type 4 and 5) prevail and may expand upstream through the southern Thuy Tu, eventually trapping relic saline waters (type 6) in the mid-section of the basin (*e.g.* August).

Seasonality of the Cau Hai basin shows a localized marine influence in the vicinity of the Tu Hien inlet during the spring months of April and May (type 5-7) and the development of restricted-circulation water masses elsewhere in the sub-basin, during the same dry months (type 3-5). In August, eastward outflow of fresh waters from the Dai Giang, Thieu-Hoa and Truoi rivers towards Tu Hien drags marine waters from the inlet across the southern Cau Hai into a clockwise gyre that brings brackish waters and ventilation to the easternmost restricted sector. Circulation restriction tends to prevail during fair weather (November 2006).

*Sam Chuon lagoon.*

The Sam Chuon basin is an embayment within the Huong river delta system, artificially cut off from riverine influx. All south-flowing Huong river branches are presently dammed off; marine waters from Thuan An hardly penetrate into the Thuy Tu channel, without being diverted into the sub-basin, thus making the marine influence negligible. For these reasons and because of the intense development of aquaculture, the Sam Chuon waters show little seasonality and marked affinities (type 2) with those typical of other low dynamic, scarcely ventilated and highly cultured basins (*e.g.* vicinities of Vinh Hien, Hai Duong and lower Tam Giang).

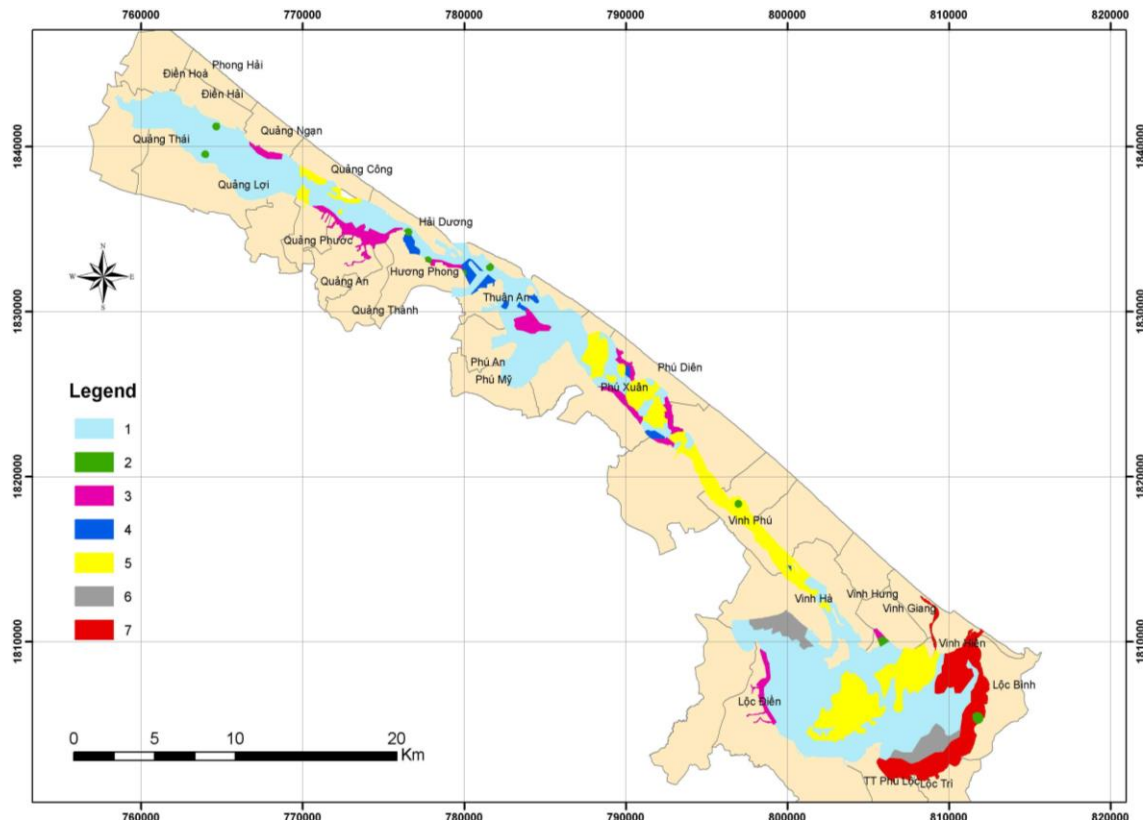


Figure 28 - Natural and anthropic habitat distribution of Tam Giang-Cau Hai. Legend: 1, hydrophytes prairies; 2, sea grass fields; 3, aquaculture ponds; 4, tidal flats; 5, sandy-mud lagoon floor; 6, muddy lagoon floor; 7, meadows flooded in the rainy season.

### Tam Giang-Cau Hai lagoon biocenotic mapping and distribution of main commercial species

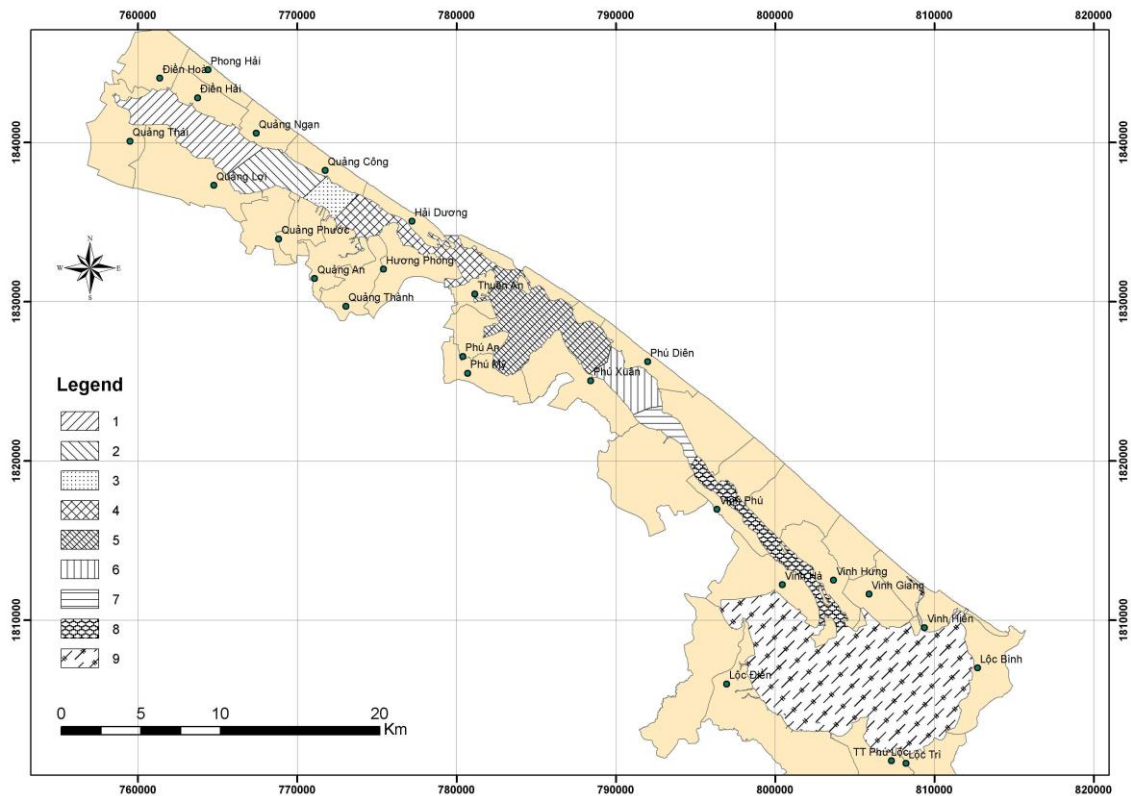
The distribution of lagoon biocenosis<sup>5</sup> is mainly driven by the salinity and its seasonal fluctuations. Figures 14 and 15 show the distribution of a suite of main *habitats* and commercial species in the lagoon, which reflects the physico-chemical characteristics of the water body and can thus be complementary elements of ecosystem definition.

Sea-bottom morphology, sediment composition, biota, macro-phytobenthos and macro-zoobenthos and vegetation coverage are parameters used in the zoning. The ecosystem of the Tam Giang-Cau Hai lagoon will be defined under the FAO code of ecosystem classification for further reference.

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<sup>5</sup> A group of interacting living animals and/or plants that form a particular ecosystem.





**Figure 29 - Distribution map of characteristic Tam Giang-Cau Hai biota and selected commercial species (natural fish population, macrophytobenthos, macrozoobenthos). Legend: 1, *Meretrix sp.* (*Trià*), fresh-water shrimps, fresh-water weeds; 2, *Meretrix sp.* (*Trià*), sea crab, thorned black crab (*ram*), *Potunus sp.* (blue crab or *ghe*); 3, *Saccostrea sp.*, *Meretrix sp.*, *Cerithidea*; 4, *Meretrix sp.*, *Potunus sp.* (blue crab or *ghe*), fresh-water *Tilapia*, *Siganus*, sea fishes; 5, *Meretrix sp.*, *Potunus sp.* (blue crab or *ghe*), seagrass, *Veneridae* and others bivalves, *Corbicula*, *Siganus*, *Metapenaeus ensis*, shrimps (*tôm go mo*); 6, *Cerithidae* (*ôc*), seaweeds; 7, *Cerithidae* (*ôc*), *Corbicula*, seaweeds; 8, *Solen sp.*, *Metapenaeus sp.*, *Cerithidae*; 9, *Corbicula*, *Portunus* (blue crab or *ghe*), *Pteria*, *Saccostrea*, *Penaeidae*.**

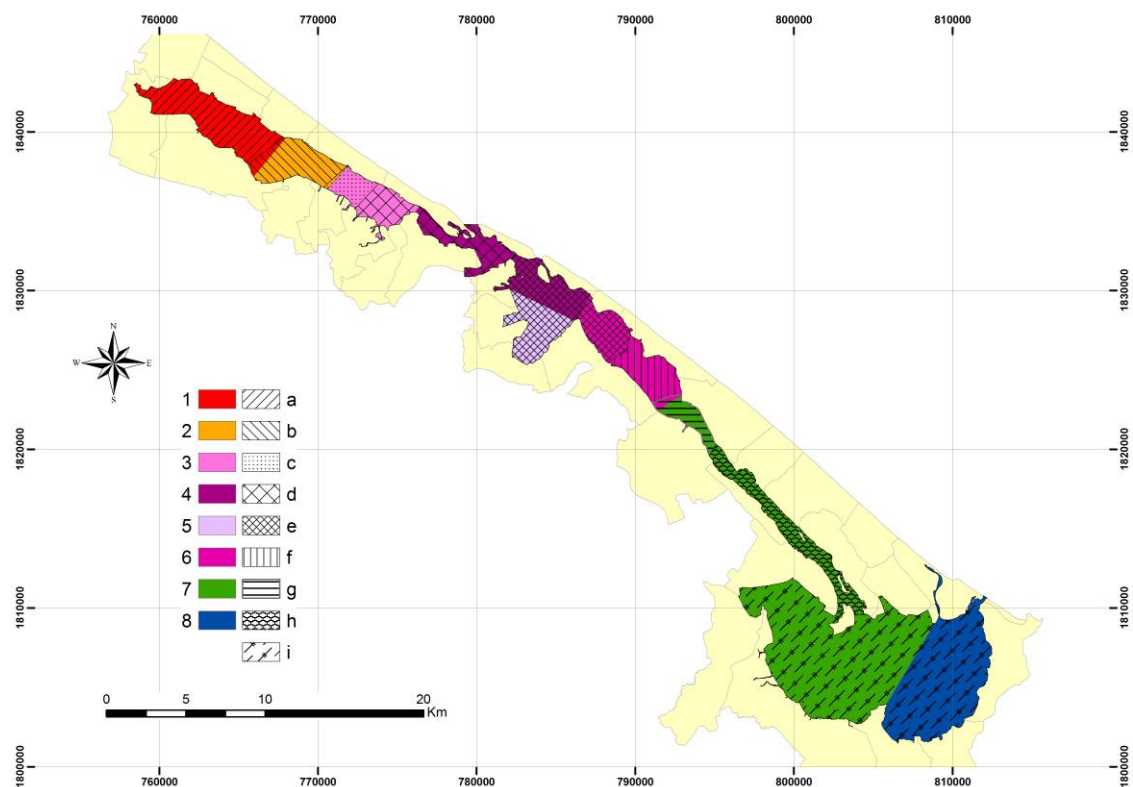
#### Tam Giang-Cau Hai lagoon ecosystem zoning.

A stack of the seasonal cluster maps integrated with information from biota distribution, lagoon-bottom features and sediment types provide a basis to ecosystem characterization and zoning.

*O’Lau ecozone.* Riverine freshwater body from O’ Lao predominates in the upper Tam Giang (O’ Lau eco-zone). The boundary of this area fluctuates seasonally, but freshwater generally predominates far south as the Quang Loi-Quan Ngan transect. Hydrophytes beds and fresh-water weeds characterize the lagoon bottom in this compartment, hosting a faunal association featuring fresh-water shrimps and the clam *Meretrix sp.* as the predominant species.

*Distal O' Lau ecozone.*

The quadrangle comprising the coastline of the Sia territory, of Quang Loi and Quang Ngan communes is characterized by transitional brackish waters (distal O' Lao ecozone). The bottom is persistently colonized by patchy schyzohaline hydrophytes prairies hosting *Meretrix sp.* clams, but a more prominently brackish assemblage characterized by a variety of crustaceans (sea crab, blue crab and black-thorned crab) replacing freshwater shrimps.



**Figure 30 - Integrated ecosystem zoning map derived from the distribution of water body characteristics (cluster analysis), lagoon-bottom features, sediment types and selected biota compositions (legend for fields a-I, see Fig. 29). Ecosystem legend: 1, Upper Tam Giang, O' Lau eco-zone; 2, Central Tam Giang, distal O' Lau eco-zone; 3, Lower Tam Giang eco-zone; 4, Huong river inner delta eco-zone; 5, Sam Chuon eco-zone; 6, Upper Thuy Tu marine relic waters; 7, Lower Thuy Tu channel and western Cau Hai continental domain; 8, Eastern Cau Hai marine and submarine domain, including mixing zone. The lower Tam Giang, the Sam Chuon and eastern Cau Hai show affinities in the way of having low-dynamic, stagnant water masses, largely of continental origin**

*Lower Tam Giang ecozone.*

The lower Tam Giang is still characterized by riverine predominance, with stronger seasonal marine influence and restricted circulation hindered by narrow passageways (e.g. Huong Phong). The influences of water masses alternatively flowing in opposite directions and the overwhelming pollutant load by aquaculture are characterizing features of the locale. Hydrophytes prairies host in this sector macro-benthic assemblages adapted to sandy bottoms and brackish-water conditions (e.g. *Meretrix sp.* associated with Cerithidea and oysters of the genus *Saccostrea*; fields 3 and 4 in Fig. 30).

#### *Huong river delta ecozone.*

The Huong river delta zone is unique in the sense that it is a lagoon compartment where marine and riverine water masses mix. Currents are enhanced by the complexities of the deltaic physiography, so that the prevalence of fluvial or marine regimes depends on seasonality and precipitations. Macrophyto- and macrozoo-benthos are characteristic of schyzohaline (fluctuating salinity) environment: sea fishes such as *Siganus* (rabbit fish) and sea crab such as *Potunus*, but also *Tilapia* are reported in this area, together with *Metapenaeus ensis* and other marine shrimp specimens.

The deltaic schyzohaline ecozone may seasonally extend southward into the upper Thuy Tu (Huong river sub-deltaic ecozone), still characterized by marine assemblages of the kind described above, together with the occurrence of other typical estuarine mollusks (e.g. *Corbicula* and Veneridae).

#### *Sam Chuon ecozone.*

The Sam Chuon embayment physically belongs to the Huong river delta complex, but hydrologically is a stand-alone compartment, isolated from both riverine and marine influx. Moreover, water masses are severely affected aquaculture effluents from the southern shores. The embayment, by biota and water characteristics, shows affinities with the lower Tam Giang.

#### *Upper Thuy Tu ecozone.*

The upper Thuy Tu is a hydrologic divide. The characteristics of this section are sandy bottoms, due to the sorting effect of currents, marine water seasonally filtering from Thuan An and interfering with episodic northerly counter flows from Cau Hai and a

peculiar marine biota consisting of sea weeds and Cerithidea mollusks. The effect of water masses converging towards this sector of Thuy Tu determines the encapsulation of pockets of modified marine relic waters, with anomalous concentration of organic pollutants from aquaculture effluents.

#### *Lower Thuy Tu ecozone.*

The lower Thuy Tu waters show similarities with those of western Cau Hai and an overall aquatic environment, set aside the morphology, comparable with that of the sub-basin. Both western Cau Hai and lower Thuy Tu waters have continental riverine affinity, suggesting seasonal penetration from Cau Hai into the channel, with some mixing with the modified marine waters of the upper Thuy Tu, somewhere in the mid-section at the height of Vinh Thanh-Vinh Phu and Vinh Xuan-Phu Da. The sandy-mud lagoon floor of the lower Thuy Tu is characterized by mollusks of Cerithidea family, specimens of the genus *Solen* and *Metapenaeus ensis*.

#### *Cau Hai ecozone.*

The Cau Hai sub-basin is divided in two parts: the low-energy, fluvial-dominated environment of the western and northern sector, with largely muddy bottoms, and the marine-dominated eastern and southern sector, with a transitional zone corresponding to the transect between Phu Loc town-Loc Tri and Vinh Hien communes. In the marine and transitional zones, sandy bottom prevails in the vicinities of the inlet (Tu Hien flood tidal delta) and in the Thuy Tu inner delta debouching into Cau Hai. The benthos characteristics are undifferentiated in the sub-basin (*Corbicula*, the oyster *Saccostrea* and the bivalve *Pteria*, along with sea crabs and Penaeidae).

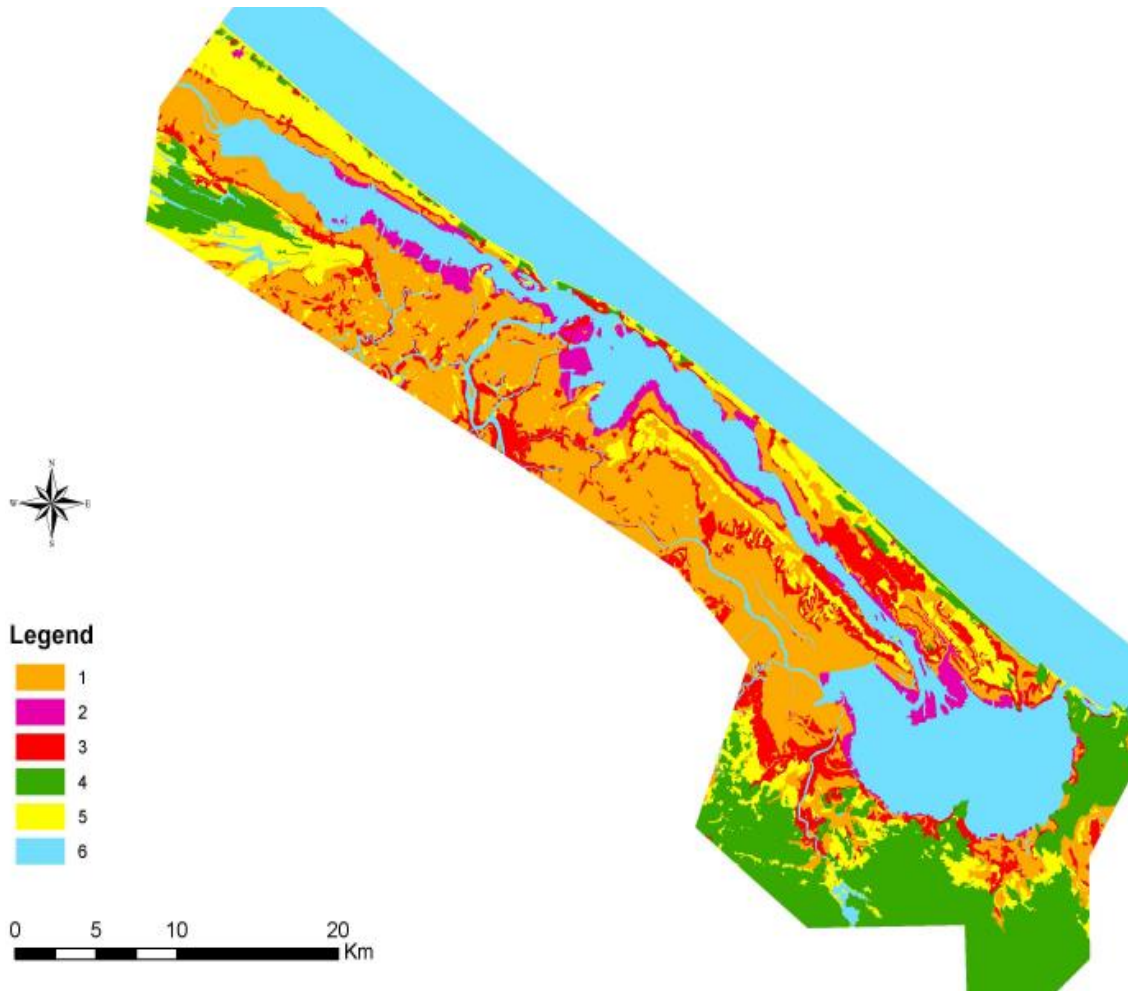
#### Non-living resources (soil, groundwater, mineral resources)

##### *Land use and land cover.*

To assist in management of Tam Giang-Cau Hai lagoon and its pertinences, a land use and land cover map 1:25,000 scale of the coastal Thua Thien Hue Province, covering 125,000 hectares and 34 coastal communes, was produced in 2006, stacking remote-sensing data from 1989, 2000 and 2006 imagery (Fig. 31).

An *ad hoc* legend was created by modifying and adapting to local characteristics, the CORINE<sup>6</sup> land cover legend.

In the detailed study and related map, the land use/land cover categories are grouped into four levels, with 5, 11, 23 and 8 headings each (Fig. 31).



**Figure 31 - Simplified land-use map of the territory surrounding the Tam Giang-Cau Hai lagoon. Legend: 1, agricultural areas (rice paddies and cultivated fields) in alluvial plain lowland; 2, aquaculture areas, potential sites of effluent waste-water discharge and sources of organic pollutants (fish feeds, fish metabolism residues etc.); 3, urban (and industrial) areas; 4, semi-natural tropical and subtropical moist broadleaf forests; 5, semi-natural sandy area, with irregular and discontinuous bush coverage; 6, inland waters and sea.**

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<sup>6</sup> CORINE, “Coordination of Information on the Environment”. In 1985 the Corine programme was initiated in the European Union; it was a prototype project working on many different environmental issues. The Corine databases and several of its programmes have been taken over by the EEA. One of these is an inventory of land cover in 44 classes, and presented as a cartographic product, at a scale of 1:100 000. This database is operationally available for most areas of Europe (<http://www.eea.europa.eu/publications/COR0-landcover>)

1 Artificial Surfaces	11 Urban Fabric	111 Continuous urban fabric		
		112 Discontinuous urban fabric		
	12 Industrial, Commercial and Transport Units	121 Industrial areas		
		122 Ports and airports		
		123 Aquaculture ponds	1231 Inland aquaculture ponds 1232 Lagoon aquaculture ponds	
	13 Mines and construction sites	131 Mineral extraction sites		
		132 Construction sites		
	14 Graves or cemeteries, monumental areas	141 Graves or cemeteries		
2 Agricultural areas	21 Arable land	211 Rice fields		
		212 Complex cultivation		
	22 Agro-forestry areas	221 Agro-forestry areas		
3 Forests and semi-natural areas	31 Forests	311 Broad-leaved forest	3111 Broad-leaved sparse forest 3112 Broad-leaved dense forest	
		312 Coniferous forest	3121 Coniferous sparse forest 3122 Coniferous dense forest	
		313 Mixed forest	3131 Mixed sparse forest 3132 Mixed dense forest	
		314 Mangrove forest		
		32 Open spaces with little or no vegetation	321 Shrub, bushes and/or herbaceous vegetation 322 Sandy inner areas 323 Sandy coastal areas	
	4 Wetlands	41 Inland wetlands	411 Inland wetlands	
	5 Water bodies	51 Inland waters	511 Water courses	
			512 Lakes or inner basins	
52 Marine waters		521 Coastal Lagoon		
		522 Sea and Ocean		

**Figure 32 - Land use/land cover categories and nomenclature defined for coastal Thua Thien Hue Province, modified after CORINE land types classifications.**

*Mineral resources and building materials.*

Exploitation of mineral resources in the floodplain/coastal zone relate to:

- Sand mining from inlets and active beaches, for construction (artisanal, manually operated)
- Sand mining from active river bed for construction and infrastructure building (artisanal and semi-industrial, manually and machine-operated)
- Laterite-soil quarrying from slopes (semi-industrial).
- Limestone rock mining for handycraft (artisanal) and cement processing (industrial).
- Sand is actively mined from inlet and beaches. Excess sediment in these areas of natural natural deposition is not harmful but rather beneficial to the hydraulic system which suffers excessive siltation and clogging.

Sand withdrawal from active rivers is intense, semi-industrially operated and continuous through the years, except during floods and storms. This activity driven by strong and likely higher future demand as a result of urbanization, industrialization and infrastructure building is harmful to the sediment budget along the coast and will likely result in enhanced coastal erosion, already severe at places.

Laterite-soil quarrying from mountainous slope is an important source of low-quality material for roads substrates, earthfill and basement for country and urban houses. This practice has been widespread in recent years along the slopes surrounding Cau Hai, next to the road for immediate and convenient supply. Laterite is abundant as a tropical soil wherever outcrop of crystalline basement rock is outcropping. Quarrying surveying and technology is rather primitive: deforestation is followed by bulldozing of soft argillaceous soil to meet the immediate demand, until the rocky substrate or unsuitable material are encountered. Transportation is by small trucks or dumpers, according to the needs of the contractor. When finished, the mining site is abandoned, vulnerable to slope instability, creep, landsliding and dispersal of contaminant laterite sludge in the lagoon.

Limestone rock, wherever of suitable mineralogical composition and purity, is mined for processing in cement industry, more often inland rather than in coastal areas. Decorative limestone is also small-scale quarried and marketed across Vietnam for household handicraft, decorations, column pedestals and tombstones. While the former is highly impacting the environment and landscape integrity, this latter is rather a popular artisanal activity in Thua Thien Hue as well as in other areas of Vietnam, more of a cultural value and should be subject of tutelage.

#### *Groundwater.*

Coastal Thua Thien Hue Province possesses extensive superficial aquifers<sup>7</sup> that, once managed, can provide adequate freshwater supply for agriculture, aquaculture and urban services of rural communities. Ground water serves the great majority of people who live in rural areas; the reason is that, among the various sources of supply, ground water is by far the most practical and safe in nature.

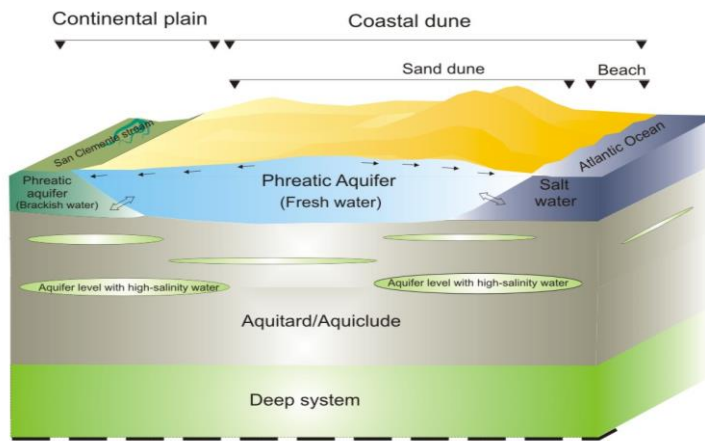
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<sup>7</sup> Water-bearing rock or sediment formations

Unlike surface waters, the advantages of groundwater are:

- it is likely to be free of pathogenic bacteria;
- generally, it may be used without further treatment;
- in many instances it can be found in the close vicinity of rural communities;
- it is often most practical and economical to obtain and distribute;
- the water-bearing aquifer from which it is drawn usually provides a natural storage at the point of intake<sup>8</sup>.

There are some disadvantages relating to its high mineral content and to the investment needed to survey, engineer and capture water from aquifers, but advantages far outnumber the few inconveniences that exist over surface, more erratic supply.



**Figure 33 - Cartoon showing a typical example of coastal phreatic aquifer from sand dunes.**

In coastal Thua Thien Hue, groundwater resides near-surface (the free-water zone of Tollman) in the modern coastal-dunes aquifers (the backshore from Phong Dien to Hai Duong and from Thuan An to the extreme south) and sub-recent dune fields of Quang Dien, Phong Dien and Phu Vang (Fig. 1 and 31).

Aquifers are easily recharged by the abundant precipitation of the province (over 3,000 mm/year) and freshwater is there available, in most instances, in the close vicinity of sites of utilization and in amounts sufficient to the community requirements.

The resource has not yet been seriously considered for potential, quality and vulnerability. Pollution is currently a severe hazard for the resource, from urban waste and industrial aquaculture recently developed on sand fields. Once polluted, aquifers are difficult to treat or remedy.

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<sup>8</sup> World Health Organization Monograph 42, Chapter 3



## **Summary and conclusions**

### **Assessment of status of coastal environment**

#### Status of quality of coastal environment

The Tam Giang-Cau Hai and vicinities maintain its rather pristine state compared to other lagoon and deltaic settings of the world, despite the high population density, the strong demand for food resources and on going vigorous development. Historical events of the second half of the past century slowed down the process of industrialization and urbanization, maintaining agriculture and fishery into artisanal conditions and the aesthetic of the natural environment into its untouched beauty.

Infrastructure development (road, bridges, expansion of cities) generated impacts: on landscape, on resources and, easing communications among villages and to the city, on the social structure itself. Some traditional occupations like local ferries, handicraft making etc. have vanished and pressure has been transferred from the aquatic environment on land, from the peripheral centers to cities. Traditional nomadic boat-dwellers, once present in dense and bursting floating villages in both the remote lagoon expanses and in the cities have been, in the turn of a quinquennium, relocated on land, with doubtful improvements in their lifestyles, livelihoods and aquatic environment but with an undoubted loss of cultural and traditional value.

Aquaculture has been another major element of perturbation of the natural system: the attempt to promote shrimp production in the nineties generated in cascade negative impacts whose repercussions are still felt today: widespread organic pollution, wiping out of endemic mangrove ecosystem, eliminating inter-subtidal spawning and nursery platform, loss of productivity, break of reproductive cycle of numerous species, loss of biodiversity, hindrance of circulation etc.

Construction and uncontrolled mining of building materials altered the harmony of the landscape by producing large red scars in the deep green of the forest: laterite mining in near the coastline deteriorated the quality of the aquatic environment by promoting the flushing of red sludge in the lagoon and increase the amount of suspended matter; uncontrolled quarrying is promoting soil erosion, land sliding and slope instability. The uncontrolled dumping of construction material or roadwork debris (e.g. construction

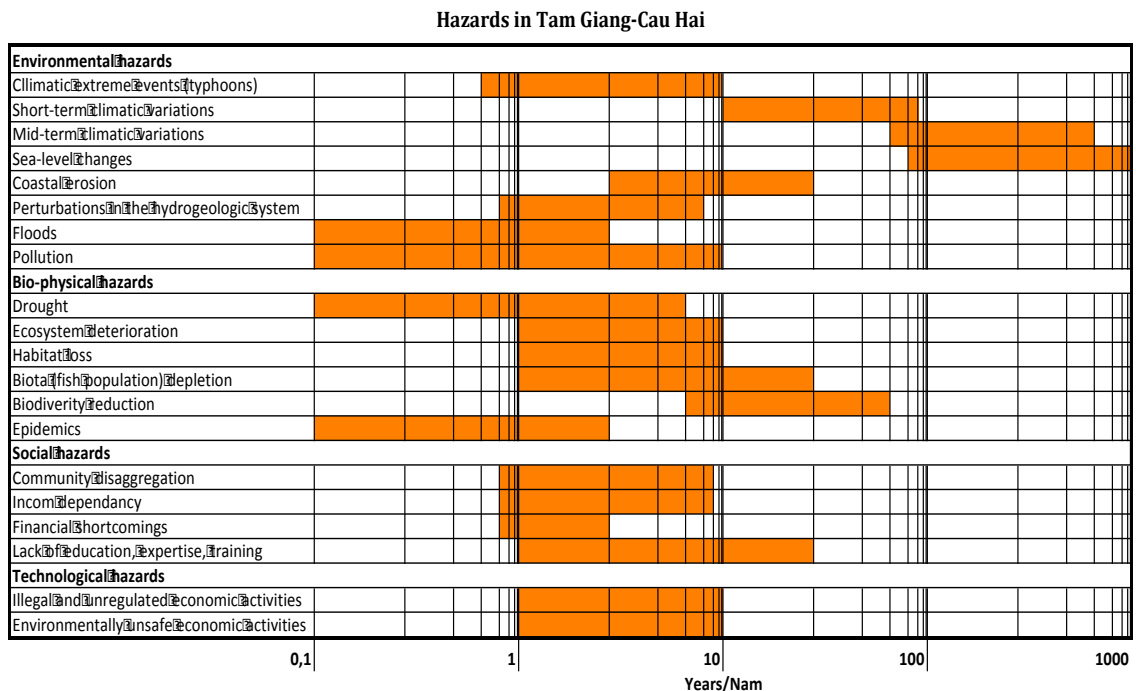
residues of the Tu Hien bridge) also deteriorated the integrity of the coastline and lagoon floor, to the detriment of fishery and living creatures.

River diversion, damming and construction of salinization-prevention dikes, besides resolving some of the impending emergencies for agriculture altered the natural waterways network and hydraulics, generating changes in the lagoon biological cycles.

Damming in the upper catchment, by reducing the discharge of rivers and sediment-laden water will further promote saline intrusion from the sea and sand mining for construction from the all active riverbeds will increase coastal erosion, which already became severe at places in the past few years (e.g. Thuan An and Hai Duong). The construction of shoreline groins and piers, neither resolve the problem nor mitigate the process (e.g. Cua Viet and Cua Tung, Quang Tri).

Despite rapid and recent changes, the environment maintains its charm and beauty, its unique character in the whole Vietnam and this is a cultural asset that should be maintained for its recreational value and use for the benefit of the forthcoming development of the touristic industry in the country.

The sources of environmental hazards, pollution and degradation in the coastal area



**Figure 34 - Hazards classification for the Tam Giang-Cau Hai and the coastal province of Thua Thien Hue. The time scale indicates the time frame and frequency at which hazard may turn into an emergency.**

Hazards<sup>9</sup> for the lagoon environment listed in Figure 20 have a natural, climatic and social origin. These can be classified as environmental, when they relate to the physical environment; bio-physical, when they affect the biota and living creatures in general; social, when hazard may impact the socio-economy of the human community at large and technological, when relate to technologically assisted human activities.

The timing and the frequency (return period<sup>10</sup>) at which a hazard may turn into an emergency may assist decision makers in designing interventions and setting priorities. Prevalent natural hazards in Tam Giang-Cau Hai lagoon and the costal area of Thua Thien Hue include pollutant dispersal from agriculture, aquaculture, industry and urban settlements; climatic hazards relate to seasonal fluctuation of river discharge (flash-floods) and the recurrence of autumnal typhoons that, if in-phase with the flash floods, can enhance the level of risk. The simplified land-use map of Fig. 31 shows the potential sources of risks.

In terms of susceptibility towards hazards and vulnerability, three levels of risk can be identified for the different zones of Tam Giang-Cau Hai: *high risk*, whenever incumbent hazards are persistent in populated areas or production sites, emergencies are frequent and/or multiple emergencies can develop simultaneously (e.g. flood and typhoon), with effects that manifest in the immediate or short term; *moderate risk*, whenever emergencies, once they manifest, are localized or controllable, affect lesser-densely populated areas and *low risk*, whenever emergencies are mild and their effects are likely to be naturally mitigated in the short term. High-risk zones require immediate attention and hazard mitigation interventions. Moderate risk zones require monitoring and rehabilitation measures. Low-risk zones require low-level of attention. With respect to these three levels of risks, the Tam Giang-Cau Hai system can be categorized has follows.

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<sup>9</sup> Hazard/emergency ranking: establishing priorities. A hazard is a situation that poses a level of threat to life, health, property or environment. Most hazards are dormant or potential: the likelihood a hazard turns into an emergency and the seriousness of the emergency whenever it manifests express the level of risk. Emergencies are episodic, cyclic, or persistent. The time frame into which the emergency develops bears consequences on establishing priorities of intervention.

<sup>10</sup> A return period also known as a recurrence interval is an estimate of the likelihood of an event, such as an earthquake, flood or a river discharge flow to occur. It is a statistical measurement typically based on historic data denoting the average recurrence interval over an extended period of time, and is usually used for risk analysis (e.g. to decide whether a project should be allowed to go forward in a zone of a certain risk, or to design structures to withstand an event with a certain return period. The following analysis assumes that the probability of the event occurring does not vary over time and is independent of past events.

### Critical issues relating to the quality of wetland habitats of Central Vietnam

- Parameters and variables of water quality, sediments, benthic associations in Tam Giang-Cau Hai lagoon and estuaries of Central Vietnam portray a rather stable situation and a persistent acceptable standard for aquaculture; however, high organic-matter input to open waters relating to biological excretion and excessive feed may create eutrophication and anoxia lethal for life.
- High-impact shrimp monoculture is unsustainable on the long term: alternative methods (e.g. clam farming) need to be studied and piloted to mitigate negative effects of effluents and aquaculture waste waters.
- Livelihood shift is necessary, having short-term impact on household welfare and economy, but unquestionably beneficial on the long term.
- Need of a viable community-based solid-waste collection and treatment plan, including the creation of a community-based collection network at commune level.
- Appropriateness of the future implementation of a wetland ecosystem carrying capacity exercise, in support of future planning and zoning for conservation and exclusion
- Need of a water-quality monitoring system for early warning of upcoming critical conditions and implementation of protection measures.

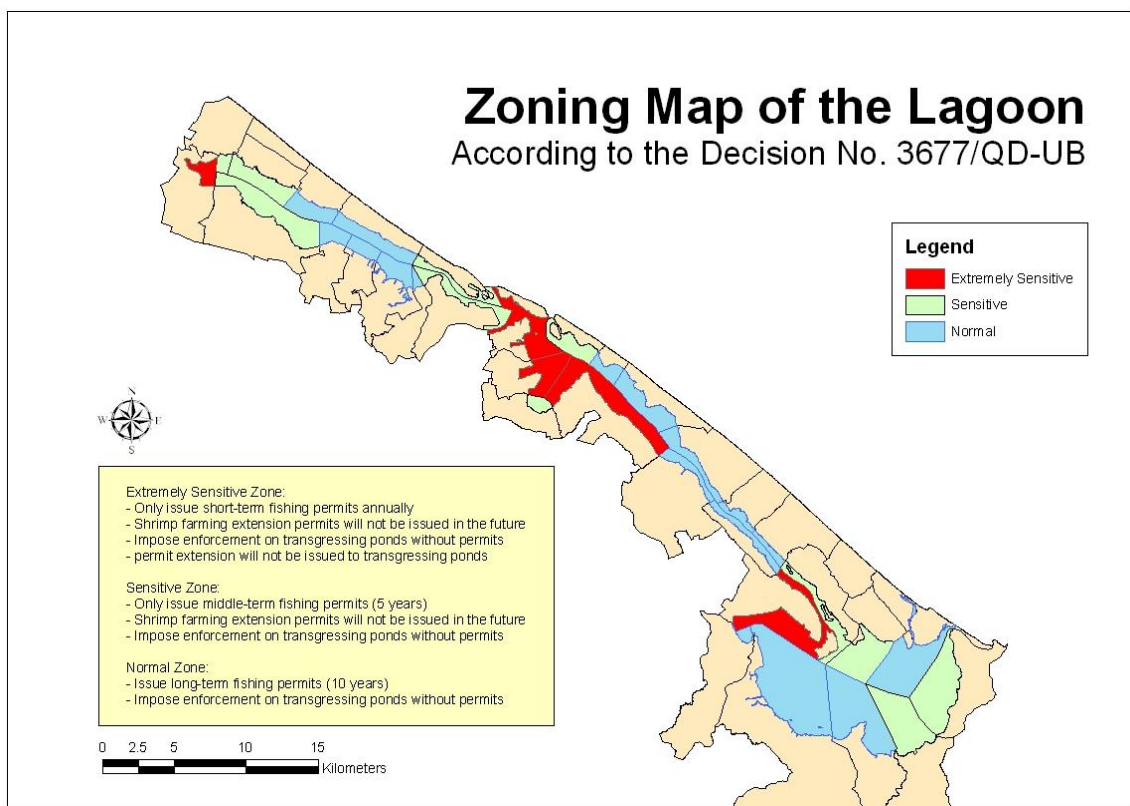
### Vulnerable ecosystems in Tam Giang-Cau Hai lagoon and coastal areas and risk assessment maps.

The sub-basins of Lower Tam Giang, Sam Chuon-upper Thuy Tu, and western Cau Hai, because of physiographic complexities and hydrological hindrance, are at present considered as high-risk zones. The intense development of aquaculture and related threats of pollution, disease outbreaks, loss of crop, income instability, etc. represent the main hazards and source of social hardship: people, their livelihoods and the environment are affected.

Concerning climatic hazards, the landward side of the lower Tam Giang coastal plain (Huong Tra and Quang Dien) is subject to poor drainage and subject to persistent floods yearly. Organic pollution is severe due to enormous development of ponds and lack of flushing. The central region from the lower Tam Giang (Hai Duong Commune) to the upper Thuy Tu (Thuan An, Phu Hai) appears to be under potential threat of coastal erosion: these zones should be reclassified as extremely sensitive, according to the

definition adopted in the Decision No. 3677 of the Thua Thien Hue People's Committee (Fig. 35). Thuy Tu is currently highly polluted and priority should be given to effluent control from aquaculture. Western Cau Hai should be eligible of high sensitiveness given its water exchange limitations

The eastern Cau Hai, the lower and middle Thuy Tu and the upper Tam Giang are classified as moderate-risk zones (sensitive zones in the terminology of the Decision No. 3677 of the Thua Thien Hue People's Committee) implying that better fishery-management practices and rehabilitation should suffice to mitigate the current state of degradation. The Tu Hien inlet is a critical zone for sea-lagoon interaction.



**Figure 35 - Zoning map of the Tam Giang-Cau Hai lagoon based on risk-level towards fishery activities, as assessed in 2004, and range of proposed interventions in support of the sector (as per Decision No. 3677/QD-UB of 2004 of the Thua Thien Hue Provincial People's Committee).**

No zones may be classified at present as low-risk, even though the Huong river delta and the upper Tam Giang (O'Lau river mouth), because of ecosystem integrity may be re-proposed for conservation and tourism. Chemical pollutants from agriculture are seasonally severe, so that better control on effluents and ban of POPs use should be emplaced.

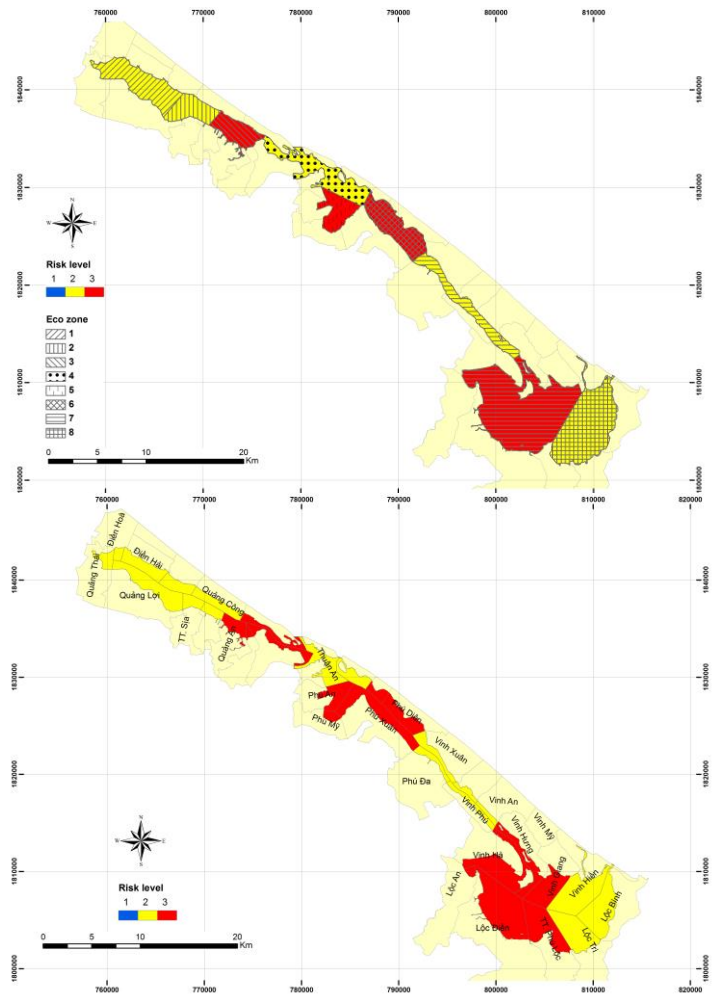


Figure 36 - Risk maps of the Tam Giang-Cau Hai lagoon representing the risk level evaluated for each of the identified ecosystems (above) and communes (below). Risk levels have been ranked according to three categories: 1, *low*, whenever emergencies are mild and their effects are likely to be naturally mitigated or nullified in the short term; 2, *moderate*, whenever emergencies, once they manifest, are circumscribed or controllable and 3, *high*, whenever emergencies are persistent and/or multiple emergencies can manifest simultaneously.

Based on latest assessments, the 2004 commune risk-level (sensitiveness) map of Tam Giang-Cau Hai should be revised according to the representation of Fig. 36

Assessment of status of coastal biological resources

Biodiversity is an asset of Tam Giang-Cau Hai. In terms of species richness, the lagoon and wetlands of Thua Thien Hue far outnumber any other lagoon in Vietnam. Count totals approximately 1000 species, most of them systematically identified: phytoplankton records the highest in number of representative species (287), fish's species are 215-230; birds, 73; zooplankton, 72; zoobenthos, 193 species, seaweeds 46, higher plant, 31; water grass 18 (of which 7 species of sea grass). Percent of specie distribution is shown in Fig. 37.

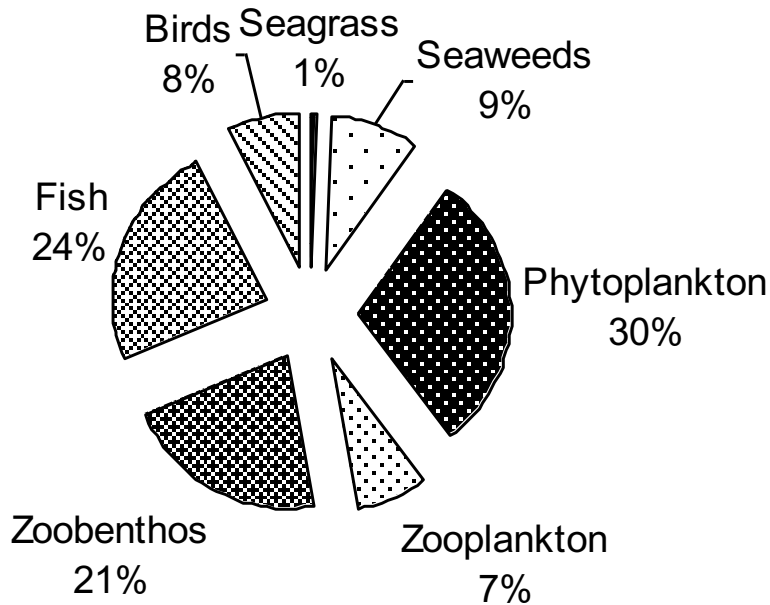


Figure 37 - Percent of species distribution in Tam Giang-Cau Hai lagoon identified during the assessments 2006-2018.

High biodiversity depends on complexities of lagoon physiography and habitats, and seasonal changes of the aquatic environment: salinity fluctuations between the dry and rainy season affect the composition of biota and species predominance. For example, in rainy season, the freshwater grass *Valisneria spiralis* is predominant, associated to the freshwater prawn *Macrobrachium sp.*, the freshwater shrimp *Palaemonetes sp.* and the clam *Corbicula*. In contrast, in the dry season, brackish species predominate, such as squids *Loligo sp.* and *Sepia sp.*, the shrimp *Penaeus sp.*, the crab *Portunus sp.*, and the fish *Siganus sp.*

A characteristic of the Tam Giang-Cau Hai lagoon is the growth of underwater and coastal prairies, ideal habitat of migrant waterfowls, including approximately thirty protected species among which the purple heron (*Ardea purpurea*), the osprey (*Pandion haliaetus*), the common kestrel (*Falco tinnunculus*) and one species, the asian dowitcher (*Limnodromus semipalmatus*), which is recorded in the Vietnam Red Book of endangered species.

There are over a hundred aquatic species of economic interest in Tam Giang-Cau Hai lagoon. From crude estimates, local fishers in the five lagoon districts produce approximately 2,500 metric tons of aquatic products (excluding seaweeds of low economic value), primarily for the local market. Among these, there were 23 marketable fish species and 34 other categories including benthic edible organisms that are considered a valuable commodity for processing of local food such as sour shrimp sauce (*tom chua*) and clam rice (*com hen*).

Among the 73 species of aquatic bird, there are five migrant types seasonally resident in the lagoon: the Eurasian coot (*Fulica atra*), the spot-billed duck (*Anas poecilorhyncha*), the graylag goose (*Anser anser*), the spotted redshank (*Tringa erythropus*), the garganey (*Anas querquedula*), with estimated several ten thousands of individuals of commercial interest.

Seagrass and seaweeds are harvested for manure and animal feed, to increase the income for local households. Surveys revealed that there are about 1021 hectares a seagrass fields in Tam Giang-Cau Hai, mostly concentrated in Con Noi, Con Dai and Ba Con (Cau Hai).

Subaqueous prairies develop at depth of 0.7 to 1.4 m, displaying different species composition depending seasons and salinity<sup>11</sup>, with massive growth of invasive freshwater plants during the rainy season and increased runoff; with increased salinity during the summer months, seagrass replaces freshwater plants and the decay of dead tissue contributes to water pollution and alkalinity.

The growth of invasive freshwater plants is a serious, yet common problem of coastal lagoons subject to fluvial water predominance and limited seawater circulation. Freshwater plants are often classified as pest plants, highly noxious to environment and difficult to eradicate. *Hydrilla* for example, forms dense mats of vegetation that interferes with recreation and destroys fish and wildlife habitats.

Despite Tam Giang-Cau Hai has a long history of scientific investigations, quantitative studies on biological resources are still non exhaustive. The biological stock of tropical, multi-species systems is a laborious elaboration of catch monitoring data: under time and budget limitation, the IMOLA project attempted a crude estimate of biomass based on bulk catch in areas where bottom morphology and navigation conditions allowed trawling. The standing stock size was expeditiously obtained applying the Pauly (1983) formula<sup>12</sup>.

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<sup>11</sup> Five species belonging to Hydrocharitaceae, Cymodoceaceae, Zosteraceae and Ruppiaceae, with predominant *Zostera japonica*, *Halophila beccarii* and *Halodule pinifolia*. Species of oligohaline waters (0.5-4‰) are *Hydrilla verticillata* (a salinity resistant, potentially invasive freshwater plant), *Vallisneria spiralis* (eel grass, proliferates on organic-rich substrates, invasive pest plant), *Myriophyllum spicatum* (watermilfoil, potentially invasive, noxious weed), *Potamogeton crispus* (noxious weed of inland waters and lakes). Euryhaline species (tolerant to wide range of salinity) are *Zostera japonica* and *Halodule pinifolia*. Mixohaline species (0,5-30‰) are *Halophila beccarii* and *Ruppia maritima*. A predominant euhaline species (normal marine) is *Halophila ovalis*.

<sup>12</sup> Pauly, D., 1983, Some simple methods for the assessment of tropical fish stock, FAO Fishery Technical Paper 234



Thirteen trawling transects using push net in O Lau, Hai Duong-Thuan An, Sam Chuon, Thuy Tu and Cau Hai. Based on captured frequency, taxonomic categories were subdivided into:

- High-frequency group. 42 species of economic interest, 7 of which rather ubiquitous, with frequency of 78-100% of total catch sessions. These include flathead grey mullet (*Mugil cephalus*), whipfin silver biddy (*Gerres filamentosus*), *Triacanthus brevirostris*, *Hemirhamphus sinensis*, freckled hawkfish (*Ambassis kopsii*), orange-spotted and white-spotted rabbitfish (*Siganus guttatus* and *Siganus oramin*), and shrimp *Metapenaeus ensis*.
- Medium-frequency group: 10 species, with frequency from 46 - 70% of total catch sessions. These include crabs (*Varuna litterata*), catfish (*Leiocassis hanamensis*), *Sparus latus*, crescent bass (*Terapon jarbua*), bartail flathead (*Platycephalus indicus*), shortnose ponyfish (*Leiognathus brevirostris*), gobids (*Oxyurichthys tentacularis*), glass eel (*Pisodonophis boro*), Commerson's anchovie (*Stolephorus commersonii*).
- Low-frequency group: 11 species with appearance frequency of 25-40% of total catch sessions. These include blue swimmer crab (*Portunus pelagicus*), clams (*Corbicula sp.*), whitefin wolf-herring (*Chirocentrus nudus*), *Tetraodon ocellatus*, trumpeter whiting (*Sillago maculatus*), white goby (*Aboma lactipes*), *Clupanodon punctatus*, Mauritius sardinella (*Sardinella jussieu*), prawns (*Penaeus merguensis* and *Penaeus monodon*).
- Very low frequency group: 14 species recorded on only 1-2 catch sessions. These species have restricted distribution: for example the freshwater prawn *Macrobrachium sp* is an indicator of freshwater habitats of O Lau and adjacent areas whilst *Squilla sp.* is only found in saline waters close to inlets; around the lagoon mouths; The flat-head goby is a rare species only found in Sam Chuon and Huong Tra.

The highest recorded diversity were in the marine-influenced lagoon tract comprised between Quang Cong to Phu Hai (Tam Giang and Sam Chuon), with number of species ranging from 30 and 32. The lowest number of species was recorded in the freshwater-dominated section of O Lau (20 species), followed by Thuy Tu (23 species) and Cau Hai (26 species) (Fig. 38). Similarly, the highest bulk biomass, as calculated from catch, was recorded in the Sam Chuon-Thuy Tu transect and the lowest value in absolute

terms was in O Lau. Based on production during the catch sessions, the bulk spot biomass value was assessed for each location, applying the Pauly (1983) relationship (Table 7).

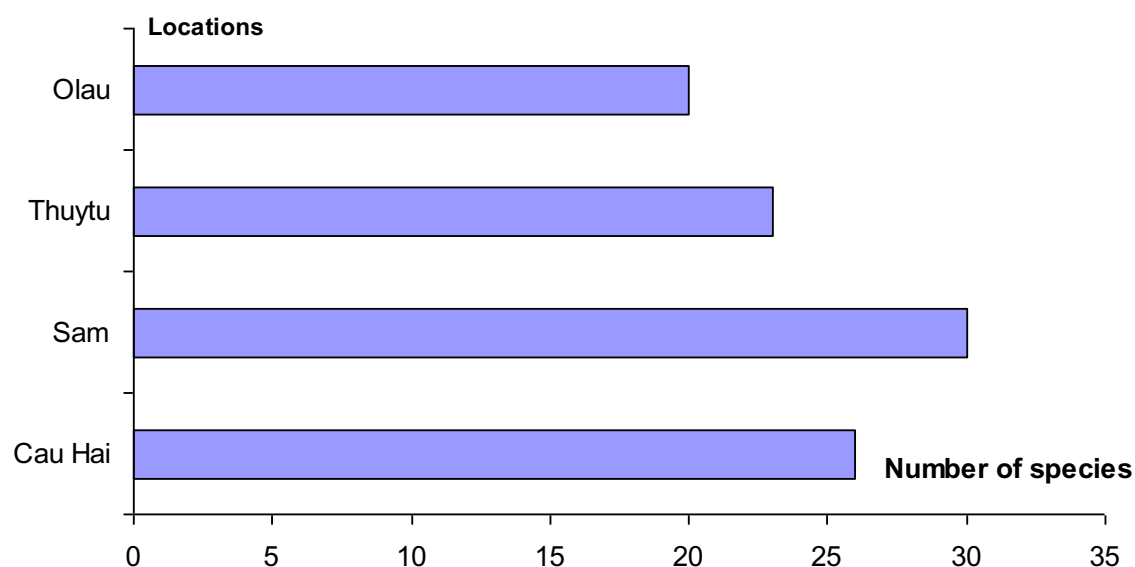


Figure 38 - Distribution of number of aquatic species in the Tam Giang-Cau Hai lagoon sub-basins.

Location	Production per catch session (tons/hectare)	Extrapolated bulk biomass (tons)
O Lau	3.293	13.172
South Thuan An	17.562	70.248
East Thuan An	5.193	20.772
West Thuan An	8.849	35.396
West Sam Chuon	4.837	19.348
North Sam Chuon	190.200	760.800
East Sam Chuon	36.997	147.988
Huong Tra	71.862	287.448
Thuy Tu	47.104	188.416
South Cau Hai	16.300	65.200
East Cau Hai	22.103	88.412
North Cau Hai	16.907	67.628
West Cau Hai	8.526	34.104

Table 7 - Production per catch session (in tons per hectare) and extrapolated bulk biomass (in tons), May 2006.

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Survey Institute of Marine Resources and Environment, April 2006 Phytoplankton Species

STATION 1			
Sampling date: 03/04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Melosira granulata	14	280
2	Leptocylindrus danicus	30	600
3	Navicula cyro	1	20
4	Cymbella sp.	1	20
5	Pleurosigma p	3	60
6	Nitzschia lorenziana	2	40
7	Pseudonitzschia p.2	2	40
8	Surirella tenera, reversa	4	80
9	Scenedesmus spp.	4	80
10	Euglena sp.1	3	60
11	Gloeotila pelagica	13	260
		77	1540

STATION 2			
Sampling date: 03/04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Melosira granulata	37	740
2	Leptocylindrus danicus	9	180
3	Chaetoceros socialis	8	160
4	Synedra aillonii	2	40
5	Diploneis mithii	1	20
6	Nitzschia lorenziana	3	60
7	Nitzschia longissima, reversa	2	40
8	Nitzschia sigma	3	60
9	Surirella tenera, reversa	2	40
10	Pediastrum sp.	1	20
11	Euglena sp.1	13	260
12	Phacus sp.1	4	80
13	Gloeotila pelagicum	73	1460
			160

STATION 3			
Sampling date: 04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Synedra aillonii	2	40
2	Nitzschia lorenziana	2	40
3	Nitzschia sigma	2	40
4	Nitzschia sp.	4	80
5	Surirella tenera, reversa	2	40
6	Euglena sp.1	2	40
7	Gloeotila pelagicum	6	120
			380

STATION 4			
Sampling date: 05/04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Diploneis mithii	7	70
2	Nitzschia lorenziana	31	310
3	Nitzschia longissima	13	130
4	Nitzschia longissima, reversa	128	1280
5	Nitzschia sp.	16	160
6	Bacillaria paxillifera	54	540
7	Pediastrum sp.	1	10
		255	2550

STATION 5			
Sampling date: 04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Melosira granulata	12	240
2	Diploneis mithii	4	80
3	Nitzschia lorenziana	11	220
4	Nitzschia longissima	21	420
5	Nitzschia longissima, reversa	140	2800
6	Nitzschia sigma	6	120
7	Nitzschia sp.	40	800
8	Bacillaria paxillifera	15	320
9	Gloeotila pelagicum	5	100
		255	5100

STATION 6			
Sampling date: 04/2006			
No	SPECIES	QUANTITY	CELLS/L

STATION 19			
Sampling date: 04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Thalassiosira spp.	2	20
2	Chaetoceros spp.	4	40
	Thalassionema nitzschoides		20
4	Synedra sp.	14	140
5	Grammatophora marina	55	550
7	Trachyneis aspera	2	20
8	Diploneis scutellum	1	10
	Gyosigma sp.		
10	Pleurosigma angulatum	1	10
11	Pleurosigma longatum	1	10
12	Pleurosigma pelagicum	1	10
13	Nitzschia longissima, reversa	3	30
14	Nitzschia sigma, intercedens	2	20
15	Nitzschia sp.1	4	40
	Nitzschia sp.		
17	Surirella ovalis	1	10
18	Dinophysis audata	9	90
19	Dinophysis ripos	2	20
20	Ceratium curca	1	10
21	Gonyaulax polygramma	2	20
22	Gonyaulax pinifera	1	10
23	Gonyaulax verior	1	10
24	Alexandrium spp.	1	10
25	Oscillatoria sp.1	249	2490
26	Oscillatoria sp.2	4	40
		373	3730

STATION 20			
Sampling date: 05/04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Thalassiosira spp.	10	200
2	Chaetoceros spp.	168	3360
3	Thalassionema nitzschoides	21	420
4	Thalassiothrix rauenfeldi	34	680
5	Synedra sp.	113	2260
6	Grammatophora marina	4	80
8	Trachyneis aspera	3	60
9	Pleurosigma pelagicum	3,5	70
10	Nitzschia lorenziana	2	40
11	Nitzschia sigma	8	160
12	Pseudonitzschia sp.	5	100
13	Surirella ovalis	0,5	10
14	Prorocentrum gracile	1	20
15	Dinophysis audata	4	80
16	Dinophysis ripos	1	20
17	Dinophysis sp.	1	20
18	Ceratium breve	0,5	10
19	Ceratium deflexum	1	20
20	Ceratium curca	1	20
21	Gonyaulax sp.	1	20
22	Protoperidinium conicum	0,5	10
23	Protoperidinium depressum	1	20
24	Protoperidinium sp.2	8	160
25	Alexandrium spp.	12	240
26	Oscillatoria sp.1	1	20
		406	8120

STATION 21			
Sampling date: 04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Coscinodiscus cf. subtilis	1	10
2	Thalassiosira spp.	18	180
	Guinardia striata		20
4	Guinardia striata	2	20
5	Chaetoceros spp.	84	840
6	Ceratium bergonii	2	20
7	Thalassionema nitzschoides	54	540
8	Thalassiothrix rauenfeldi	16	160
9	Synedra sp.	116	1160

1	<i>Melosira</i> sp.	2	20
2	<i>Navicula</i> sp.	5	50
3	<i>Diploneis</i> sp.	1	10
4	<i>Nitzschia</i> sp.	5	50
5	<i>Nitzschia</i> sp.	1	10
6	<i>Nitzschia</i> sp.	1	10
7	<i>Nitzschia</i> sp.	9	90
8	<i>Nitzschia</i> sp.	5	50
9	<i>Bacillaria</i> sp.	3	30
10	<i>Surirella</i> sp.	1	10
11	<i>Campylodiscus</i> sp.	1	10
		34	340

STATION 7			
Sampling date: 03/04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassiosira</i> sp.	1	20
2	<i>Thalassiothrix</i> sp.	1	20
3	<i>Navicula</i> sp.	1	20
4	<i>Navicula</i> sp.	3	60
5	<i>Trachyneis</i> sp.	6	120
6	<i>Diploneis</i> sp.	8	160
7	<i>Nitzschia</i> sp.	13	260
8	<i>Nitzschia</i> sp.	2	40
9	<i>Nitzschia</i> sp.	36	720
10	<i>Nitzschia</i> sp.	86	1720
11	<i>Nitzschia</i> sp.	6	120
12	<i>Pseudonitzschia</i> sp.	8	160
13	<i>Surirella</i> sp.	1	20
		172	3440

STATION 8			
Sampling date: 03/04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Synedra</i> sp.	3	60
2	<i>Grammatophora</i> sp.	2	40
3	<i>Navicula</i> sp.	2	40
4	<i>Diploneis</i> sp.	3	60
5	<i>Pleurosigma</i> sp.	4	80
6	<i>Amphiprora</i> sp.	1	20
7	<i>Nitzschia</i> sp.	14	280
8	<i>Nitzschia</i> sp.	11	220
9	<i>Nitzschia</i> sp.	80	1600
10	<i>Nitzschia</i> sp.	2	40
11	<i>Nitzschia</i> sp.	19	380
12	<i>Bacillaria</i> sp.	8	160
13	<i>Surirella</i> sp.	3	60
14	<i>Surirella</i> sp.	1	20
15	<i>Campylodiscus</i> sp.	1	20
16	<i>Prorocentrum</i> sp.	1	20
		155	3100

STATION 9			
Sampling date: 03/04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Cyclotella</i> sp.	2	40
2	<i>Synedra</i> sp.	1	20
3	<i>Grammatophora</i> sp.	1	20
4	<i>Navicula</i> sp.	1	20
5	<i>Navicula</i> sp.	0,5	10
6	<i>Trachyneis</i> sp.	1	20
7	<i>Diploneis</i> sp.	2	40
8	<i>Pleurosigma</i> sp.	1	20
9	<i>Nitzschia</i> sp.	16	320
10	<i>Nitzschia</i> sp.	3	60
11	<i>Nitzschia</i> sp.	166	3320
12	<i>Nitzschia</i> sp.	2	40
13	<i>Nitzschia</i> sp.	53	1060
14	<i>Bacillaria</i> sp.	262	5240
15	<i>Pseudonitzschia</i> sp.	29	580
16	<i>Surirella</i> sp.	1	20
		541,5	10830

STATION 10			
Sampling date: 04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Coccolodiscus</i> sp.	1	10
	<i>Thalassiosira</i> sp.	2	20

10	<i>Cocconeis</i> sp.	1	10
11	<i>Navicula</i> sp.	1	10
12	<i>Nitzschia</i> sp.	2	20
13	<i>Nitzschia</i> sp.	2	20
14	<i>Bacillaria</i> sp.	5	50
15	<i>Pseudonitzschia</i> sp.	8	80
16	<i>Dinophysis</i> sp.	1	10
17	<i>Gonyaulax</i> sp.	8	80
18	<i>Gonyaulax</i> sp.	1	10
19	<i>Protoperidium</i> sp.	12	120
20	<i>Protoperidium</i> sp.	3	30
21	<i>Protoperidium</i> sp.	1	10
22	<i>Protoperidium</i> sp.	4	40
23	<i>Alexandrium</i> sp.	22	220
		365	3650

STATION 22			
Sampling date: 04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassiosira</i> sp.	10	200
2	<i>Guinardia</i> sp.	2	40
3	<i>Chaetoceros</i> sp.	26	520
4	<i>Thalassonema</i> sp.	8	160
5	<i>Thalassiothrix</i> sp.	12	240
6	<i>Nitzschia</i> sp.	24	480
7	<i>Grammatophora</i> sp.	1	20
8	<i>Cocconeis</i> sp.	9	180
9	<i>Navicula</i> sp.	1	20
10	<i>Navicula</i> sp.	1	20
11	<i>Pleurosigma</i> sp.	1	20
12	<i>Pleurosigma</i> sp.	1	20
13	<i>Amphiprora</i> sp.	1	20
14	<i>Nitzschia</i> sp.	2	40
15	<i>Nitzschia</i> sp.	1	20
16	<i>Nitzschia</i> sp.	1	20
17	<i>Prorocentrum</i> sp.	2	40
18	<i>Prorocentrum</i> sp.	0,5	10
19	<i>Dinophysis</i> sp.	6	60
20	<i>Polykrikos</i> sp.	0,5	10
21	<i>Ceratium</i> sp.	33	660
22	<i>Ceratium</i> sp.	0,5	10
23	<i>Ceratium</i> sp.	1	20
24	<i>Gonyaulax</i> sp.	3	60
25	<i>Protoperidium</i> sp.	8	160
26	<i>Protoperidium</i> sp.	3	60
27	<i>Protoperidium</i> sp.	2	40
28	<i>Protoperidium</i> sp.	3	60
29	<i>Protoperidium</i> sp.	1	20
30	<i>Protoperidium</i> sp.	5	100
31	<i>Alexandrium</i> sp.	2	40
		171,5	3430

STATION 23			
Sampling date: 05/04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Asteromphalus</i> sp.	1	10
2	<i>Thalassiosira</i> sp.	9	90
3	<i>Chaetoceros</i> sp.	Nhieu	Nhieu
4	<i>Thalassonema</i> sp.	3	30
5	<i>Thalassiothrix</i> sp.	13	130
6	<i>Synedra</i> sp.	12	120
7	<i>Grammatophora</i> sp.	7	70
8	<i>Cocconeis</i> sp.	20	200
9	<i>Navicula</i> sp.	2	20
10	<i>Navicula</i> sp.	4	40
11	<i>Trachyneis</i> sp.	2	20
12	<i>Diploneis</i> sp.	2	20
13	<i>Pleurosigma</i> sp.	1	10
14	<i>Nitzschia</i> sp.	4	40
15	<i>Pseudonitzschia</i> sp.	1	10
16	<i>Campylodiscus</i> sp.	2	20
17	<i>Prorocentrum</i> sp.	6	60
18	<i>Gymnodinium</i> sp.	2	20
19	<i>Polykrikos</i> sp.	4	40
20	<i>Ceratium</i> sp.	10	100
21	<i>Gonyaulax</i> sp.	1	10
22	<i>Gonyaulax</i> sp.	4	40
23	<i>Gonyaulax</i> sp.	2	20
24	<i>Protoperidium</i> sp.	9	90
25	<i>Protoperidium</i> sp.	1	10

3	<i>Skeletonema</i> <i>costatum</i>	18	180
4	<i>Leptocylindrus</i> <i>panicus</i>	1	30
5	<i>Biddulphia</i> <i>sp.</i>	1	10
6	<i>Thalassiothrix</i> <i>rauenfeldii</i>	2	20
7	<i>Synedra</i> <i>oillonii</i>	1	10
8	<i>Navicula</i> <i>membranacea</i>	7	70
9	<i>Pleurosigma</i> <i>angulatum</i>	1	10
10	<i>Nitzschia</i> <i>sigma</i>	24	240
11	<i>Nitzschia</i> <i>sp.</i>	2	20
12	<i>Surirella</i> <i>ovalis</i>	3	30
13	<i>Protoperidium</i> <i>sp.</i>	2	20
		92	920

STATION#11			
Sampling Date: 05/04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira</i> <i>granulata</i>	10	100
2	<i>Coscinodiscus</i> <i>subtilis</i>	1	10
3	<i>Thalassiosira</i> <i>sp.</i>	40	400
4	<i>Skeletonema</i> <i>costatum</i>	12	120
5	<i>Chaetoceros</i> <i>curvisetus</i>	16	160
6	<i>Chaetoceros</i> <i>sp.</i>	8	80
7	<i>Thalassionema</i> <i>nitzschoides</i>	1	10
8	<i>Nitzschia</i> <i>longissima</i>	4	40
9	<i>Nitzschia</i> <i>sigma</i>	25	250
11	<i>Pseudonitzschia</i> <i>sp.1</i>	1	10
12	<i>Surirella</i> <i>ovalis</i>	6	60
13	<i>Ceratium</i> <i>breve</i>	1	10
14	<i>Gonyaulax</i> <i>sp.</i>	17	170
15	<i>Protoperidium</i> <i>ellucidum</i>	1	10
16	<i>Protoperidium</i> <i>sp.2</i>	7	70
17	<i>Alexandrium</i> <i>sp.</i>	4	40
18	<i>Oscillatoria</i> <i>sp.2</i>	2	20
		159	1590

STATION#12			
Sampling Date: 05/04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Coscinodiscus</i> <i>onesianus</i>	1	10
2	<i>Thalassiosira</i> <i>sp.</i>	20	200
3	<i>Chaetoceros</i> <i>sp.</i>	2	20
4	<i>Thalassionema</i> <i>nitzschoides</i>	2	20
5	<i>Thalassiothrix</i> <i>rauenfeldii</i>	6	60
6	<i>Pleurosigma</i> <i>angulatum</i>	1	10
7	<i>Nitzschia</i> <i>longissima</i>	3	30
8	<i>Nitzschia</i> <i>sigma</i>	11	110
9	<i>Nitzschia</i> <i>sigma</i> <i>intercedens</i>	2	20
10	<i>Pseudonitzschia</i> <i>sp.1</i>	4	40
11	<i>Surirella</i> <i>ovalis</i>	1	10
12	<i>Gonyaulax</i> <i>sp.</i>	5	50
13	<i>Protoperidium</i> <i>sp.</i>	21	210
		101	1010

STATION#13			
Sampling Date: 04/04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira</i> <i>granulata</i>	3	60
2	<i>Cyclotella</i> <i>triata</i>	1	20
3	<i>Cyclotella</i> <i>compta</i>	1	20
4	<i>Cyclotella</i> <i>sp.</i>	1	20
5	<i>Coscinodiscus</i> <i>steromphalus</i>	1	20
6	<i>Coscinodiscus</i> <i>onesianus</i>	1	20
7	<i>Coscinodiscus</i> <i>subtilis</i>	1	20
8	<i>Coscinodiscus</i> <i>sp.</i>	1	20
9	<i>Thalassiosira</i> <i>sp.</i>	16	320
10	<i>Leptocylindrus</i> <i>panicus</i>	4	80
11	<i>Chaetoceros</i> <i>sp.</i>	27	540
12	<i>Biddulphia</i> <i>longicirris</i>	0.5	10
13	<i>Thalassionema</i> <i>nitzschoides</i>	2	40
14	<i>Thalassiothrix</i> <i>rauenfeldii</i>	5	100
15	<i>Navicula</i> <i>palpebralis</i>	3	60
16	<i>Navicula</i> <i>legans</i>	1	20
17	<i>Amphiprora</i> <i>lata</i>	1	20
18	<i>Nitzschia</i> <i>lorenziana</i>	1	20
19	<i>Nitzschia</i> <i>longissima</i>	5	100
20	<i>Nitzschia</i> <i>sigma</i>	4	80
21	<i>Nitzschia</i> <i>sigma</i> <i>intercedens</i>	0.5	10
22	<i>Nitzschia</i> <i>sp.</i>	1	20

26	<i>Protoperidium</i> <i>virgens</i>	9	90
27	<i>Protoperidium</i> <i>depressum</i>	1	10
28	<i>Protoperidium</i> <i>ellucidum</i>	1	10
29	<i>Protoperidium</i> <i>sp.</i>	4	40
30	<i>Alexandrium</i> <i>sp.</i>	1	10
		128	1280

STATION#24			
Sampling Date: 05/04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassiosira</i> <i>sp.</i>	5	50
2	<i>Thalassionema</i> <i>nitzschoides</i>	1	10
3	<i>Thalassiothrix</i> <i>rauenfeldii</i>	7	70
4	<i>Synedra</i> <i>sp.</i>	15	150
5	<i>Grammatophora</i> <i>marina</i>	3	30
6	<i>Cocconeis</i> <i>scutellum</i>	7	70
7	<i>Trachyneis</i> <i>aspera</i>	2	20
8	<i>Nitzschia</i> <i>lorenziana</i>	1	10
9	<i>Nitzschia</i> <i>sigma</i>	3	30
10	<i>Pseudonitzschia</i> <i>sp.</i>	2	20
11	<i>Campylodiscus</i> <i>cheneis</i>	1	10
12	<i>Campylodiscus</i> <i>angulatum</i>	1	10
13	<i>Proocentrum</i> <i>mexicanum</i>	9	90
14	<i>Dinophysis</i> <i>audata</i>	30	300
15	<i>Gymnodinium</i> <i>sp.</i>	2	20
16	<i>Ceratium</i> <i>surca</i>	3	30
17	<i>Gonyaulax</i> <i>spinifera</i>	6	60
18	<i>Gonyaulax</i> <i>verior</i>	1	10
19	<i>Protoperidium</i> <i>teini</i>	3	30
20	<i>Protoperidium</i> <i>tonicum</i>	1	10
21	<i>Protoperidium</i> <i>virgens</i>	4	40
22	<i>Protoperidium</i> <i>ellucidum</i>	1	10
23	<i>Protoperidium</i> <i>sp.</i>	18	180
		97	970

STATION#25			
Sampling Date: 07/04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira</i> <i>lumuloides</i>	2	20
2	<i>Chaetoceros</i> <i>sp.</i>	80	800
3	<i>Thalassiothrix</i> <i>rauenfeldii</i>	1	10
4	<i>Synedra</i> <i>sp.</i>	5	50
5	<i>Grammatophora</i> <i>marina</i>	5	50
6	<i>Navicula</i> <i>legans</i>	1	10
7	<i>Diploneis</i> <i>smithii</i>	1	10
8	<i>Pleurosigma</i> <i>angulatum</i>	1	10
9	<i>Nitzschia</i> <i>lorenziana</i>	2	20
10	<i>Nitzschia</i> <i>sigma</i>	6	60
11	<i>Pseudonitzschia</i> <i>sp.</i>	5	50
12	<i>Proocentrum</i> <i>sp.</i>	1	10
13	<i>Gymnodinium</i> <i>anguineum</i>	1	10
14	<i>Gyrodinium</i> <i>spirale</i>	2	20
15	<i>Ceratium</i> <i>surca</i>	10	100
16	<i>Gonyaulax</i> <i>sp.</i>	2	20
17	<i>Gonyaulax</i> <i>spinifera</i>	4	40
18	<i>Protoperidium</i> <i>tonicum</i>	3	30
19	<i>Protoperidium</i> <i>sp.</i>	16	160
20	<i>Alexandrium</i> <i>sp.</i>	1	10
		149	1490

STATION#26			
Sampling Date: 07/04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassiosira</i> <i>sp.</i>	2	20
2	<i>Thalassionema</i> <i>nitzschoides</i>	1	10
3	<i>Thalassiothrix</i> <i>rauenfeldii</i>	2	20
4	<i>Synedra</i> <i>sp.</i>	4	40
5	<i>Grammatophora</i> <i>marina</i>	7	70
6	<i>Navicula</i> <i>legans</i>	1	10
7	<i>Trachyneis</i> <i>aspera</i>	2	20
8	<i>Pleurosigma</i> <i>naviculaceum</i>	1	10
9	<i>Nitzschia</i> <i>lorenziana</i>	5	50
10	<i>Nitzschia</i> <i>longissima</i>	3	30
11	<i>Pseudonitzschia</i> <i>sp.</i>	1	10
12	<i>Campylodiscus</i> <i>cheneis</i>	1	10
13	<i>Proocentrum</i> <i>mexicanum</i>	4	40
14	<i>Dinophysis</i> <i>audata</i>	1	10
15	<i>Gymnodinium</i> <i>sp.</i>	1	10
16	<i>Gyrodinium</i> <i>spirale</i>	1	10

23	<i>Pseudonitzschia</i> sp. 1	3	60
24	<i>Dinophysis</i> cf. <i>ortii</i>	1	20
25	<i>Dinophysis</i> cf. <i>ortii</i>	1	20
26	<i>Closterium</i> sp.	3,5	70
		<b>93,5</b>	<b>1870</b>

**STATION 14**

Sampling date: 04/04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Leptocylindrus</i> <i>planicus</i>	3	60
2	<i>Chaetoceros</i> sp.	11	220
3	<i>Synedra</i> <i>pulchella</i>	8	160
4	<i>Navicula</i> <i>quadrata</i>	0,5	10
5	<i>Diploneis</i> <i>ornatus</i>	1	20
6	<i>Pleurosigma</i> <i>angulatum</i>	3	60
7	<i>Amphiprora</i> <i>palata</i>	5	100
8	<i>Nitzschia</i> <i>lorenziana</i>	7	140
9	<i>Nitzschia</i> <i>longissima</i>	16	320
10	<i>Nitzschia</i> <i>sigma</i>	4	80
11	<i>Nitzschia</i> sp.	20	400
12	<i>Bacillaria</i> <i>paxillifera</i>	20	400
13	<i>Prorocentrum</i> <i>gracile</i>	0,5	10
14	<i>Pediastrum</i> sp.	1	20
15	<i>Scenedesmus</i> sp.	30	600
16	<i>Closterium</i> sp.	4	80
17	<i>Gloetila</i> <i>pelagica</i>	23	460
		<b>157</b>	<b>3140</b>

**STATION 15**

Sampling date: 04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira</i> <i>granulata</i>	8	160
2	<i>Skeletonema</i> <i>costatum</i>	6	120
3	<i>Chaetoceros</i> <i>curvisetus</i>	40	800
4	<i>Chaetoceros</i> sp.	7	140
5	<i>Synedra</i> <i>pulchella</i>	9	180
6	<i>Navicula</i> <i>palpebralis</i>	1	20
7	<i>Pleurosigma</i> <i>angulatum</i>	3	60
8	<i>Nitzschia</i> <i>longissima</i>	9	180
9	<i>Nitzschia</i> <i>longissima</i> <i>reversa</i>	2	40
10	<i>Nitzschia</i> <i>sigma</i>	1	20
11	<i>Nitzschia</i> <i>sigma</i> <i>intercedens</i>	0,5	10
12	<i>Nitzschia</i> sp.	18	360
13	<i>Surirella</i> <i>tenerrima</i> <i>tenervosa</i>	0,5	10
14	<i>Protoperidium</i> sp.	1	20
15	<i>Peridinium</i> <i>linguacorne</i>	2	40
16	<i>Pediastrum</i> sp.	2	40
17	<i>Scenedesmus</i> sp.	26	520
18	<i>Staurastrum</i> sp.	3	60
19	<i>Closterium</i> sp.	4	80
		<b>325</b>	<b>6500</b>

**STATION 16**

Sampling date: 04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Cyclotella</i> sp.	1	20
2	<i>Coscinodiscus</i> <i>asteromphalus</i>	1	20
3	<i>Coscinodiscus</i> <i>onesianus</i>	3	60
4	<i>Coscinodiscus</i> <i>onesianus</i> <i>commutata</i>	1	20
5	<i>Coscinodiscus</i> <i>nodulifer</i>	1	20
6	<i>Thalassiosira</i> sp.	7	140
7	<i>Skeletonema</i> <i>costatum</i>	8	160
8	<i>Chaetoceros</i> sp.	78	1560
9	<i>Thalassonema</i> <i>nitzschoides</i>	8	160
10	<i>Thalassiothrix</i> <i>rauenfeldii</i>	19	380
11	<i>Pleurosigma</i> <i>affine</i>	1	20
12	<i>Nitzschia</i> <i>lorenziana</i>	1	20
13	<i>Nitzschia</i> <i>longissima</i>	2	40
14	<i>Nitzschia</i> <i>longissima</i> <i>reversa</i>	2	40
15	<i>Pseudonitzschia</i> sp.	1	20
16	<i>Dinophysis</i> <i>audata</i>	6	120
17	<i>Dinophysis</i> cf. <i>ortii</i>	1	20
18	<i>Dinophysis</i> sp.	1	20
19	<i>Gyrodinium</i> <i>spirale</i>	1	20
20	<i>Ceratium</i> <i>richoceros</i>	1	20
21	<i>Protoperidium</i> sp.	1	20
22	<i>Peridinium</i> <i>linguacorne</i>	4	80
		<b>156</b>	<b>3120</b>

17	<i>Ceratium</i> <i>surca</i>	12	120
18	<i>Chaetoceros</i> sp.	5	100
19	<i>Gonyaulax</i> <i>spinifera</i>	3	30
20	<i>Protoperidium</i> <i>depressum</i>	2	20
21	<i>Protoperidium</i> <i>ellucidum</i>	1	10
22	<i>Protoperidium</i> <i>leonis</i>	1	10
23	<i>Protoperidium</i> sp.	14	140
		<b>71</b>	<b>710</b>

**STATION 27**

Sampling date: 07/04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassiosira</i> sp.	2	20
3	<i>Thalassiothrix</i> <i>rauenfeldii</i>	2	20
4	<i>Synedra</i> sp.	5	50
5	<i>Cocconeis</i> <i>cutellum</i>	6	60
6	<i>Pleurosigma</i> <i>pelagicum</i>	1	10
7	<i>Nitzschia</i> <i>lorenziana</i>	3	30
8	<i>Nitzschia</i> <i>longissima</i>	1	10
9	<i>Nitzschia</i> <i>sigma</i>	10	100
10	<i>Prorocentrum</i> <i>mexicanum</i>	3	30
11	<i>Ceratium</i> <i>surca</i>	1	10
12	<i>Ceratium</i> <i>macroceros</i>	1	10
13	<i>Gonyaulax</i> <i>verior</i>	1	10
14	<i>Protoperidium</i> <i>tonicum</i>	1	10
15	<i>Protoperidium</i> <i>divergens</i>	1	10
16	<i>Protoperidium</i> sp.	15	150
		<b>121</b>	<b>1210</b>

**STATION 28**

Sampling date: 07/04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira</i> <i>granulata</i>	10	100
2	<i>Rhizosolenia</i> <i>valina</i>	19	180
3	<i>Chaetoceros</i> sp.	1	10
4	<i>Thalassiothrix</i> <i>rauenfeldii</i>	2	20
5	<i>Cocconeis</i> <i>cutellum</i>	10	100
6	<i>Pleurosigma</i> sp.	1	10
7	<i>Nitzschia</i> <i>longissima</i>	1	10
8	<i>Nitzschia</i> <i>sigma</i>	4	40
9	<i>Ceratium</i> <i>surca</i>	9	90
10	<i>Gonyaulax</i> <i>verior</i>	1	10
11	<i>Protoperidium</i> <i>teini</i>	6	60
12	<i>Alexandrium</i> sp.	1	10
		<b>47</b>	<b>470</b>

**STATION 29**

Sampling date: 07/04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Cyclotella</i> <i>comata</i>	1	10
2	<i>Thalassionema</i> <i>nitzschoides</i>	1	10
3	<i>Thalassiothrix</i> <i>rauenfeldii</i>	1	10
4	<i>Synedra</i> sp.	7	70
5	<i>Nitzschia</i> <i>lorenziana</i>	2	20
6	<i>Nitzschia</i> <i>longissima</i>	3	30
7	<i>Bacillaria</i> <i>paxillifera</i>	1	10
8	<i>Pseudonitzschia</i> sp. 1	4	40
9	<i>Gloetila</i> <i>pelagica</i>	4	40
		<b>24</b>	<b>240</b>

**STATION 30**

Sampling date: 07/04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Coscinodiscus</i> <i>onesianus</i>	1	20
2	<i>Lauderia</i> <i>boealis</i>	1	20
3	<i>Leptocylindrus</i> <i>planicus</i>	2	40
4	<i>Guinardia</i> <i>laccida</i>	7	140
5	<i>Guinardia</i> <i>striata</i>	10	200
6	<i>Rhizosolenia</i> <i>valina</i>	0,5	10
7	<i>Chaetoceros</i> sp.	11	220
8	<i>Eucampia</i> <i>ornata</i>	1,5	30
9	<i>Thalassionema</i> <i>nitzschoides</i>	8	160
10	<i>Thalassiothrix</i> <i>rauenfeldii</i>	121	2420
11	<i>Synedra</i> <i>gailonii</i>	2	40
12	<i>Navicula</i> <i>palpebralis</i>	1	20
13	<i>Navicula</i> <i>membranacea</i>	14	280
14	<i>Navicula</i> <i>legans</i>	1	20

STATION#17			
No	Sampling date: 04/2006		
	SPECIES	QUANTITY	CELLS/L
1	<i>Coscinodiscus asteromphalus</i>	2	40
2	<i>Coscinodiscus subtilis</i>	62	1240
3	<i>Thalassiosira</i> sp.	14	280
4	<i>Skeletonema costatum</i>	1	20
5	<i>Chaetoceros</i> sp.	62	1240
6	<i>Thalassionema nitzschoides</i>	9	180
7	<i>Thalassiothrix frauenfeldii</i>	9	180
8	<i>Synedra caillonii</i>	0.5	10
9	<i>Pleurosigma affine</i>	0.5	10
10	<i>Pleurosigma angulatum</i>	0.5	10
11	<i>Pleurosigma pelagicum</i>	1	20
12	<i>Nitzschia borenziana</i>	3	60
13	<i>Nitzschia longissima</i>	4	80
14	<i>Nitzschia longissima</i> , <i>reversa</i>	4	80
15	<i>Nitzschia sigma</i> , <i>intercedens</i>	1	20
16	<i>Bacillaria paxilliferis</i>	3	60
17	<i>Pseudonitzschia</i> sp.	6	120
18	<i>Dinophysis caudata</i>	5	100
19	<i>Dinophysis</i> cf. <i>ortii</i>	2	40
20	<i>Dinophysis</i> <i>nitra</i>	1	20
21	<i>Protoperdinium divergens</i>	2	40
22	<i>Protoperdinium oceanicum</i>	1	20
23	<i>Protoperdinium</i> sp.	5	100
24	<i>Gloetilia pelagica</i>	4	80
		<b>138.5</b>	<b>2770</b>

STATION#18			
No	Sampling date: 04/2006		
	SPECIES	QUANTITY	CELLS/L
1	<i>Coscinodiscus subtilis</i>	1	10
2	<i>Thalassiosira</i> sp.	9	90
3	<i>Skeletonema costatum</i>	8	80
4	<i>Chaetoceros</i> sp.	36	360
5	<i>Thalassiothrix frauenfeldii</i>	2	20
6	<i>Synedra</i> sp.	2	20
7	<i>Navicula legans</i>	1	10
8	<i>Gyrosigma</i> sp.	1	10
9	<i>Pleurosigma fasciola</i>	1	10
10	<i>Nitzschia borenziana</i>	6	60
11	<i>Nitzschia longissima</i>	4	40
12	<i>Nitzschia longissima</i> , <i>reversa</i>	3	30
13	<i>Nitzschia sigma</i>	1	10
14	<i>Pseudonitzschia</i> sp.	4	40
15	<i>Dinophysis</i> cf. <i>ortii</i>	1	10
16	<i>Pediastrum</i> sp.	1	10
		<b>81</b>	<b>810</b>

15	<i>Trachyneis</i> <i>spersa</i>	1	20
16	<i>Pleurosigma affine</i>	1	20
17	<i>Pleurosigma angulatum</i>	3	60
18	<i>Pleurosigma naviculaceum</i>	10	200
19	<i>Nitzschia borenziana</i>	1	20
20	<i>Nitzschia longissima</i>	2	40
22	<i>Pseudonitzschia</i> sp. 1	1	20
23	<i>Campylodiscus cheneis</i>	0.5	10
24	<i>Proocentrum</i> <i>americanus</i>	1	20
25	<i>Dinophysis caudata</i>	8	160
26	<i>Dinophysis</i> sp.	0.5	10
27	<i>Ceratium breve</i>	0.5	10
28	<i>Ceratium flexum</i>	0.5	10
29	<i>Ceratium furca</i>	3	60
30	<i>Ceratium minus</i>	3	60
31	<i>Ceratium ofoidii</i>	1	20
32	<i>Ceratium richoceros</i>	1	20
33	<i>Protoperdinium teinii</i>	1	20
34	<i>Protoperdinium oceanicum</i>	0.5	10
35	<i>Protoperdinium divergens</i>	3	60
36	<i>Goniadoma polyedra</i>	1	20
37	<i>Dictyocha speculum</i>	0.5	10
38	<i>Oscillatoria</i> sp2	6	120
		<b>240</b>	<b>4800</b>

STATION#32			
No	Sampling date: 06/04/2006		
	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassiothrix frauenfeldii</i>	4	40
2	<i>Synedra</i> sp.	2	20
3	<i>Navicula</i> <i>lyra</i>	1	10
5	<i>Gyrosigma strigile</i>	1	10
5	<i>Nitzschia longissima</i>	5	50
6	<i>Nitzschia longissima</i> , <i>reversa</i>	2	20
	<i>Nitzschia sigma</i> , <i>intercedens</i>	3	30
8	<i>Nitzschia</i> sp.	8	80
9	<i>Pseudonitzschia</i> sp. 1	2	20
10	<i>Ceratium furca</i>	1	10
11	<i>Gonyaulax</i> <i>superior</i>	1	10
12	<i>Gonyaulax</i> <i>rotundata</i>	2	20
13	<i>Protoperdinium teinii</i>	4	40
14	<i>Protoperdinium</i> sp.	1	10
15	<i>Alexandrium</i> sp	2	20
		<b>39</b>	<b>390</b>

STATION#33			
No	Sampling date: 06/04/2006		
	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassiothrix frauenfeldii</i>	3	30
2	<i>Nitzschia borenziana</i>	1	10
3	<i>Nitzschia longissima</i> , <i>reversa</i>	1	10
4	<i>Pseudonitzschia</i> sp. 1	2	20
5	<i>Campylodiscus angulatum</i>	1	10
6	<i>Ceratium furca</i>	1	10
7	<i>Protoperdinium teinii</i>	1	10
8	<i>Alexandrium</i> sp	20	200
9	<i>Oscillatoria</i> sp2	1	10
		<b>31</b>	<b>310</b>

STATION#37			
No	Sampling date: 06/04/2006		
	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassiothrix frauenfeldii</i>	2	20
2	<i>Synedra</i> sp.	1	10
3	<i>Diploneis smithii</i>	1	10
4	<i>Nitzschia longissima</i>	2	20
5	<i>Nitzschia sigma</i>	1	10
6	<i>Pseudonitzschia</i> sp.	1	10
7	<i>Alexandrium</i> sp.	16	160
		<b>24</b>	<b>240</b>

STATION#34			
No	Sampling date: 06/04/2006		
	SPECIES	QUANTITY	CELLS/L
1	<i>Chaetoceros</i> sp.	64	640
2	<i>Thalassiothrix frauenfeldii</i>	1	10
3	<i>Synedra</i> sp.	1	10
4	<i>Grammatophora marino</i>	4	40

5	<i>Nitzschia longissima</i>	1	10
6	<i>Nitzschia sigma</i>	5	50
7	<i>Nitzschia</i> sp.	2	20
8	<i>Pseudonitzschia</i> spp.	3	30
9	<i>Alexandrium</i> spp.	268	2680
		349	3490

STATION 35			
Sampling Date: 06/04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Chaetoceros</i> sp.	4	40
2	<i>Thalassiothrix frauenfeldii</i>	1	10
3	<i>Nitzschia orenziana</i>	4	40
4	<i>Nitzschia longissima</i>	2	20
5	<i>Nitzschia longissima</i> , <i>reversa</i>	1	10
6	<i>Nitzschia sigma</i>	3	30
7	<i>Gonyaulax</i> sp.	1	10
8	<i>Alexandrium pseudogonyaulax</i>	586	5860
		602	6020

STATION 36			
Sampling Date: 06/04/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassionema nitzschioides</i>	2	20
2	<i>Cocconeis cutellum</i>	2	20
3	<i>Gyrosigma trigile</i>	1	10
4	<i>Nitzschia longissima</i>	2	20
5	<i>Nitzschia sigma</i>	7	70
6	<i>Pseudonitzschia</i> sp. 1	2	20
7	<i>Alexandrium</i> spp.	1234	12340
		1250	12500

**Survey Institute of Marine Resources and Environment, April 2006 Zooplankton Species**

STATION A1			
Sampling Date: 03/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Penilia schmackeri</i>	1	50
2	<i>Paracalanus parvus</i>	14	700
3	<i>Schmackeria tubia</i>	1	50
4	<i>Allodiaptomus</i> sp.	1	50
5	<i>Oithona similis</i>	2	100
6	<i>Thermocyclops hyalinus</i>	34	1700
7	<i>Corycaeus ahli</i>	1	50
8	<i>Oikopleura dioica</i>	1	50
9	ATAM	1	50
10	<i>Acartiella sinensis</i>	1	50
<b>Total</b>		<b>57</b>	<b>2850</b>

STATION A2			
Sampling Date: 03/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	2	20
2	<i>Acartia lausi</i>	8	80
3	<i>Oithona similis</i>	2	20
4	<i>Thermocyclops hyalinus</i>	1	10
5	<i>Cypridina noctiluca</i>	2	20
<b>Total</b>		<b>15</b>	<b>150</b>

STATION A3			
Sampling Date: 04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	11	550
2	<i>Pseudodiaptomus ansisus</i>	1	50
3	<i>Acartia pacifica</i>	3	150
4	<i>Oithona similis</i>	6	300
5	<i>Oithona brevicornis</i>	12	600
6	<i>Thermocyclops hyalinus</i>	6	300
7	<i>Euterpina cutifront</i>	1	50
8	<i>Harpacticoda</i>	4	200
9	<i>Acarbella sinensis</i>	7	350
<b>Total</b>		<b>51</b>	<b>2550</b>

STATION A4			
Sampling Date: 05/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Acartiella sinensis</i>	66	3300
<b>Total</b>		<b>66</b>	<b>3300</b>

STATION A5			
Sampling Date: 04/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Acartiella sinensis</i>	202	40400
<b>Total</b>		<b>202</b>	<b>40400</b>

STATION A6			
Sampling Date: 06/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Piaphanosoma sarsi</i>	1	50
2	<i>Acartia lausi</i>	50	2500
3	<i>Harpacticoda</i>	1	50
4	<i>Acarbella sinensis</i>	234	11700
<b>Total</b>		<b>286</b>	<b>14300</b>

STATION A7			
Sampling Date: 08/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Schmackeria tubulosa</i>	2	100
2	<i>Acartia pacifica</i>	7	350
3	<i>Thermocyclops hyalinus</i>	18	900

STATION G20			
Sampling Date: 05/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Sagitta serassa</i>	1	50
2	<i>Pseudodiaptomus ansisus</i>	2	100
3	<i>Temora turbinata Dana</i>	1	50
4	<i>Labidocera minuta</i>	2	100
5	<i>Acartia lausi</i>	4	200
6	<i>Euterpina cutifront</i>	1	50
7	<i>Oikopleura dioica</i>	2	100
8	C. on	2	100
<b>Total</b>		<b>15</b>	<b>750</b>

STATION G21			
Sampling Date: 04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Pseudodiaptomus ansisus</i>	2	100
2	<i>Acartia lausi</i>	1	50
3	<i>Oithona dana</i>	4	200
4	<i>Oithona similis</i>	1	50
5	<i>Oncaea venusta</i>	6	300
6	<i>Microcyclops varican</i>	4	200
7	<i>Clytemnestra scutellata</i>	1	50
8	<i>Amphipoda</i>	1	50
9	<i>Oikopleura dioica</i>	3	150
10	<i>Copepoda</i>	10	500
<b>Total</b>		<b>33</b>	<b>1650</b>

STATION H22			
Sampling Date: 05/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	15	1500
2	<i>Centropages persini</i>	1	100
3	<i>Oithona similis</i>	10	1000
4	<i>Oncaea venusta</i>	1	100
5	<i>Oikopleura dioica</i>	4	300
6	AT Balanus	11	1100
7	<i>Cypridina noctiluca</i>	8	800
<b>Total</b>		<b>49</b>	<b>4900</b>

STATION H23			
Sampling Date: 05/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	3	150
2	<i>Centropages persini</i>	5	250
3	<i>Temora turbinata Dana</i>	1	50
4	<i>Labidocera minuta</i>	1	50
5	<i>Acartia lausi</i>	1	50
6	<i>Oithona similis</i>	1	50
7	<i>Thermocyclops hyalinus</i>	3	150
8	<i>Oikopleura dioica</i>	7	350
9	AT Gastropoda	12	600
10	AT Balanus	2	100
11	<i>Cypridina noctiluca</i>	2	100
<b>Total</b>		<b>38</b>	<b>1900</b>

STATION H24			
Sampling Date: 05/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	1	50
2	<i>Centropages persini</i>	1	50
3	<i>Labidocera minuta</i>	3	150
4	<i>Acartia lausi</i>	1	50
5	<i>Oithona similis</i>	1	50
6	<i>Euterpina cutifront</i>	1	50
7	<i>Oikopleura dioica</i>	8	400



4	ATBrachyura	2	100
5	Citon	1	50
6	ATAm	40	2000
7	Cypridinaoctiluca	1	50
8	Acartiella sinensis	3	150
<b>Total</b>		<b>74</b>	<b>3700</b>

STATION 8			
Sampling Date: 04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	Paracalanusparvus	1	50
2	Oncaeaconifera	1	50
3	ATBivalvia	1	50
4	Acartiella sinensis	8	400
<b>Total</b>		<b>11</b>	<b>550</b>

STATION 9			
Sampling Date: 06/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	Paracalanusparvus	6	60
2	Oithonaana	1	10
3	Acarbella sinensis	10	100
<b>Total</b>		<b>17</b>	<b>170</b>

STATION 10			
Sampling Date: 05/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	Acartia lausi	1	10
2	Amphipoda	1	10
<b>Total</b>		<b>2</b>	<b>20</b>

STATION 11			
Sampling Date: 05/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	Paracalanusparvus	19	190
2	Centropagesborsini	2	20
3	Acartia lausi	51	510
4	Oithona similis	1	10
5	Euterpinae cutifront	2	20
6	Hyalocylis triata	1	10
7	Oikopleura dioica	19	190
8	ATBivalvia	5	50
9	ATActinotrocha	2	20
<b>Total</b>		<b>102</b>	<b>1020</b>

STATION 12			
Sampling Date: 05/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	Oikopleura dioica	1	50
2	Cypridinaoctiluca	1	50
3	Acartiella sinensis	31	1550
<b>Total</b>		<b>33</b>	<b>1650</b>

STATION 13			
Sampling Date: 03/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	Podon schmackeri	1	10
2	Paracalanusparvus	28	280
3	Oithona similis	2	20
4	Oikopleura dioica	2	20
<b>Total</b>		<b>33</b>	<b>330</b>

STATION 15			
Sampling Date: 04/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	Paracalanusparvus	2	20
2	Acartia lausi	1	10
3	Euterpinae cutifront	1	10

8	ATBivalvia	1	50
9	ATGastropoda	1	50
10	Citon	1	50
11	Cypridinaoctiluca	1	50
<b>Total</b>		<b>20</b>	<b>1000</b>

STATION 25			
Sampling Date: 07/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	Paracalanusparvus	6	300
2	Labidocera minuta	3	150
3	Acartia lausi	1	50
4	Corycaeus andrewsi	1	50
5	Corycaeus alhi	1	50
6	Oikopleura dioica	1	50
7	ATGastropoda	3	150
8	Cypridinaoctiluca	1	50
<b>Total</b>		<b>17</b>	<b>850</b>

STATION 26			
Sampling Date: 07/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	Paracalanusparvus	6	60
2	Acartia lausi	6	60
3	Oithona similis	9	90
4	Oikopleura dioica	3	30
5	Copepoda	12	120
6	Cypridinaoctiluca	24	240
<b>Total</b>		<b>60</b>	<b>600</b>

STATION 27			
Sampling Date: 07/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	Paracalanusparvus	8	400
2	Pseudodiaptomus sinensis	2	100
3	Acartia lausi	1	50
4	Oithona similis	23	1150
5	Corycaeus andrewsi	1	50
6	Corycaeus alhi	1	50
7	Euterpinae cutifront	2	100
8	Amphipoda	1	50
9	Citon	1	50
10	Cypridinaoctiluca	1	50
11	Acarbella sinensis	1	50
<b>Total</b>		<b>42</b>	<b>2100</b>

STATION 28			
Sampling Date: 07/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	Paracalanusparvus	8	400
2	Acartia lausi	2	100
3	Oithona brevicornis	27	1350
4	Corycaeus alhi	3	150
5	ATActinotrocha	1	50
6	Citon	2	100
7	Cypridinaoctiluca	1	50
<b>Total</b>		<b>44</b>	<b>2200</b>

STATION 29			
Sampling Date: 07/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	Paracalanusparvus	1	50
2	Acartia lausi	1	50
3	ATBrachyura	1	50
<b>Total</b>		<b>3</b>	<b>150</b>

STATION 30			
Sampling Date: 07/04/2006			
No	SPECIES	QUANTITY	INDS/M3

4	<i>Cypridina</i> <i>noctiluca</i>	1	10
<b>Total</b>		<b>5</b>	<b>50</b>

STATION#16			
Sampling Date: 04/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> <i>parvus</i>	32	320
2	<i>Centropages</i> <i>persini</i>	1	10
3	<i>Acartia</i> <i>lausi</i>	3	30
4	<i>Oithona</i> <i>similis</i>	2	20
5	<i>Oikopleura</i> <i>dioica</i>	5	50
<b>Total</b>		<b>43</b>	<b>430</b>

STATION#17			
Sampling Date: 04/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Thermocyclops</i> <i>thalinus</i>	1	50
2	<i>Corycaeus</i> <i>thalhi</i>	6	300
3	<i>Oikopleura</i> <i>dioica</i>	12	600
<b>Total</b>		<b>19</b>	<b>950</b>

STATION#18			
Sampling Date: 04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> <i>parvus</i>	3	30
2	<i>Acartia</i> <i>lausi</i>	1	10
3	<i>Oithona</i> <i>similis</i>	4	40
4	<i>Euterpina</i> <i>cutifront</i>	3	30
5	<i>Acartia</i> <i>sinensis</i>	3	30
<b>Total</b>		<b>14</b>	<b>140</b>

No	SPECIES	QUANTITY	INDS/M3
1	<i>Sagitta</i> <i>enflata</i>	6	60
2	<i>Sagitta</i> <i>sp.</i>	1	10
3	<i>Penilia</i> <i>schmackeri</i>	1	10
4	<i>Canthocalanus</i> <i>pauper</i>	1	10
5	<i>Acrocalanus</i> <i>fibber</i>	1	10
6	<i>Paracalanus</i> <i>parvus</i>	22	220
7	<i>Centropages</i> <i>persini</i>	1	10
8	<i>Temora</i> <i>turbinata</i> <i>Dana</i>	5	50
9	<i>Acartia</i> <i>lausi</i>	3	30
10	<i>Pontellina</i> <i>plumata</i>	3	30
11	<i>Oithona</i> <i>mana</i>	2	20
12	<i>Oithona</i> <i>similis</i>	6	60
13	<i>Corycaeus</i> <i>andrewsi</i>	1	10
14	<i>Corycaeus</i> <i>thalhi</i>	1	10
15	<i>Euterpina</i> <i>cutifront</i>	10	100
16	<i>Oikopleura</i> <i>dioica</i>	6	60
17	<i>Acartia</i> <i>sinensis</i>	4	40
<b>Total</b>		<b>74</b>	<b>740</b>

STATION#32			
Sampling Date: 06/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> <i>parvus</i>	1	10
2	<i>Temora</i> <i>turbinata</i> <i>Dana</i>	1	10
3	<i>Labidocera</i> <i>minuta</i>	2	20
4	<i>Acartia</i> <i>lausi</i>	8	80
5	<i>Oithona</i> <i>similis</i>	1	10
6	<i>Cypridina</i> <i>noctiluca</i>	1	10
<b>Total</b>		<b>14</b>	<b>140</b>

STATION#33			
Sampling Date: 06/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> <i>parvus</i>	1	10
2	<i>Acartia</i> <i>lausi</i>	12	120
3	<i>Oikopleura</i> <i>dioica</i>	1	10
<b>Total</b>		<b>14</b>	<b>140</b>

STATION#37			
Sampling Date: 06/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> <i>parvus</i>	3	30
2	<i>Acartia</i> <i>lausi</i>	1	10
3	<i>Oithona</i> <i>similis</i>	1	10
4	<i>AT</i> <i>gastropoda</i>	1	10
5	<i>Cypridina</i> <i>noctiluca</i>	4	40
6	<i>Acartia</i> <i>sinensis</i>	9	90
<b>Total</b>		<b>19</b>	<b>190</b>

STATION#34			
Sampling Date: 06/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> <i>parvus</i>	9	450
2	<i>Temora</i> <i>turbinata</i> <i>Dana</i>	2	100
3	<i>Oithona</i> <i>similis</i>	3	150
4	<i>Cypridina</i> <i>noctiluca</i>	3	150
<b>Total</b>		<b>17</b>	<b>850</b>

STATION#35			
Sampling Date: 06/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Labidocera</i> <i>minuta</i>	2	20
2	<i>Acartia</i> <i>lausi</i>	4	40
3	<i>Oithona</i> <i>similis</i>	5	50
4	<i>Cypridina</i> <i>noctiluca</i>	6	60
<b>Total</b>		<b>17</b>	<b>170</b>

STATION#36			
Sampling Date: 06/04/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> <i>parvus</i>	4	40
<b>Total</b>		<b>4</b>	<b>40</b>

Survey Institute of Marine Resources and Environment, May 2006

Site	Station	T/B	Depth(m)	Turbidity	Temp(°C)	Flow velocity(m/s)		BottomSedimentType	TotalSolid(SS)	Suspende dSolid(SS)	pH	Salinity (ppt)	DOE(mg/l)	NO <sub>2</sub> (mg/l)	NO <sub>3</sub> (mg/l)	NH <sub>4</sub> (mg/l)	Alkalinity(pH)	TotalP	TotalN	ChlorophyllaC	ZooBenthos(Biomass)	Phytoplankton(Ind/cell/l)	Microalgae	ToxicAlgae(cell/l)	TotalColiforms	E.coli	Vibrio
						Tide(Flood)	Tide(Ebb)																				
A	1	T		0.6	28.5					6.45	1.0	5.20	0.003	<0.05	0.06			0.02	<0.05	1.9				2800			
		M																									
		R	0.5				Mud																				
	2	T		0.5	28.4						6.23	1.0	5.20	0.005	<0.05	<0.02			0.04	<0.05	1.4				46000		
		M																									
		R	1.0				Mud&Clay																				
3	T		1.0	31.3						6.45	1.0	7.30	<0.002	<0.05	0.04			0.03	0.06	0.5				2700			
	M																										
	R	0.7				Mud&Clay&Sand																					
B	4	T		1.0	31.5					7.87	10.0	7.70	<0.002	0.07	0.03			0.03	1.51	2.6				46000			
		M																									
		R	0.9				Mud&Clay																				
	5	T		1.2	29.9						8.15	14.5	7.50	0.004	0.05	0.04			0.07	0.79	8.2				>240000		
		M																									
		R	1.5				Mud&Clay																				
6	T		1.0	31.6						8.96	10.0	8.30	<0.002	0.05	0.02			0.02	0.75	0.6				1500			
	M																										
	R	0.4				Mud&Clay&Sand																					
C	7	T		1.8	31.7					8.21	14.5	7.50	<0.002	0.06	0.02			0.04	0.99	3.5				21000			
		M																									
		R	2.7				Mud&Clay																				
	8	T		1.0	31.5						8.65	13.0	7.40	0.002	0.06	0.02			0.04	0.83	3.3				15000		
		M																									
		R	1.3				Mud&Clay																				
9	T		0.7	32.4						8.26	14.5	7.60	0.003	0.06	0.05			0.04	1.07	2.6				1100			
	M																										
	R	0.7				Mud&Sand																					
D	10	T		1.0	30.5					7.99	18.0	6.10	<0.002	0.07	<0.02			0.06	1.31	4.2				2000			
		M																									
		R	1.0				Mud&Sand																				
	11	T		1.2	30.5						8.10	18.0	6.10	<0.002	0.06	0.06			0.05	1.88	4.3				2000		
		M																									
		R	1.8				Mud&Sand																				
12	T		1.0	31.2						8.10	17.0	6.20	<0.002	0.06	0.04			0.04	1.74	5.9				2800			
	M																										
	R	1.4				Mud&Clay																					
E	13	T		2.0	29.0					8.21	30.0	7.10	<0.002	0.07	0.02			0.04	1.08	1.1				1100			
		M																									
		R	11.0				Mud&Sand																				
	14	T		1.5	28.1						8.23	31.5	7.50	<0.002	0.07	0.02			0.03	1.64	1.0				>240000		
		M																									
		R	1.0				Sand																				
15	T		1.1	31.0						8.14	13.5	7.50	0.007	0.21	0.02			0.04	0.92	0.2				2800			
	M																										
	R	0.8				Mud																					
F	16	T		0.9	30.0					8.03	23.0	6.15	0.004	0.08	0.05			0.03	0.54	3.2				2100			
		M																									
		R	1.0				Mud&Sand																				
	17	T		1.0	30.1						8.07	24.0	6.80	0.003	0.06	0.05			0.02	0.90	3.6				110000		
		M																									
		R	1.8				Mud																				
18	T		1.0	30.5						7.96	21.0	6.00	0.002	0.07	0.04			0.03	1.72	1.5				2300			
	M																										
	R	0.9				Mud&Clay&Sand																					
G	19	T		1.4	31.8					8.34	25.0	7.90	0.002	0.08	0.05			0.04	1.47	3.2				4300			
		M																									
		R	1.3				Mud&Sand																				
	20	T		1.1	31.6						8.09	24.0	6.90	0.004	0.07	0.04			0.04	1.32	4.3				24000		
		M																									
		R	2.7				Mud&Sand																				
21	T		0.6	32.3						8.21	25.0	7.80	0.002	0.05	0.05			0.03	1.63	2.8				4300			
	M																										
	R	0.3				Mud&Sand																					
22	T		0.9	32.0						8.35	23.0	7.80	0.004	0.10	0.08			0.04	0.95	6.1				15000			
	M																										
	R	1.5				Mud&Sand																					
H	T		1.0	31.1						8.15	22.5	7.40	0.004	0.08	0.04			0.04	0.94	1.7				>240000			
	M																										
	R	3.7				Mud&Sand																					
I	T		1.2	31.0						8.22	21.5	7.60	0.003	0.06	0.04			0.06	1.81	4.7				4300			
	M																										
	R	2.0				Mud&Sand																					
J	T																										

**Survey of Marine Resources and Environment, May 2006 Phytoplankton Species**

STATION 1			
Sampling Date: 26/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira granulata</i>	6	120
2	<i>Melosira granulata</i>	1090	2180
3	<i>Asterionella</i> sp.	1145	22900
4	<i>Nitzschia lorenziana</i>	2	40
5	<i>Dinophysis</i> sp.	1	20
6	<i>Alexandrium</i> sp.	3	60
7	<i>Pediastrum</i> sp.	10	200
8	<i>Scenedesmus quadricauda</i>	15	300
9	<i>Spirogyra</i> sp.	1	20
10	<i>Staurastrum</i> sp.	90	1800
11	<i>Phacus</i> cf. <i>longicauda</i>	1	20
12	<i>Dinoflagellate</i> group	1	20
		<b>2365</b>	<b>47300</b>

STATION 2			
Sampling Date: 26/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira granulata</i>	610	6100
2	<i>Thalassiothrix frauenfeldii</i>	5	50
3	<i>Asterionella</i> sp.	300	3000
4	<i>Gyrosigma</i> sp.	1	10
5	<i>Surirella tenera</i> var. <i>hervosa</i>	1	10
6	<i>Protoperidinium</i> sp.	1	10
7	<i>Peridinium</i> sp.	50	500
8	<i>Alexandrium</i> sp.	6	60
9	<i>Dictyochaetula</i>	1	10
10	<i>Pediastrum</i> sp.	5	50
11	<i>Scenedesmus quadricauda</i>	4	40
12	<i>Staurastrum</i> sp.	161	1610
13	<i>Euglena</i> sp.2	1	10
		<b>1146</b>	<b>11460</b>

STATION 3			
Sampling Date: 26/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassiosira</i> sp.	1	20
2	<i>Chaetoceros</i> sp.	8	160
3	<i>Synedra gailionii</i>	1	20
4	<i>Amphiprorata</i>	1	20
5	<i>Nitzschia longissima</i> var. <i>reversa</i>	1	20
6	<i>Nitzschia sigma</i>	2	40
7	<i>Surirella valis</i>	2	40
8	<i>Surirella tenera</i> var. <i>hervosa</i>	2	40
9	<i>Gymnodinium sanguineum</i>	1	20
10	<i>Oscillatoria</i> sp.2	4	80
11	<i>Pediastrum</i> sp.	2	40
12	<i>Scenedesmus</i> sp.	19	380
13	<i>Gloeotila pelagicum</i>	6	120
14	<i>Dinoflagellates</i> group	489	9780
		<b>539</b>	<b>10780</b>

STATION 4			
Sampling Date: 26/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira granulata</i>	6	60
2	<i>Asterionella</i> sp.	195	1950
3	<i>Diploneis smithii</i>	3	30
4	<i>Pleurosigma</i> sp.	1	10
5	<i>Nitzschia lorenziana</i>	1	10
6	<i>Nitzschia longissima</i> var. <i>reversa</i>	7	70
7	<i>Nitzschia palpebralis</i>	1	10
9	<i>Peridinium</i> sp.	173	1730
8	<i>Ceratium</i> sp.	1	10
10	<i>Alexandrium</i> sp.	1	10
11	<i>Anabaena</i> sp.	8	80
12	<i>Pediastrum simplex</i>	1	10
13	<i>Staurastrum</i> sp.	4	40
14	<i>Phacus</i> sp.1	2	20
15	<i>Dinoflagellate</i> group	4	40
		<b>408</b>	<b>4080</b>

STATION 5			
Sampling Date: 26/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Cyclotella</i> sp.	1	10
2	<i>Thalassiosira</i> sp.	7	70
3	<i>Navicula elegans</i>	2	20

STATION 20			
Sampling Date: 28/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Coscinodiscus</i> cf. <i>subtilis</i>	1	10
3	<i>Trachyneis aspera</i>	1	10
4	<i>Pleurosigma angulatum</i>	1	10
5	<i>Pleurosigma angulatum</i>	2	20
6	<i>Nitzschia lorenziana</i>	1	10
7	<i>Nitzschia longissima</i> var. <i>reversa</i>	12	20
8	<i>Nitzschia sigma</i>	14	140
9	<i>Prorocentrum</i> <i>americanus</i>	5	50
10	<i>Metadinophysis sinensis</i>	1	10
11	<i>Gymnodinium sanguineum</i>	1	10
12	<i>Ceratium furca</i>	2	20
13	<i>Gonyaulax polygramma</i>	3	30
14	<i>Gonyaulax spinifera</i>	3	30
15	<i>Protoperidinium steinii</i>	5	50
16	<i>Protoperidinium pellucidum</i>	18	180
17	<i>Protoperidinium</i> sp.	44	440
18	<i>Peridinium quinquecarne</i>	6	60
19	<i>Alexandrium</i> sp.	2	20
20	<i>Dinoflagellates</i> group	5	50
		<b>128</b>	<b>1280</b>

STATION 21			
Sampling Date: 28/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Coscinodiscus bipartitus</i>	1	10
2	<i>Actinocyclus splendens</i>	1	10
3	<i>Thalassiosira</i> sp.	208	2080
4	<i>Pleurosigma affine</i>	2	20
5	<i>Pleurosigma angulatum</i>	1	10
6	<i>Pleurosigma</i> sp.	1	10
7	<i>Nitzschia sigma</i>	7	70
9	<i>Metadinophysis sinensis</i>	1	10
10	<i>Ceratium furca</i>	2	20
11	<i>Ceratium minus</i>	1	10
12	<i>Gonyaulax</i> sp.	2	20
13	<i>Protoperidinium pellucidum</i>	13	130
14	<i>Protoperidinium</i> sp.	1	10
15	<i>Peridinium quinquecarne</i>	6	60
16	<i>Scrippsiella</i> sp.	2	20
17	<i>Dictyochaetula</i>	2	20
18	<i>Dictyochaetula</i>	1	10
19	<i>Trichodesmium thirocun</i>	1	10
21	<i>Diatom</i>	2	20
		<b>271</b>	<b>2710</b>

STATION 22			
Sampling Date: 28/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassiosira</i> sp.	7	70
2	<i>Grammatophora marina</i>	3	30
3	<i>Cocconeis cutellum</i>	131	1310
4	<i>Gyrosigma pencei</i>	1	10
5	<i>Pleurosigma affine</i>	7	70
6	<i>Pleurosigma angulatum</i>	1	10
7	<i>Pleurosigma</i> sp.	4	40
8	<i>Amphiprorata</i>	1	10
9	<i>Nitzschia longissima</i> var. <i>reversa</i>	1	10
10	<i>Nitzschia sigma</i>	10	100
11	<i>Nitzschia sigma</i> var. <i>intercedens</i>	1	10
12	<i>Gymnodinium sanguineum</i>	2	20
13	<i>Ceratium furca</i>	3	30
14	<i>Gonyaulax</i> sp.	15	150
15	<i>Protoperidinium pellucidum</i>	4	40
16	<i>Protoperidinium pentagonum</i>	1	10
17	<i>Protoperidinium</i> sp.2	2	20
18	<i>Protoperidinium</i> sp.	13	130
19	<i>Peridinium</i> sp.	11	110
20	<i>Alexandrium</i> sp.	1	10
21	<i>Alexandrium pseudogonyaulax</i>	8	80
22	<i>Anabaena</i> sp.	34	340
23	<i>Dinoflagellate</i> group	21	210
24	<i>Diatom</i>	2	20
		<b>284</b>	<b>2840</b>

No	SPECIES	QUANTITY	CELLS/L
4	<i>Trachyneis</i> <i>aspera</i>	3	30
5	<i>Diploneis</i> <i>mithii</i>	4	40
6	<i>Gyrosigma</i> <i>sp.</i>	1	10
7	<i>Pleurosigma</i> <i>pelagicum</i>	1	10
8	<i>Nitzschia</i> <i>longissima</i>	2	20
9	<i>Nitzschia</i> <i>sigma</i>	18	180
10	<i>Suirella</i> <i>ovalis</i>	7	70
11	<i>Prorocentrum</i> <i>mexicanum</i>	1	10
12	<i>Gonyaulax</i> <i>polygramma</i>	2	20
13	<i>Gonyaulax</i> <i>verior</i>	1	10
14	<i>Protoperidinium</i> <i>steinii</i>	1	10
15	<i>Protoperidinium</i> <i>sp.2</i>	15	150
16	<i>Peridinium</i> <i>sp.</i>	1	10
17	<i>Alexandrium</i> <i>sp.</i>	1	10
18	<i>Oscillatoria</i> <i>sp.2</i>	5	50
19	<i>Dinoflagellate</i> <i>group</i>	54	540
		127	1270

STATION#B6			
Sampling Date: 26/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Chaetoceros</i> <i>subtilis</i>	4	40
2	<i>Diploneis</i> <i>mithii</i>	2	20
3	<i>Pleurosigma</i> <i>sp.</i>	1	10
4	<i>Nitzschia</i> <i>sigma</i>	1	10
5	<i>Protoperidinium</i> <i>sp.2</i>	3	30
6	<i>Peridinium</i> <i>quinquecornu</i>	1	10
7	<i>Peridinium</i> <i>sp.</i>	104	1040
8	<i>Alexandrium</i> <i>sp.</i>	54	540
9	<i>Oscillatoria</i> <i>sp.1</i>	13	130
10	<i>Anabaena</i> <i>sp.</i>	1	10
11	<i>Spirogyra</i> <i>tonia</i>	5	50
12	<i>Dinoflagellate</i> <i>group</i>	3	30
13	<i>Diatom</i>	1	10
		194	1940

STATION#C7			
Sampling Date: 26/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Coscinodiscus</i> <i>steromphalus</i>	1	10
2	<i>Thalassiosira</i> <i>sp.</i>	3	30
3	<i>Navicula</i> <i>palpebralis</i>	2	20
4	<i>Navicula</i> <i>cancelata</i>	2	20
5	<i>Diploneis</i> <i>mithii</i>	2	20
6	<i>Gyrosigma</i> <i>sp.</i>	1	10
7	<i>Pleurosigma</i> <i>sp.</i>	4	40
8	<i>Nitzschia</i> <i>lorenziana</i>	1	10
9	<i>Nitzschia</i> <i>longissima</i>	1	10
10	<i>Nitzschia</i> <i>sigma</i>	11	110
11	<i>Nitzschia</i> <i>sp.</i>	3	30
12	<i>Suirella</i> <i>genera</i>	3	30
13	<i>Suirella</i> <i>sp.</i>	2	20
14	<i>Campylodiscus</i> <i>sp.</i>	4	40
15	<i>Prorocentrum</i> <i>marginalatum</i>	2	20
16	<i>Gyrodinium</i> <i>spirale</i>	1	10
17	<i>Gonyaulax</i> <i>sp.</i>	2	20
18	<i>Protoperidinium</i> <i>sp.</i>	2	20
19	<i>Peridinium</i> <i>sp.</i>	36	360
20	<i>Scenedesmus</i> <i>sp.</i>	8	80
21	<i>Dinoflagellate</i> <i>group</i>	4	40
		97	970

STATION#C8			
Sampling Date: 26/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Cyclotella</i> <i>sp.</i>	1	10
2	<i>Thalassiosira</i> <i>sp.</i>	2	20
3	<i>Odontella</i> <i>sinensis</i>	1	10
4	<i>Thalassiothrix</i> <i>trauenfeldii</i>	1	10
5	<i>Grammatophora</i> <i>marina</i>	1	10
6	<i>Navicula</i> <i>cancelata</i>	3	30
7	<i>Trachyneis</i> <i>aspera</i>	10	100
8	<i>Diploneis</i> <i>mithii</i>	1	10
9	<i>Diploneis</i> <i>trabro</i>	1	10
10	<i>Gyrosigma</i> <i>sp.</i>	1	10
11	<i>Pleurosigma</i> <i>pelagicum</i>	1	10
12	<i>Amphora</i> <i>lineata</i>	2	20
13	<i>Nitzschia</i> <i>lorenziana</i>	4	40
14	<i>Nitzschia</i> <i>longissima</i>	3	30
15	<i>Nitzschia</i> <i>sigma</i>	7	70
16	<i>Nitzschia</i> <i>sp.</i>	4	40
17	<i>Suirella</i> <i>ovalis</i>	1	10
18	<i>Campylodiscus</i> <i>undulatus</i>	1	10

STATION#H23			
Sampling Date: 28/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Cocconeis</i> <i>cutellum</i>	10	100
2	<i>Pleurosigma</i> <i>affine</i>	4	40
3	<i>Pleurosigma</i> <i>sp.</i>	3	30
4	<i>Nitzschia</i> <i>sigma</i>	6	60
5	<i>Suirella</i> <i>gemma</i>	1	10
6	<i>Prorocentrum</i> <i>mexicans</i>	4	40
7	<i>Metadinophysis</i> <i>sinensis</i>	2	20
8	<i>Gymnodinium</i> <i>anguineum</i>	16	160
9	<i>Gonyaulax</i> <i>sp.</i>	3	30
10	<i>Gonyaulax</i> <i>verior</i>	1	10
11	<i>Gonyaulax</i> <i>rotundata</i>	1	10
12	<i>Protoperidinium</i> <i>pellucidum</i>	5	50
13	<i>Protoperidinium</i> <i>pentagonum</i>	1	10
14	<i>Protoperidinium</i> <i>sp.2</i>	7	70
15	<i>Protoperidinium</i> <i>sp.</i>	7	70
16	<i>Scrippsiella</i> <i>sp.</i>	2	20
17	<i>Alexandrium</i> <i>sp.</i>	5	50
18	<i>Alexandrium</i> <i>pseudogonyaulax</i>	5	50
19	<i>Dinoflagellate</i> <i>group</i>	51	510
		139	1390

STATION#H24			
Sampling Date: 28/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassiosira</i> <i>sp.</i>	4	40
2	<i>Cocconeis</i> <i>cutellum</i>	16	160
3	<i>Trachyneis</i> <i>aspera</i>	5	50
4	<i>Gyrosigma</i> <i>balticum</i>	1	10
5	<i>Pleurosigma</i> <i>angulatum</i>	8	80
6	<i>Pleurosigma</i> <i>oviculaceum</i>	3	30
7	<i>Pleurosigma</i> <i>pelagicum</i>	4	40
8	<i>Amphora</i> <i>lineata</i>	1	10
9	<i>Amphora</i> <i>hyalina</i>	1	10
10	<i>Amphora</i> <i>quadrata</i>	1	10
11	<i>Nitzschia</i> <i>longissima</i>	2	20
12	<i>Nitzschia</i> <i>sigma</i>	2	20
13	<i>Suirella</i> <i>ovalis</i>	1	10
14	<i>Campylodiscus</i> <i>undulatus</i>	3	30
15	<i>Prorocentrum</i> <i>mexicans</i>	3	30
16	<i>Prorocentrum</i> <i>mexicanum</i>	1	10
17	<i>Gymnodinium</i> <i>anguineum</i>	2	20
18	<i>Gonyaulax</i> <i>sp.</i>	9	90
19	<i>Gonyaulax</i> <i>polygramma</i>	2	20
20	<i>Protoperidinium</i> <i>conicum</i>	3	30
21	<i>Protoperidinium</i> <i>pellucidum</i>	3	30
22	<i>Protoperidinium</i> <i>depressum</i>	1	10
23	<i>Protoperidinium</i> <i>sp.</i>	18	180
24	<i>Protoperidinium</i> <i>sp.</i>	18	180
25	<i>Alexandrium</i> <i>sp.</i>	5	50
26	<i>Oscillatoria</i> <i>sp.2</i>	3	30
27	<i>Dinoflagellate</i> <i>group</i>	18	180
		128	1280

STATION#H25			
Sampling Date: 30/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Synedra</i> <i>pulchella</i>	1	10
2	<i>Cocconeis</i> <i>cutellum</i>	2	20
3	<i>Trachyneis</i> <i>aspera</i>	1	10
4	<i>Pleurosigma</i> <i>angulatum</i>	1	10
5	<i>Prorocentrum</i> <i>mexicans</i>	1	10
6	<i>Gonyaulax</i> <i>polygramma</i>	6	60
7	<i>Protoperidinium</i> <i>steinii</i>	1	10
8	<i>Protoperidinium</i> <i>pellucidum</i>	1	10
9	<i>Protoperidinium</i> <i>sp.</i>	12	120
10	<i>Protoperidinium</i> <i>sp.</i>	18	180
11	<i>Scrippsiella</i> <i>spinifera</i>	1	10
12	<i>Oscillatoria</i> <i>sp.2</i>	1	10
		47	470

STATION#H26			
Sampling Date: 28/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassiosira</i> <i>sp.</i>	1	10
2	<i>Cocconeis</i> <i>cutellum</i>	4	40
3	<i>Pleurosigma</i> <i>affine</i>	2	20
4	<i>Pleurosigma</i> <i>sp.</i>	2	20
5	<i>Nitzschia</i> <i>sigma</i>	4	40
6	<i>Nitzschia</i> <i>sp.</i>	1	10
7	<i>Gymnodinium</i> <i>anguineum</i>	2	20

19	<i>Campylodiscus angulatum</i>	5	50
20	<i>Gyrodinium spirale</i>	1	10
21	<i>Gonyaulax verior</i>	1	10
22	<i>Pediastrum</i> sp.	1	10
23	<i>Dinoflagellates</i> group	70	700
			<b>1230</b>

STATION#9			
Sampling Date: 26/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira granulata</i> var. <i>angustissima</i>	12	120
2	<i>Cyclotella</i> sp.	1	10
3	<i>Coscinodiscus</i> sp.	1	10
4	<i>Actinocyclus splendens</i>	1	10
5	<i>Thalassiosira</i> sp.	1	10
6	<i>Navicula</i> sp.	1	10
7	<i>Navicula palpebralis</i>	1	10
8	<i>Navicula cancellata</i>	50	500
9	<i>Diploneis smithii</i>	2	20
10	<i>Gyrosigma balticum</i>	15	150
11	<i>Gyrosigma penneri</i>	9	90
12	<i>Gyrosigma</i> sp.	1	10
13	<i>Pleurosigma</i> sp.	10	100
14	<i>Nitzschia longissima</i> var. <i>reversa</i>	1	10
15	<i>Nitzschia</i> sp. 1	1	10
16	<i>Nitzschia</i> sp.	7	70
17	<i>Campylodiscus richensis</i>	5	50
18	<i>Prorocentrum</i> sp.	1	10
19	<i>Ceratium furca</i>	1	10
20	<i>Peridinium</i> sp.	36	360
21	<i>Dinoflagellates</i> group	1	10
22	<i>Diatom</i>	4	40
		<b>174</b>	<b>1740</b>

STATION#10			
Sampling Date: 27/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Coscinodiscus hanesianus</i> var. <i>commutata</i>	1	10
2	<i>Thalassiosira</i> sp.	5	50
3	<i>Nitzschia arenziana</i>	1	10
4	<i>Nitzschia longissima</i>	1	10
5	<i>Nitzschia sigma</i>	9	90
6	<i>Dinophysis quadrata</i>	1	10
7	<i>Gyrodinium spirale</i>	1	10
8	<i>Gonyaulax</i> sp.	7	70
9	<i>Protoperdinium pellucidum</i>	4	40
10	<i>Protoperdinium punctulatum</i>	2	20
11	<i>Peridinium quinquecorne</i>	1	10
12	<i>Peridinium</i> sp.	12	120
13	<i>Scripsiella</i> sp.	1	10
14	<i>Dinoflagellates</i> group	22	220
		<b>70</b>	<b>700</b>

STATION#11			
Sampling Date: 27/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira</i> sp.	8	80
2	<i>Cyclotella</i> sp.	2	20
3	<i>Coscinodiscus</i> sp.	2	20
4	<i>Thalassiosira</i> sp.	8	80
5	<i>Biddulphia longicirrus</i>	1	10
6	<i>Odontella mobiliensis</i>	1	10
7	<i>Odontella sinensis</i>	0	0
8	<i>Thalassiothrix frauenfeldii</i>	3	30
9	<i>Grammatophora marina</i>	3	30
10	<i>Navicula cancellata</i>	3	30
11	<i>Mastogloia</i> sp.	1	10
12	<i>Pleurosigma</i> sp.	4	40
13	<i>Amphora quadrata</i>	1	10
14	<i>Nitzschia arenziana</i>	10	100
15	<i>Nitzschia longissima</i>	2	20
16	<i>Nitzschia sigma</i>	12	120
17	<i>Surrella gemma</i>	2	20
18	<i>Surrella</i> sp.	6	60
19	<i>Prorocentrum minimum</i>	3	30
20	<i>Metadinophysis sinensis</i>	1	10
21	<i>Gonyaulax verior</i>	1	10
22	<i>Peridinium quinquecorne</i>	2	20
23	<i>Peridinium</i> sp.	31	310
24	<i>Alexandrium</i> sp.	9	90
25	<i>Alexandrium pseudogonyaulax</i>	1	10
26	<i>Dinoflagellates</i> group	16	160

8	<i>Gonyaulax</i> sp.	7	70
9	<i>Protoperdinium</i> sp.	5	50
10	<i>Peridinium quinquecorne</i>	1	10
11	<i>Peridinium</i> sp.	9	90
12	<i>Alexandrium</i> sp.	1	10
13	<i>Alexandrium pseudogonyaulax</i>	1	10
14	<i>Oscillatoria</i> sp. 2	1	10
15	<i>Dinoflagellates</i> group	10	100
		<b>54</b>	<b>540</b>

STATION#27			
Sampling Date: 30/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Cyclotella</i> sp.	1	10
2	<i>Thalassiosira</i> sp.	1	10
3	<i>Climacophenia noniligera</i>	1	10
4	<i>Cocconeis scutellum</i>	9	90
5	<i>Pleurosigma affine</i>	1	10
6	<i>Nitzschia longissima</i>	1	10
7	<i>Nitzschia sigma</i>	4	40
8	<i>Dinophysis quadrata</i>	2	20
9	<i>Gymnodinium sanguineum</i>	4	40
10	<i>Gonyaulax</i> sp.	2	20
11	<i>Gonyaulax verior</i>	1	10
12	<i>Protoperdinium pellucidum</i>	4	40
13	<i>Protoperdinium</i> sp.	2	20
14	<i>Peridinium quinquecorne</i>	3	30
15	<i>Alexandrium pseudogonyaulax</i>	1	10
16	<i>Alexandrium pseudogonyaulax</i>	2	20
17	<i>Pyrophacus</i> sp.	2	20
18	<i>Trichodesmium thirocun</i>	1	10
19	<i>Dinoflagellates</i> group	37	370
20	<i>Diatom</i>	4	40
21		<b>88</b>	<b>880</b>

STATION#28			
Sampling Date: 30/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassiosira</i> sp.	5	50
2	<i>Thalassionema nitzschoides</i>	2	20
3	<i>Thalassiothrix frauenfeldii</i>	1	10
4	<i>Synedra gailonii</i>	2	20
5	<i>Cocconeis scutellum</i>	7	70
6	<i>Navicula legatis</i>	1	10
7	<i>Trachyneis spera</i>	1	10
8	<i>Pleurosigma affine</i>	2	20
9	<i>Amphora lineata</i>	1	10
10	<i>Nitzschia longissima</i>	1	10
11	<i>Nitzschia sigma</i>	4	40
12	<i>Gonyaulax</i> sp.	2	20
13	<i>Gonyaulax verior</i>	3	30
14	<i>Protoperdinium pellucidum</i>	2	20
15	<i>Protoperdinium</i> sp.	16	160
		<b>50</b>	<b>500</b>

STATION#29			
Sampling Date: 30/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Biddulphia reticulum</i>	1	10
2	<i>Synedra pulchella</i>	1	10
3	<i>Navicula</i> sp.	3	30
4	<i>Trachyneis spera</i>	2	20
5	<i>Gyrosigma</i> sp.	1	10
6	<i>Nitzschia</i> sp.	1	10
7	<i>Gymnodinium</i> sp.	1	10
8	<i>Gonyaulax</i> sp.	2	20
9	<i>Gonyaulax spinifera</i>	1	10
10	<i>Protoperdinium</i> sp.	22	220
11	<i>Protoperdinium</i> sp.	1	10
12	<i>Alexandrium</i> sp.	1	10
13	<i>Dinoflagellates</i> group	40	400
		<b>77</b>	<b>770</b>

STATION#30			
Sampling Date: 30/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Coscinodiscus asteromphalus</i>	0,5	10
2	<i>Asteromphalus leveanus</i>	1	20
3	<i>Thalassiosira</i> sp.	5	120
4	<i>Guinardia laccida</i>	1	20
5	<i>Guinardia striata</i>	11	220
6	<i>Pseudosolenia calcar-avis</i>	1	10
7	<i>Thalassiothrix frauenfeldii</i>	6	120

		133	1330
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STATION D12			
Sampling Date: 27/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Coscinodiscus onesianus	1	10
2	Coscinodiscus sp.	1	10
3	Thalassiosira spp.	6	60
4	Pleurosigma pelagicum	1	10
5	Amphiproralata	1	10
6	Nitzschia lorenziana	1	10
7	Nitzschia longissima	1	10
8	Nitzschia sigma	1	10
9	Surirella ovalis	1	10
10	Surirella tenera	2	20
11	Prorocentrum mexicanum	1	10
12	Prorocentrum mexicanum	1	10
13	Dinophysis cf. fortii	1	10
14	Gymnodinium sanguineum	1	10
15	Ceratium furca	1	10
16	Gonyaulax polygramma	3	30
17	Protoperidinium steinii	3	90
18	Protoperidinium sp.	13	130
19	Peridinium quinquecorne	9	90
20	Alexandrium spp.	2	20
		61	610

STATION E13			
Sampling Date: 27/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Paralia sulcata	2	20
2	Melosira cummuloides	3	30
3	Coscinodiscus steromphalus	1	10
4	Coscinodiscus culus-iridis	1	10
5	Coscinodiscus onesianus	4	40
6	Coscinodiscus sp.	2	20
7	Thalassiosira sp.	17	170
8	Lauderia borealis	2	20
9	Guinardia laccida	7	70
10	Guinardia striata	1	10
11	Pseudosolenia caravis	1	10
12	Cerataulina bergonii	6	60
13	Ditylum brightwellii	2	20
14	Thalassiosira nitzschoides	2	20
15	Thalassiothrix frauenfeldii	95	950
16	Cocconeis scutellum	4	40
17	Navicula membranacea	4	40
18	Pleurosigma sp.	4	40
19	Campylodiscus cheeneis	1	10
20	Prorocentrum mexicanum	2	20
21	Dinophysis laudata	7	70
22	Dinophysis cf. rotundata	1	10
23	Gymnodinium sanguineum	1	10
24	Gyrodinium spirale	2	20
25	Polykrikos schwartzii	1	10
26	Ceratium furca	7	70
27	Ceratium cf. ofoidii	1	10
28	Ceratium furca	1	10
29	Protoperidinium divergens	2	20
30	Protoperidinium pellucidum	6	60
31	Protoperidinium sp. 2	5	50
32	Scrippsiella sp.	1	10
33	Alexandrium sp.	2	20
34	Goniodoma polyedra	1	10
35	Zygabikodinium sp.	1	10
36	Dictyocha fibula	1	10
37	Anabaena sp.	1	10
38	Dinoflagellate group	6	60
39	Diatom	2	20
		215	2150

STATION E14			
Sampling Date: 27/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Coscinodiscus onesianus	1	20
2	Coscinodiscus sp.	2	40
3	Asteromphalus leveanus	0.5	10
4	Thalassiosira sp.	21	420
5	Lauderia borealis	6	120
6	Guinardia laccida	0.5	10
7	Guinardia striata	1	10
8	Rhizosolenia crassispina	1	20
9	Biddulphia regia	1	20

8	Navicula membranacea	7	140
9	Trachyneis aspera	2	40
10	Pleurosigma affine	1	20
11	Pleurosigma pelagicum	2	40
12	Nitzschia longissima	3	60
13	Nitzschia sigma	2	40
14	Metadinophysis sinensis	1	20
15	Dinophysis laudata	2	40
16	Dinophysis cf. fortii	0.5	10
17	Noctiluca scintillans	0.5	10
18	Ceratium furca	6	120
19	Ceratium furca	1	20
21	Gonyaulax sp.	1	20
22	Protoperidinium pelagicum	1	20
23	Protoperidinium pelagicum	1	20
24	Protoperidinium ruber	1	20
25	Protoperidinium sp.	1	20
26	Protoperidinium sp.	1	10
27	Alexandrium sp.	1	20
28	Lingulodinium sp.	1	20
29	Podolampas palmipes	1	20
		75.5	1510

STATION K32			
Sampling Date: 29/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Thalassiosira spp.	2	20
2	Nitzschia sigma	1	10
3	Gonyaulax sp.	1	10
4	Protoperidinium sp.	2	20
5	Protoperidinium sp.	11	110
6	Alexandrium sp.	1	10
7	Alexandrium pseudogonyaulax	5	50
8	Pyrophacus sp.	1	10
9	Closterium setaceum	2	20
10	Dinoflagellate group	11	110
		49	490

STATION K33			
Sampling Date: 05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Cyclotella sp.	1	10
2	Thalassiosira sp.	3	30
3	Thalassiothrix frauenfeldii	2	20
4	Pleurosigma pelagicum	6	60
5	Gonyaulax spinifera	3	30
6	Protoperidinium sp.	82	820
7	Protoperidinium sp.	3	30
8	Alexandrium sp.	1	10
		103	1030

STATION K37			
Sampling Date: 29/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Thalassiosira spp.	3	30
2	Thalassiothrix frauenfeldii	3	30
3	Cyclotella sp.	1	10
4	Nitzschia longissima	1	10
5	Gonyaulax sp.	1	10
6	Protoperidinium pelagicum	1	10
7	Protoperidinium pellucidum	2	20
8	Protoperidinium sp.	7	70
9	Alexandrium sp.	6	60
10	Dinoflagellates group	26	260
		52	520

STATION E14			
Sampling Date: 29/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Cyclotella omta	1	20
2	Cyclotella sp.	1	20
3	Thalassiosira sp.	3	60
4	Grammatophora narina	1	20
5	Trachyneis aspera	1	20
6	Pleurosigma pelagicum	1	20
7	Amphiproralata	1	20
8	Nitzschia lorenziana	2	40
9	Nitzschia longissima	1	20
10	Nitzschia longissima	1	20
11	Nitzschia sigma	6	120
12	Gonyaulax sp.	3	60
13	Protoperidinium steinii	1	20

10	Ditylum	1	20
11	Thalassionema	4	80
12	Thalassiothrix	34	680
13	Thalassiothrix	1	20
14	Synedra	1	20
15	Trachyneis	1	20
16	Diploleis	1	20
17	Pleurosigma	2	40
18	Pleurosigma	1	20
19	Amphora	0,5	10
20	Nitzschia	1	20
21	Nitzschia	1	20
22	Nitzschia	2	40
23	Bacillaria	2	40
24	Surirella	1	20
25	Prorocentrum	6	120
26	Prorocentrum	2	40
27	Dinophysis	2	40
28	Gymnodinium	1	20
29	Gymnodinium	0,5	10
30	Ceratium	1	20
31	Ceratium	1	20
32	Gonyaulax	35	700
33	Protoperidinium	1	20
34	Lingulodinium	1	20
35	Dictyocha	1	20
36	Dictyocha	1	20
37	Dictyocha	1	20
		143	2860

STATION 15			
Sampling date: 27/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Melosira	2	20
2	Melosira	4	40
3	Thalassiosira	28	280
4	Thalassiothrix	2	20
5	Synedra	1	10
6	Cocconeis	4	40
7	Navicula	1	10
8	Diploleis	1	10
9	Gyrosigma	1	10
10	Pleurosigma	5	50
11	Nitzschia	6	60
12	Nitzschia	4	40
13	Peridinium	25	250
14	Oscillatoria	3	30
15	Spirulina	2	20
16	Dinoflagellate	2	20
17	Diatom	4	40
		96	960

STATION 16			
Sampling date: 28/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Coscinodiscus	1	10
2	Coscinodiscus	4	40
3	Thalassiosira	125	1250
4	Thalassiothrix	2	20
5	Cocconeis	3	30
6	Pleurosigma	2	20
7	Pleurosigma	7	70
8	Amphora	1	10
9	Nitzschia	16	160
10	Nitzschia	13	130
11	Nitzschia	9	90
12	Prorocentrum	2	20
13	Dinophysis	1	10
14	Gymnodinium	1	10
15	Ceratium	3	30
16	Gonyaulax	1	10
17	Protoperidinium	1	10
18	Protoperidinium	1	10
19	Peridinium	1	10
20	Dinoflagellate	1	10
		195	1950

STATION 17			
Sampling date: 28/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Coscinodiscus	1	10
2	Coscinodiscus	3	30
3	Thalassiosira	39	390

14	Protoperidinium	2	40
15	Alexandrium	4	80
		29	580

STATION 35			
Sampling date: 29/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Diatom	1	10
2	Thalassionema	1	10
3	Cocconeis	2	20
4	Pleurosigma	1	10
5	Nitzschia	1	10
6	Nitzschia	2	40
7	Nitzschia	3	30
8	Nitzschia	1	10
9	Nitzschia	1	10
10	Prorocentrum	1	10
11	Gymnodinium	5	50
12	Gonyaulax	1	10
13	Protoperidinium	1	10
14	Protoperidinium	1	10
15	Protoperidinium	1	10
16	Protoperidinium	1	10
17	Protoperidinium	22	220
		50	500

STATION 36			
Sampling date: 29/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Biddulphia	1	10
2	Cocconeis	9	90
3	Gyrosigma	3	30
4	Pleurosigma	1	10
5	Amphiprora	1	10
6	Nitzschia	1	10
7	Nitzschia	3	30
8	Nitzschia	9	90
9	Surirella	1	10
10	Ceratium	1	10
11	Gonyaulax	1	10
12	Gonyaulax	1	10
13	Protoperidinium	2	20
14	Protoperidinium	9	90
15	Alexandrium	2	20
		45	450



4	<i>Guinardia</i> <i>flaccida</i>	1	10
5	<i>Thalassiothrix</i> <i>trauenfeldii</i>	2	20
6	<i>Pleurosigma</i> <i>angulatum</i>	1	10
7	<i>Amphiprora</i> <i>platata</i>	1	10
8	<i>Nitzschia</i> <i>longissima</i>	2	20
9	<i>Nitzschia</i> <i>longissima</i> <i>reversa</i>	1	10
10	<i>Nitzschia</i> <i>sigma</i>	6	60
11	<i>Bacillaria</i> <i>paxillifera</i>	4	40
12	<i>Surirella</i> <i>emma</i>	1	10
13	<i>Prorocentrum</i> <i>minicans</i>	10	100
14	<i>Gymnodinium</i> <i>sanguineum</i>	10	100
15	<i>Ceratium</i> <i>chofoidii</i>	1	10
16	<i>Gonyaulax</i> <i>sp</i>	10	100
17	<i>Gonyaulax</i> <i>spinifera</i>	1	10
18	<i>Protoperidinium</i> <i>steinii</i>	1	10
19	<i>Protoperidinium</i> <i>pellucidum</i>	18	180
20	<i>Peridinium</i> <i>quinqueorne</i>	35	350
21	<i>Alexandrium</i> <i>sp</i>	1	10
22	<i>Dinoflagellates</i> <i>group</i>	25	250
		174	1740

STATION 18			
Sampling Date: 28/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassiosira</i> <i>sp</i>	8	80
2	<i>Cocconeis</i> <i>scutellum</i>	19	190
3	<i>Pleurosigma</i> <i>affine</i>	9	90
4	<i>Nitzschia</i> <i>sigma</i>	4	40
5	<i>Surirella</i> <i>emma</i>	2	20
6	<i>Prorocentrum</i> <i>minicans</i>	1	10
7	<i>Metadinophysis</i> <i>sinensis</i>	2	20
8	<i>Dinophysis</i> <i>audata</i>	1	10
9	<i>Gymnodinium</i> <i>sanguineum</i>	163	1630
10	<i>Gonyaulax</i> <i>sp</i>	18	180
11	<i>Gonyaulax</i> <i>superior</i>	2	20
12	<i>Gonyaulax</i> <i>rotundata</i>	2	20
13	<i>Protoperidinium</i> <i>pellucidum</i>	11	110
14	<i>Protoperidinium</i> <i>sp. 1 (to, inhagan)</i>	29	290
15	<i>Protoperidinium</i> <i>sp</i>	1	10
16	<i>Peridinium</i> <i>quinqueorne</i>	3	30
17	<i>Scrippsiella</i> <i>sp</i>	15	150
18	<i>Alexandrium</i> <i>sp</i>	14	140
19	<i>Pyrophacus</i> <i>sp</i>	2	20
20	<i>Dinoflagellate</i> <i>group</i>	6	60
		312	3120

STATION 19			
Sampling Date: 28/05/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassiosira</i> <i>sp</i>	6	60
2	<i>Grammatophora</i> <i>marina</i>	2	20
3	<i>Cocconeis</i> <i>scutellum</i>	3	30
4	<i>Trachyneis</i> <i>aspera</i>	2	20
5	<i>Gyrosigma</i> <i>strigile</i>	1	10
6	<i>Pleurosigma</i> <i>naviculaceum</i>	1	10
7	<i>Pleurosigma</i> <i>pelagicum</i>	4	40
8	<i>Amphora</i> <i>yalina</i>	1	10
9	<i>Nitzschia</i> <i>longissima</i> <i>reversa</i>	5	50
10	<i>Nitzschia</i> <i>sigma</i>	5	50
11	<i>Prorocentrum</i> <i>minicans</i>	3	30
12	<i>Dinophysis</i> <i>audata</i>	2	20
13	<i>Gymnodinium</i> <i>sanguineum</i>	2	20
14	<i>Ceratium</i> <i>purca</i>	6	60
15	<i>Gonyaulax</i> <i>polygramma</i>	5	50
16	<i>Gonyaulax</i> <i>spinifera</i>	2	20
17	<i>Protoperidinium</i> <i>steinii</i>	11	110
18	<i>Protoperidinium</i> <i>pellucidum</i>	21	210
19	<i>Protoperidinium</i> <i>sp</i>	11	110
20	<i>Peridinium</i> <i>quinqueorne</i>	10	100
21	<i>Alexandrium</i> <i>minutum</i>	2	20
22	<i>Dinoflagellates</i> <i>group</i>	35	350
		140	1400

**Survey Institute of Marine Resources and Environment, May 2006 Zooplankton Species**

STATION 191			
Sampling Date: 26/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Penilia schmackeri</i>	2	200
2	<i>Paracalanus parvus</i>	3	300
3	<i>Labidocera roery</i>	1	100
4	<i>Oithona similis</i>	9	900
5	<i>Thermocyclops yalinus</i>	38	3800
6	<i>Diaphanosoma sarsi</i>	1	100
7	AT Bivalvia	1	100
8	AT T m	1	100
<b>Total</b>		<b>56</b>	<b>5600</b>

STATION 192			
Sampling Date: 26/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus crassirostris</i>	15	750
2	<i>Acartia rythraea</i>	19	950
3	<i>Oithona similis</i>	6	300
4	<i>Thermocyclops yalinus</i>	24	1200
5	<i>Moinodaphnia oleyii</i>	6	300
6	<i>Diaphanosoma sarsi</i>	18	900
7	<i>Bosmina longistris</i>	18	900
<b>Total</b>		<b>106</b>	<b>5300</b>

STATION 193			
Sampling Date: 26/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	1	10
2	<i>Acartia lausi</i>	5	50
3	<i>Oithona similis</i>	1	10
4	AT Bivalvia	1	10
<b>Total</b>		<b>8</b>	<b>80</b>

STATION 194			
Sampling Date: 26/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Eucalanus subcrassus giesb.</i>	2	100
2	Larva of shrimp	1	50
3	<i>Paracalanus crassirostris</i>	36	1800
4	<i>Temora burbinata dana</i>	1	50
5	<i>Acartia lausi</i>	7	350
6	<i>Acartia rythraea</i>	4	200
7	<i>Oithona similis</i>	5	250
8	<i>Oithona brevicornis</i>	2	100
9	<i>Thermocyclops yalinus</i>	5	250
10	<i>Clytemnestra cutellata</i>	1	50
11	<i>Diaphanosoma sarsi</i>	1	50
12	<i>Bosmina longistris</i>	5	250
<b>Total</b>		<b>70</b>	<b>3500</b>

STATION 195			
Sampling Date: 28/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Eucalanus subcrassus giesb.</i>	1	50
2	<i>Paracalanus parvus</i>	11	550
3	<i>Acartia lausi</i>	2	100
<b>Total</b>		<b>14</b>	<b>700</b>

STATION 196			
Sampling Date: 26/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Acartia lausi</i>	3	30
2	<i>Oithona similis</i>	1	10
3	AT Bivalvia	3	30
<b>Total</b>		<b>7</b>	<b>70</b>

STATION 197			
Sampling Date: 26/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	3	150
2	<i>Labidocera minuta</i>	1	50
3	<i>Acartia pacifica</i>	4	200
4	<i>Oithona similis</i>	1	50
5	<i>Microsetella norvegica</i>	2	100
<b>Total</b>		<b>11</b>	<b>550</b>

STATION 199			
Sampling Date: 28/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Penilia schmackeri</i>	1	50
2	<i>Paracalanus parvus</i>	4	200
3	<i>Acartia lausi</i>	3	150
4	<i>Acartia rythraea</i>	1	50
5	<i>Oithona similis</i>	15	750
6	<i>Oncaea venusta</i>	1	50
7	<i>Microsetella norvegica</i>	1	50
8	<i>Macrosetella gracilis</i>	12	600
9	<i>Copepoda</i>	6	300
<b>Total</b>		<b>44</b>	<b>2200</b>

STATION 199			
Sampling Date: 26/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Sagitta delicata</i>	1	50
2	<i>Canthocalanus pauper</i>	2	100
3	<i>Undinula vulgaris dana</i>	1	50
4	<i>Paracalanus parvus</i>	22	1100
5	<i>Labidocera roery</i>	1	50
6	<i>Acartia lausi</i>	9	450
7	<i>Oithona brevicornis</i>	1	50
8	<i>Oithona lumifera</i>	1	50
9	<i>Microsetella norvegica</i>	13	650
10	C. ton	1	50
<b>Total</b>		<b>50</b>	<b>2500</b>

STATION 201			
Sampling Date: 28/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	33	1650
2	<i>Acartia lausi</i>	4	200
3	<i>Oithona brevicornis</i>	10	500
4	<i>Microsetella norvegica</i>	5	250
5	<i>Euterpina acutifront</i>	1	50
6	<i>Luciferopus</i>	1	50
7	<i>Oikopleura dioica</i>	1	50
8	AT T m	2	100
<b>Total</b>		<b>57</b>	<b>2850</b>

STATION 202			
Sampling Date: 28/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus crassirostris</i>	58	5800
2	<i>Pseudodiaptomus marinus</i>	1	100
3	<i>Schmackeria cordioides</i>	1	100
4	<i>Labidocera minuta</i>	1	100
5	<i>Acartia lausi</i>	33	3300
6	<i>Oithona dana</i>	21	2100
7	<i>Oithona brevicornis</i>	6	600
8	<i>Oithona lumifera</i>	1	100
9	<i>Microsetella norvegica</i>	1	100
10	<i>Euterpina acutifront</i>	3	300
11	<i>Oikopleura dioica</i>	1	100
12	<i>Copepoda</i>	1	100
13	AT T m	1	100
<b>Total</b>		<b>129</b>	<b>12900</b>

STATION 203			
Sampling Date: 28/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Sagitta delicata</i>	1	100
2	<i>Paracalanus parvus</i>	197	19700
3	<i>Centropages curcatus</i>	1	100
4	<i>Acartia lausi</i>	6	600
5	<i>Oithona similis</i>	16	1600
6	<i>Oithona brevicornis</i>	1	100
7	<i>Macrosetella gracilis</i>	1	100
<b>Total</b>		<b>223</b>	<b>22300</b>

STATION 204			
Sampling Date: 28/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	8	400

STATION 28			
Sampling Date: 26/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	9	90
2	<i>Acartia lausi</i>	6	60
3	<i>Microsetella norvegica</i>	1	10
Total		16	160

STATION 29			
Sampling Date: 26/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	5	250
2	<i>Acartia lausi</i>	4	200
3	<i>Oithona similis</i>	1	50
4	<i>Corycaeus ahali</i>	2	100
5	<i>Microsetella norvegica</i>	18	900
6	<i>Macrosetella gracilis</i>	12	600
7	<i>Euterpina acutifront</i>	2	100
8	<i>AT bivalvia</i>	1	50
Total		45	2250

STATION 30			
Sampling Date: 27/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	5	500
2	<i>Schmackeria tubosa</i>	1	100
3	<i>Oithona similis</i>	1	100
4	<i>Oithona brevicornis</i>	2	200
5	<i>Oithona lumifera</i>	8	800
6	<i>Thermocyclops hyalinus</i>	16	1600
7	<i>Diaphanosoma sarsi</i>	2	200
8	<i>AT bivalvia</i>	1	100
9	<i>Mesocyclops sp.</i>	1	100
10	<i>Bosmina longistris</i>	1	100
Total		38	3800

STATION 31			
Sampling Date: 27/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	2	100
2	<i>Acartia lausi</i>	2	100
3	<i>Oithona similis</i>	6	300
4	<i>Corycaeus ahali</i>	1	50
5	<i>Microsetella norvegica</i>	1	50
6	<i>Oikopleura dioica</i>	1	50
Total		13	650

STATION 32			
Sampling Date: 27/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Sagitta delicata</i>	1	50
2	<i>Eucalanus subcrassus Giesb.</i>	1	50
3	<i>Paracalanus parvus</i>	4	200
4	<i>Labidocera minuta</i>	1	50
5	<i>Acartia lausi</i>	11	550
6	<i>Oithona allax</i>	1	50
7	<i>Oithona similis</i>	1	50
8	<i>Oncaea venusta</i>	1	50
Total		21	1050

STATION 33			
Sampling Date: 27/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	37	1850
2	<i>Schmackeria sp.</i>	1	50
3	<i>Acartia lausi</i>	4	200
4	<i>Oithona similis</i>	30	1500
5	<i>Oithona brevicornis</i>	39	1950
6	<i>Corycaeus ahali</i>	4	200
7	<i>Microsetella norvegica</i>	7	350
Total		122	6100

STATION 34			
Sampling Date: 27/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Sagitta delicata</i>	2	100
2	<i>Paracalanus parvus</i>	23	1150

2	<i>Acartia pacifica</i>	2	100
3	<i>Oncaea venusta</i>	2	100
4	<i>AT actinotrocha</i>	1	50
Total		13	650

STATION 30			
Sampling Date: 30/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Sagitta crassa</i>	1	50
2	<i>Acrocalanus gilber</i>	2	100
3	<i>Paracalanus parvus</i>	32	1600
4	<i>Acartia lausi</i>	2	100
5	<i>Oithona similis</i>	19	950
6	<i>Oithona brevicornis</i>	6	300
7	<i>Euterpina acutifront</i>	2	100
8	<i>Oikopleura dioica</i>	11	550
9	<i>AT gastropoda</i>	1	50
10	<i>Dyphyes sp.</i>	1	50
Total		77	3850

STATION 32			
Sampling Date: 29/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Eucalanus subcrassus Giesb.</i>	1	50
2	<i>Paracalanus parvus</i>	9	450
3	<i>Centropages curcatus</i>	1	50
4	<i>Acartia lausi</i>	1	50
5	<i>Acartia rhythroa</i>	6	300
6	<i>Oithona similis</i>	14	700
7	<i>Oithona brevicornis</i>	6	300
8	<i>Thermocyclops hyalinus</i>	1	50
9	<i>Moinadaphnia macleayi</i>	1	50
10	<i>Hyalocylis triata</i>	1	50
11	<i>Diaphanosoma sarsi</i>	1	50
12	<i>Creisesis sp.</i>	1	50
Total		37	1850

STATION 33			
Sampling Date: 26/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	30	1500
2	<i>Labidocera minuta</i>	1	50
3	<i>Acartia lausi</i>	3	150
4	<i>Oithona dana</i>	1	50
5	<i>Oithona similis</i>	5	250
Total		40	2000

STATION 37			
Sampling Date: 29/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	20	200
2	<i>AT bivalvia</i>	1	10
3	<i>Copepoda</i>	6	60
Total		27	270

STATION 34			
Sampling Date: 29/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Undinula vulgaris Dana</i>	1	100
2	<i>Eucalanus subcrassus Giesb.</i>	1	100
3	<i>Paracalanus parvus</i>	75	7500
4	<i>Oithona similis</i>	1	100
5	<i>Oithona brevicornis</i>	8	800
6	<i>Corycaeus ahali</i>	2	200
7	<i>Microsetella norvegica</i>	1	100
8	<i>AT bivalvia</i>	1	100
Total		90	9000

STATION 35			
Sampling Date: 29/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	46	2300
2	<i>Acartia rhythroa</i>	1	50
3	<i>Oithona similis</i>	12	600
4	<i>AT bivalvia</i>	1	50
Total		60	3000

STATION 36			
Sampling Date: 29/05/2006			
No	SPECIES	QUANTITY	INDS/M3

3	<i>Acartia pacifica</i>	6	300
4	<i>Oithona similis</i>	19	950
5	<i>Corycaeus ahli</i>	1	50
6	<i>Microsetella norvegica</i>	1	50
7	Copepod	1	50
	<b>Total</b>	<b>53</b>	<b>2650</b>

NO	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	15	1500
2	<i>Oithona similis</i>	14	1400
3	<i>Oithona brevicornis</i>	11	1100
4	Copepoda	12	1200
	<b>Total</b>	<b>52</b>	<b>5200</b>

STATION#15			
Sampling date: 27/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	12	600
2	<i>Oithona similis</i>	4	200
3	Copepoda	1	50
	<b>Total</b>	<b>17</b>	<b>850</b>

STATION#16			
Sampling date: 28/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	1	100
2	<i>Labidocera minuta</i>	1	100
3	<i>Acartia lausi</i>	3	300
4	<i>Oithona sp.</i>	3	300
5	<i>Corycaeus andrewsi</i>	1	100
6	<i>Corycaeus ahli</i>	3	300
7	<i>Microsetella norvegica</i>	18	1800
8	<i>Macrosetella gracilis</i>	24	2400
9	<i>Euterpina acutifront</i>	1	100
	<b>Total</b>	<b>55</b>	<b>5500</b>

STATION#17			
Sampling date: 28/05/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	7	350
2	<i>Labidocera minuta</i>	3	150
3	<i>Acartia lausi</i>	3	150
4	<i>Oithona similis</i>	14	700
5	<i>Corycaeus ahli</i>	4	200
6	<i>Microsetella norvegica</i>	1	50
7	<i>Oikopleura dioica</i>	3	150
8	AT Gastropoda	1	50
9	Copepoda	6	300
	<b>Total</b>	<b>42</b>	<b>2100</b>



Survey Institute of Marine Resources and Environment, August 2006 Phytoplankton Species

STATION#1			
Sampling Date: 8/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Melosira granulata	9120	182400
2	Thalassiosira spp.	3	60
3	Diploneis smithii	1	20
4	Pleurosigma sp.	1	20
5	Nitzschia sigma	26	520
6	Nitzschia sp.	2	40
7	Prorocentrum micans	1	20
8	Ceratium furca	2	40
9	Oscillatoria p2	10	200
10	Pediastrum sp.	7	140
11	Phacus sp.1	4	80
		9177	183540

STATION#2			
Sampling Date: 8/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Melosira granulata	5856	117120
2	Thalassionema nitzschioides	2	40
3	Thalassiothrix frauenfeldii	1	20
4	Nitzschia sigma	4	80
5	Oscillatoria p2	9	180
6	Anabaena sp	1	20
7	Pediastrum sp.	2	40
8	Scenedesmus spp.	8	160
9	Phacus sp.1	9	180
		5888	117760

STATION#3			
Sampling Date: 8/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Melosira granulata	3648	72960
2	Skeletonema costatum	10	200
3	Nitzschia longissima	2	40
4	Nitzschia sigma	20	400
5	Pseudonitzschia sp.1 (to)	3	60
6	Prorocentrum mexicanum	1	20
7	Oscillatoria p1	9	180
8	Oscillatoria p2	11	220
9	Pediastrum sp.	5	100
10	Scenedesmus spp.	184	3680
11	Phacus sp.1	3	60
12	Tioidoc	85	1700
		3976	79520

STATION#4			
Sampling Date: 8/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Melosira granulata	8736	174720
2	Melosira spp.	30	600
3	Thalassiosira spp.	2	40
4	Navicula palpebralis	1	20
5	Trachyneis sp.	1	20
6	Pleurosigma delicatum	1	20
7	Nitzschia borenziana	9	180
8	Nitzschia longissima	7	140
9	Nitzschia sigma	7	140
10	Nitzschia sigma antarcedens	2	40
11	Pseudonitzschia spp.	3	60
12	Oscillatoria p2	130	2600
13	Pediastrum sp.	6	120
14	Scenedesmus spp.	148	2960
		9089	181780

STATION#5			
Sampling Date: 8/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Melosira granulata	11488	229760
2	Melosira aerensis	17	340
3	Melosira spp.	32	640
4	Trachyneis sp.	23	460
5	Pleurosigma sp	1	20
6	Nitzschia sigma	6	120
7	Surirella sigma	1	20
8	Oscillatoria p2	35	700
9	Pediastrum sp.	5	100
10	Scenedesmus spp.	4	80
11	Glosterium sp	2	40
12	Phacus sp.1	2	40
		11616	232320

STATION#6

STATION#19			
Sampling Date: 9/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Thalassiosira spp.	6	240
2	Pleurosigma sp	1	40
3	Amphora quadrata	5	200
4	Nitzschia longissima reversa	8	320
5	Prorocentrum micans	79	3160
6	Prorocentrum sigmoides	2	80
7	Prorocentrum hathyrum	14	560
8	Dinophysis hudgei	14	560
9	Gymnodinium sp	5	200
10	Gymnodinium mikimotoi	17	680
11	Gyrodinium spirale	21	840
12	Ceratium furca	33	1320
13	Ceratium pusillus	3	120
14	Gonyaulax sp	1	40
15	Protoperidinium steinii	12	480
16	Protoperidinium pellucidum	11	440
17	Protoperidinium leonis	24	960
18	Protoperidinium sp.	16	640
19	Peridinium quinquecorne	1	40
20	Gyrodinium sp.	5	200
21	Alexandrium sp	4	160
22	Alexandrium pseudogonyaulax	18	720
23	Diplopsalis sp.	1	40
		665	26600

STATION#20			
Sampling Date: 20/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Asteromphalus leveanus	1	40
2	Thalassiosira spp.	17	680
3	Odontella mobilensis	0.5	20
4	Synedra pulchella	4	160
5	Pleurosigma angulatum	10	400
6	Pleurosigma oviculaceum	30	1200
7	Nitzschia longissima	2	80
8	Nitzschia sigma	2	80
9	Nitzschia sigma antarcedens	4	160
10	Nitzschia sp.	6	240
11	Prorocentrum micans	12	480
12	Prorocentrum mexicanum	46	1840
13	Prorocentrum gracile	32	1280
14	Dinophysis rotundata	2	80
15	Gonyaulax polygramma	1	40
16	Protoperidinium steinii	35	1400
17	Protoperidinium depressum	2	80
18	Protoperidinium pellucidum	11	440
19	Alexandrium spp.	17	680
20	Diplopsalis sp.	1	40
		241,5	9660

STATION#21			
Sampling Date: 20/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Thalassiosira spp.	1	20
2	Skeletonema costatum	26	520
3	Chaetoceros diversus	2	40
4	Navicula sp	1	20
5	Pleurosigma sp	62	1240
6	Nitzschia longissima	1	20
7	Nitzschia sigma	1	20
8	Prorocentrum micans	92	1840
9	Prorocentrum hathyrum	11	220
10	Dinophysis hudgei	1	20
11	Gymnodinium sp	5	100
12	Gyrodinium spirale	11	220
13	Polykrikos schwartzii	1	20
14	Ceratium furca	1	20
15	Protoperidinium steinii	41	820
16	Protoperidinium pellucidum	6	120
17	Protoperidinium sp.	13	260
18	Peridinium quinquecorne	1	20
19	Nitzschia sp.	1	20
20	Scrippsiella sp.	1	20
21	Alexandrium spp.	6	120
22	Alexandrium pseudogonyaulax	15	300
23	Pyrophaucus sp	25	500
24	Hermesium sp.	6	120
		334	6680

STATION#22

Station#7 Sampling date: 18/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Melosira granulata	115	2300
2	Melosira uegensis	2	40
3	Skeletonema costatum	10	200
4	Nitzschia paleobralis	1	20
5	Diploneis mithis	1	20
6	Gyrosigma strigile	1	20
7	Amphiproraolata	1	20
8	Nitzschia longissima	3	60
9	Nitzschia sigma	2	40
10	Nitzschia sigma intercedens	2	40
11	Protoperidinium sp.	4	80
12	Oscillatoria sp2	7	140
13	Pediastrum sp.	2	40
14	Scenedesmus sp.	54	1080
		<b>230</b>	<b>4600</b>

Station#7 Sampling date: 18/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Melosira granulata	8256	165120
2	Melosira uegensis	2	40
3	Thalassiosira spp.	2	40
4	Skeletonema costatum	6	120
5	Diatoma longatum	4	80
6	Navicula paleobralis	10	200
7	Trachyneis aspera	2	40
8	Pleurosigma aviculaceum	2	40
9	Pleurosigma elagicum	1	20
10	Nitzschia corenziana	1	20
11	Nitzschia longissima	13	260
12	Nitzschia sigma	94	1880
13	Nitzschia sigma intercedens	10	200
14	Pseudonitzschia sp. (ta)	13	260
15	Oscillatoria sp2	29	580
16	Pediastrum sp.	2	40
17	Scenedesmus sp.	10	200
18	Scenedesmus sp.	32	640
19	Phacus sp1.	2	40
		<b>8532</b>	<b>170640</b>

Station#8 Sampling date: 18/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Melosira granulata	1728	34560
2	Melosira uegensis	93	1860
3	Thalassiosira spp.	5	100
4	Navicula paleobralis	5	100
5	Pleurosigma spp.	3	60
6	Pleurosigma spp.	2	40
7	Nitzschia corenziana	1	20
8	Nitzschia longissima	6	120
9	Nitzschia sigma	15	300
10	Nitzschia sigma intercedens	3	60
11	Pseudonitzschia sp.	1	20
12	Protoperidinium sp.	1	20
13	Oscillatoria sp2	6	120
14	Pediastrum sp.	10	200
15	Scenedesmus sp.	90	1800
16	Tetrachlorella umbricus var. piculatus	24	480
		<b>2003</b>	<b>40060</b>

Station#9 Sampling date: 18/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Melosira granulata	744	14880
2	Coscinodiscus subutilis	1	20
3	Thalassiosira spp.	4	80
4	Diatoma longatum	2	40
5	Grammatophora marina	2	40
6	Navicula paleobralis	3	60
7	Navicula sp.	1	20
8	Pleurosigma sp.	4	80
9	Pleurosigma aviculaceum	1	20
10	Pleurosigma elagicum	2	40
11	Amphiprora quadrata	3	60
12	Nitzschia sigma	11	220
13	Surirella tenera var. servosa	1	20
14	Campylodiscus longulatum	1	20
15	Protoperidinium sp.	6	120
16	Oscillatoria sp2	8	160
17	Pediastrum sp.	8	160
18	Scenedesmus sp.	120	2400
19	Closterium sp.	2	40
20	Phacus sp1.	1	20
		<b>928</b>	<b>18560</b>

Station#10 Sampling date: 20/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Thalassiosira spp.	1	40
2	Skeletonema costatum	10	400
3	Synedra sp.	2	80
4	Grammatophora marina	1	40
5	Navicula sp.	1	40
6	Pleurosigma angulatum	4	160
7	Pleurosigma sp.	15	600
8	Nitzschia corenziana	2	80
9	Nitzschia longissima	5	200
10	Nitzschia longissima reversa	45	1800
11	Nitzschia sigma	19	760
12	Nitzschia closterium	1	40
13	Prorocentrum nicanans	2	80
14	Prorocentrum hathyrum	5	200
15	Gymnodinium sanguineum	3	120
16	Polykrikos schwartzii	0,5	20
17	Gonyaulax sp.	7	280
18	Gonyaulax sp.	1	40
19	Protoperidinium steinii	2	80
20	Protoperidinium pellucidum	3	120
21	Protoperidinium sp.	6	240
22	Peridinium sp.	3	120
23	Alexandrium sp.	1	40
24	Alexandrium pseudogonyaulax	8	320
25	Obolus sp.	8	320
26	Pyrophacus sp.	7	280
27	Oscillatoria sp2	2	80
28	Anabaena sp.	3	120
29	Tetrahymena	112	4480
		<b>280,5</b>	<b>11220</b>

Station#13 Sampling date: 20/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Thalassiosira spp.	2	40
2	Skeletonema costatum	73	1460
3	Chaetoceros sp.	4	80
4	Pleurosigma sp.	12	240
5	Nitzschia longissima	1	20
6	Nitzschia longissima reversa	5	100
7	Nitzschia sigma	7	140
8	Prorocentrum nicanans	7	140
9	Prorocentrum hathyrum	21	420
10	Prorocentrum sp.	1	40
11	Dinophysis judayi	1	40
12	Gymnodinium sanguineum	1	40
13	Polykrikos schwartzii	1	20
14	Ceratum furca	1	20
15	Gonyaulax sp.	16	320
16	Gonyaulax sp.	2	40
17	Gonyaulax rotundata	1	20
18	Protoperidinium pellucidum	1	20
19	Protoperidinium sp.	29	580
20	Peridinium quinquecarne	1	20
21	Obolus sp.	10	200
22	Pyrophacus sp.	11	220
23	Hermesium sp.	2	40
24	Oscillatoria sp2	2	40
25	Tetrahymena	235	4700
		<b>452</b>	<b>9040</b>

Station#14 Sampling date: 20/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Skeletonema costatum	44	1760
2	Thalassiosira frauenfeldii	1	40
3	Navicula sp.	0,5	20
4	Diploneis sp.1	1	40
5	Pleurosigma sp.	8	320
6	Nitzschia corenziana	1	40
7	Nitzschia longissima	1	40
8	Nitzschia longissima reversa	8	320
9	Nitzschia sigma	1	40
10	Prorocentrum nicanans	2	80
11	Prorocentrum sp.	9	360
12	Dinophysis judayi	1	40
13	Polykrikos schwartzii	2	80
14	Gonyaulax sp.	12	480
15	Gonyaulax sp.	1	40
16	Gonyaulax sp.	1	40
17	Protoperidinium pellucidum	1	40
18	Protoperidinium sp.	43	1720
19	Peridinium quinquecarne	4	160
20	Peridinium sp.	1	40
21	Alexandrium pseudogonyaulax	11	440
22	Fragilidium sp.	1	40

STATION#10			
Sampling Date: 19/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Melosira granulata	6	120
2	Melosira granulata angustissima	121	2420
3	Thalassiosira spp.	16	320
4	Skeletonema costatum	27	540
5	Leptocylindrus dianicus	7	140
6	Navicula palpebralis	5	100
7	Nitzschia longissima	261	5220
8	Nitzschia sigma	2160	43200
9	Nitzschia sigma intercedens	1	20
11	Oscillatoria p2	3	60
12	Pediastrum sp.	4	80
13	Scenedesmus spp.	66	1320
14	Phacus sp1.	1	20
		<b>2692</b>	<b>53840</b>

STATION#11			
Sampling Date: 19/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Melosira granulata	116	2320
2	Thalassiosira spp.	11	220
3	Skeletonema costatum	3	60
4	Diatoma longatum	3	60
5	Nitzschia longissima	108	2160
6	Nitzschia sigma	688	13760
7	Pseudonitzschia spp.	1	20
8	Oscillatoria p2	5	100
9	Pediastrum sp.	8	160
10	Scenedesmus spp.	108	2160
11	Phacus sp1.	1	20
		<b>1079</b>	<b>21580</b>

STATION#12			
Sampling Date: 19/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Melosira granulata	31	620
2	Thalassiosira spp.	12	240
3	Skeletonema costatum	4	80
4	Diatoma longatum	5	100
5	Synedra pulchella	1	20
6	Synedra aillonii	1	20
7	Nitzschia longissima	2	40
8	Nitzschia sigma	7	140
9	Pseudonitzschia spp.	11	220
10	Alexandrium sp.	1	20
11	Oscillatoria p2	4	80
12	Anabaena sp.	3	60
13	Pediastrum sp.	8	160
14	Scenedesmus spp.	160	3200
15	Phacus sp1	1	20
16	Gloeotila spp	57	1140
		<b>310</b>	<b>6200</b>

STATION#13			
Sampling Date: 19/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Melosira granulata	97	1940
2	Thalassiosira spp.	23	460
3	Skeletonema costatum	12	240
4	Diatoma longatum	1	20
5	Nitzschia longissima	324	6480
6	Nitzschia sigma	1	20
7	Pseudonitzschia spp.	1	20
8	Protoperidinium spp.	3	60
9	Alexandrium sp.	1	20
10	Oscillatoria p2	11	220
11	Anabaena sp.	3	60
12	Scenedesmus spp.	60	1200
		<b>732</b>	<b>14640</b>

STATION#14			
Sampling Date: 19/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Melosira granulata	158	3160
2	Thalassiosira spp.	7	140
3	Skeletonema costatum	8	160
4	Biculophora pulchella	1	20
5	Diatoma longatum	5	100
6	Thalassiothrix frauenfeldii	1	20
7	Synedra aillonii	1	20
8	Trachyneis sp.	1	20
9	Pleurosigma normanii	1	20
10	Nitzschia longissima	1	20
11	Gymnodinium sanguineum	1	20

23	Oblea sp	25	1000
24	Pyrophacus sp	5	200
25	Hermesium sp.	2	80
26	Tiham	54	2160
		<b>244,5</b>	<b>9780</b>

STATION#25			
Sampling Date: 21/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Thalassiosira spp.	1	40
2	Skeletonema costatum	62	2480
3	Pleurosigma sp.	2	80
5	Prorocentrum micans	1	40
6	Prorocentrum minimum	1	40
7	Prorocentrum thalium	1	40
9	Ceratium furca	0,5	20
10	Gonyaulax sp	2	80
11	Protoperidinium pellucidum	1	40
12	Protoperidinium sp.	7	280
13	Peridinium quinquecorne	1	40
14	Alexandrium sp.	1	40
15	Alexandrium gonyaulax	5	200
17	Pyrophacus sp	21	840
18	Hermesium sp.	1	40
19	Anabaena sp.	9	360
20	Tiham	244	9760
		<b>384,5</b>	<b>15380</b>

STATION#26			
Sampling Date: 21/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Skeletonema costatum	58	2320
2	Chaetoceros sp.	5	200
3	Synedra sp.	2	80
4	Pleurosigma sp.	2	80
5	Nitzschia longissima reversa	1	40
7	Protoperidinium onicum	1	40
8	Protoperidinium sp.	4	160
9	Peridinium quinquecorne	6	240
11	Diplopsalis sp.	1	40
12	Oblea sp	3	120
13	Pyrophacus sp	4	160
14	Hermesium sp.	2	80
15	Anabaena sp	4	80
16	Pediastrum sp.	1	40
17	Scenedesmus sp.	12	480
18	Tiham	214	8560
		<b>338</b>	<b>13520</b>

STATION#27			
Sampling Date: 21/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Skeletonema costatum	39	1560
2	Chaetoceros sp.	4	160
3	Gyrodinium aureolum	1	40
4	Pleurosigma sp.	3	120
5	Nitzschia longissima	3	120
6	Nitzschia longissima reversa	1	40
7	Nitzschia sigma	1	40
9	Prorocentrum micans	3	120
10	Prorocentrum thalium	2	80
11	Gonyaulax sp	4	160
12	Protoperidinium sp.	1	40
13	Peridinium sp.	0,5	20
14	Alexandrium sp	3	120
15	Fragilidium sp.	1	40
16	Oblea sp	7	280
17	Pyrophacus sp	6	240
18	Hermesium sp.	3	120
19	Oscillatoria sp1	3	120
20	Anabaena sp	17	680
21	Tiham	261	10440
		<b>366</b>	<b>14640</b>

STATION#28			
Sampling Date: 21/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	Skeletonema costatum	95	1900
2	Synedra sp.	2	40
3	Pleurosigma sp.	2	40
4	Pleurosigma aviculaceum	1	20



12	<i>Protoperidinium</i> sp.	3	60
13	<i>Oscillatoria</i> sp.2	15	300
14	<i>Spirulina</i> sp.	1	120
15	<i>Anabaena</i> sp.	6	120
16	<i>Pediastrum</i> sp.	1	20
17	<i>Scenedesmus</i> sp.	30	600
18	<i>Phacus</i> sp.1	1	20
		<b>248</b>	<b>4960</b>

STATION#15			
Sampling date: 9/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira</i> sp. <i>granulata</i>	176	3520
2	<i>Thalassiosira</i> sp.	14	280
3	<i>Diatoma</i> sp. <i>longatum</i>	2	40
4	<i>Nitzschia</i> sp. <i>longissima</i>	7	140
5	<i>Nitzschia</i> sp. <i>sigma</i>	15	300
6	<i>Nitzschia</i> sp.	5	120
7	<i>Oscillatoria</i> sp.2	14	280
8	<i>Spirulina</i> sp.	5	100
9	<i>Anabaena</i> sp.	9	180
10	<i>Pediastrum</i> sp.	8	160
11	<i>Scenedesmus</i> sp.	1	20
12	<i>Phacus</i> sp.1	8	160
		<b>345</b>	<b>6900</b>

STATION#16			
Sampling date: 9/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassiosira</i> sp.	49	1960
2	<i>Skeletonema</i> sp. <i>ostatum</i>	8	320
3	<i>Pleurosigma</i> sp. <i>elagicum</i>	1	40
4	<i>Nitzschia</i> sp. <i>longissima</i>	1	40
5	<i>Nitzschia</i> sp. <i>sigma</i>	1	40
6	<i>Prorocentrum</i> sp. <i>americanus</i>	0,5	20
7	<i>Prorocentrum</i> sp. <i>hathynum</i>	34	1360
8	<i>Prorocentrum</i> sp. <i>racile</i>	1	40
9	<i>Gymnodinium</i> sp.	3	120
10	<i>Gymnodinium</i> sp. <i>conguineum</i>	3	120
11	<i>Protoperidinium</i> sp. <i>pellucidum</i>	4	160
12	<i>Protoperidinium</i> sp.	6	240
13	<i>Scenedesmus</i> sp.	81	1620
14	<i>Oscillatoria</i> sp.2	30	1200
15	<i>Spirulina</i> sp.	5	200
16	<i>Anabaena</i> sp.	5	200
17	<i>Nham</i> sp. <i>indico</i>	103	4120
		<b>259,5</b>	<b>10380</b>

STATION#17			
Sampling date: 9/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira</i> sp. <i>granulata</i>	3	60
2	<i>Thalassiosira</i> sp.	67	1340
3	<i>Skeletonema</i> sp. <i>ostatum</i>	4	80
4	<i>Nitzschia</i> sp. <i>longissima</i>	3	60
5	<i>Nitzschia</i> sp. <i>sigma</i>	7	140
6	<i>Pseudonitzschia</i> sp.	1	20
7	<i>Prorocentrum</i> sp. <i>hathynum</i>	3	60
8	<i>Alexandrium</i> sp.	13	260
9	<i>Oscillatoria</i> sp.2	99	1980
10	<i>Spirulina</i> sp.	14	280
11	<i>Anabaena</i> sp.	17	340
12	<i>Scenedesmus</i> sp.	8	160
13	<i>Nham</i> sp. <i>indico</i>	101	2020
		<b>342</b>	<b>6840</b>

STATION#18			
Sampling date: 9/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira</i> sp. <i>granulata</i> var. <i>angustissima</i>	10	200
2	<i>Skeletonema</i> sp. <i>ostatum</i>	8	160
3	<i>Synedra</i> sp.	2	40
4	<i>Pleurosigma</i> sp.	1	20
5	<i>Nitzschia</i> sp. <i>longissima</i> var. <i>reversa</i>	14	80
6	<i>Nitzschia</i> sp.	4	80
7	<i>Peridinium</i> sp. <i>quaquecome</i>	1	20
8	<i>Anabaena</i> sp.	780	15600
9	<i>Microcystis</i> sp.	16	320
10	<i>Scenedesmus</i> sp. <i>quadricauda</i>	16	320
11	<i>Scenedesmus</i> sp. <i>armatus</i>	400	8000
12	<i>Scenedesmus</i> sp. <i>cuminatus</i> var. <i>cuminatus</i>	8	160
13	<i>Phacus</i> sp.2	1	20
14	<i>Phacus</i> sp.1	1	20
		<b>1308</b>	<b>26160</b>

5	<i>Nitzschia</i> sp. <i>longissima</i>	2	40
6	<i>Nitzschia</i> sp. <i>sigma</i> var. <i>intercedens</i>	1	20
7	<i>Prorocentrum</i> sp. <i>hathynum</i>	1	20
8	<i>Anabaenopsis</i> sp. <i>aciborski</i>	2150	43000
9	<i>Pediastrum</i> sp.	2	40
10	<i>Scenedesmus</i> sp. <i>arinatus</i>	8	160
11	<i>Scenedesmus</i> sp. <i>quadricauda</i>	4	80
12	<i>Scenedesmus</i> sp. <i>cuminatus</i> var. <i>cuminatus</i>	20	400
13	<i>Staurastrum</i> sp.	1	20
14	<i>Thiodam</i>	15	300
		<b>2304</b>	<b>46080</b>

STATION#29			
Sampling date: 11/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira</i> sp. <i>granulata</i> var. <i>angustissima</i>	294	5880
2	<i>Skeletonema</i> sp. <i>ostatum</i>	525	10500
3	<i>Synedra</i> sp.	2	40
4	<i>Navicula</i> sp.	1	40
5	<i>Nitzschia</i> sp. <i>sigma</i>	1	20
6	<i>Nitzschia</i> sp.	2	40
7	<i>Peridinium</i> sp.	2	40
8	<i>Prorocentrum</i> sp.	1	20
9	<i>Anabaenopsis</i> sp.	510	10200
10	<i>Pediastrum</i> sp.	5	100
11	<i>Scenedesmus</i> sp.	8	160
12	<i>Scenedesmus</i> sp. <i>arinatus</i>	8	160
13	<i>Scenedesmus</i> sp. <i>quadricauda</i>	12	240
14	<i>Scenedesmus</i> sp. <i>cuminatus</i> var. <i>cuminatus</i>	16	320
15	<i>Staurastrum</i> sp.	1	20
		<b>1395</b>	<b>27900</b>

STATION#30			
Sampling date: 11/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira</i> sp. <i>granulata</i> var. <i>angustissima</i>	109	4360
2	<i>Coscinodiscus</i> sp.	0,5	20
3	<i>Skeletonema</i> sp. <i>ostatum</i>	18	720
4	<i>Synedra</i> sp.	4	160
5	<i>Pleurosigma</i> sp.	1	40
6	<i>Nitzschia</i> sp. <i>sigma</i>	2	80
7	<i>Peridinium</i> sp.	6	240
8	<i>Oscillatoria</i> sp.1	1	40
9	<i>Anabaenopsis</i> sp. <i>aciborski</i>	547,5	21900
10	<i>Microcystis</i> sp.	1	40
11	<i>Pediastrum</i> sp.	3	120
12	<i>Scenedesmus</i> sp. <i>arinatus</i>	4	160
13	<i>Scenedesmus</i> sp. <i>quadricauda</i>	8	320
14	<i>Scenedesmus</i> sp. <i>cuminatus</i> var. <i>cuminatus</i>	8	320
15	<i>Staurastrum</i> sp.	1	20
16	<i>Thiodam</i>	2	80
		<b>719</b>	<b>28760</b>

STATION#31			
Sampling date: 11/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Skeletonema</i> sp. <i>ostatum</i>	141	2820
2	<i>Thalassiothrix</i> sp. <i>rauenfeldii</i>	1	20
3	<i>Pleurosigma</i> sp.	6	120
4	<i>Nitzschia</i> sp. <i>lorenziana</i>	1	20
5	<i>Nitzschia</i> sp. <i>longissima</i> var. <i>reversa</i>	5	120
6	<i>Nitzschia</i> sp. <i>sigma</i> var. <i>intercedens</i>	4	80
7	<i>Oscillatoria</i> sp.1	3	60
8	<i>Anabaenopsis</i> sp.	2001	40020
		<b>2161</b>	<b>43220</b>

STATION#32			
Sampling date: 12/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Skeletonema</i> sp. <i>ostatum</i>	70	1400
2	<i>Pleurosigma</i> sp.	23	460
3	<i>Nitzschia</i> sp. <i>longissima</i>	1	20
4	<i>Nitzschia</i> sp. <i>longissima</i> var. <i>reversa</i>	29	580
5	<i>Nitzschia</i> sp. <i>sigma</i>	1	20
6	<i>Prorocentrum</i> sp. <i>americanus</i>	6	120
7	<i>Prorocentrum</i> sp. <i>hathynum</i>	2	40
8	<i>Gonyaulax</i> sp.	1	20
9	<i>Gonyaulax</i> sp. <i>acrior</i>	1	20
10	<i>Gonyaulax</i> sp. <i>rotundata</i>	2	40
11	<i>Protoperidinium</i> sp. <i>steinii</i>	7	140
12	<i>Protoperidinium</i> sp. <i>pellucidum</i>	3	60
13	<i>Peridinium</i> sp.	12	240
14	<i>Alexandrium</i> sp.	1	20
15	<i>Alexandrium</i> sp.	1	20
16	<i>Alexandrium</i> sp. <i>pseudogonyaulax</i>	1	20
17	<i>Diplopsalis</i> sp.	1	20
18	<i>Obolus</i> sp.	1	20

19	<i>Pyrophacus</i> sp	28	560
20	<i>Hermesium</i> sp	3	60
21	<i>Oscillatoria</i> sp1	1	20
22	<i>Euglena</i> sp1	1	20
23	Total	77	1540
		<b>280</b>	<b>5600</b>

STATION K33			
Sampling date: 22/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Skeletonema</i> oostatum	50	1000
2	<i>Chaetoceros</i> sp	7	140
3	<i>Synedra</i> sp	1	20
4	<i>Achnanthes</i> longipes	1	20
5	<i>Pleurosigma</i> sp	1	20
6	<i>Nitzschia</i> longissima	8	160
7	<i>Nitzschia</i> longissima, reversa	6	120
8	<i>Nitzschia</i> sigma	3	60
9	<i>Protoperdinium</i> sp	1	20
10	<i>Alexandrium</i> pseudogonyaulax	2	40
11	<i>Pyrophacus</i> sp	5	100
12	<i>Hermesium</i> sp	1	20
13	Total	2	40
		<b>107</b>	<b>2140</b>

STATION L35			
Sampling date: 22/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Skeletonema</i> oostatum	53	1060
2	<i>Chaetoceros</i> sp	47	940
3	<i>Synedra</i> sp	6	120
4	<i>Grammatophara</i> marina	13	260
5	<i>Cocconeis</i> scutellum	1	20
6	<i>Pleurosigma</i> affine	2	40
7	<i>Nitzschia</i> corenziana	1	20
8	<i>Nitzschia</i> longissima	1	20
9	<i>Nitzschia</i> longissima, reversa	34	680
10	<i>Nitzschia</i> sigma	7	140
11	<i>Gymnodinium</i> sanguineum	1	20
12	<i>Ceratium</i> furca	13	260
13	<i>Gonyaulax</i> sp	2	40
14	<i>Gonyaulax</i> rotundata	5	100
15	<i>Protoperdinium</i> laudicans	1	20
16	<i>Protoperdinium</i> pellucidum	2	40
17	<i>Protoperdinium</i> sp	2	40
18	<i>Peridinium</i> quinquecorne	1	20
19	<i>Peridinium</i> sp	18	360
20	<i>Alexandrium</i> sp	2	40
21	<i>Alexandrium</i> pseudogonyaulax	2	40
22	<i>Pyrophacus</i> sp	1	20
23	<i>Oscillatoria</i> sp2	4	80
24	<i>Anabaenopsis</i> sp	14	280
25	Total	1	20
		<b>239</b>	<b>4780</b>

STATION L36			
Sampling date: 22/08/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira</i> sp	17	340
2	<i>Thalassiosira</i> sp	26	520
3	<i>Skeletonema</i> oostatum	2	40
4	<i>Guinardia</i> laccida	2	40
5	<i>Biddulphia</i> tubia	1	20
6	<i>Ditylum</i> mol	1	20
7	<i>Diatoma</i> longatum	5	100
8	<i>Thalassionema</i> nitzschioides	5	100
9	<i>Thalassiothrix</i> frauenfeldii	4	80
10	<i>Navicula</i> myra	2	40
11	<i>Trachyneis</i> spera	8	160
12	<i>Diploneis</i> bombus	1	20
13	<i>Gyrosigma</i> trigile	3	60
14	<i>Gyrosigma</i> sp	11	220
15	<i>Pleurosigma</i> affine	11	220
16	<i>Pleurosigma</i> angulatum	3	60
17	<i>Pleurosigma</i> oviculaceum	25	500
18	<i>Pleurosigma</i> pelagicum	2	40
19	<i>Amphiproprata</i> lata	69	1380
20	<i>Nitzschia</i> corenziana	8	160
21	<i>Nitzschia</i> longissima	45	900
22	<i>Nitzschia</i> longissima, intercedens	45	900
23	<i>Bacillaria</i> axillifera	10	200
24	<i>Pseudonitzschia</i> sp	1	20
25	<i>Campylodiscus</i> undulatus	2	40
26	<i>Prorocentrum</i> laudicans	1	20
27	<i>Protoperdinium</i> pellucidum	2	40
		<b>343</b>	<b>6860</b>

**Survey Institute of Marine Resources and Environment, August 2006 Zooplankton Species**

STATION 11			
Sampling Date: 18/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	8	400
2	<i>Oithona similis</i>	22	1100
3	<i>Copepoda</i>	12	600
4	<i>Microcyclops varicans</i>	9	450
		<b>51</b>	<b>2550</b>

STATION 12			
Sampling Date: 18/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	18	900
2	<i>Acartia lausi</i>	4	200
3	<i>Oithona similis</i>	50	2500
4	<i>Copepoda</i>	138	6900
		<b>210</b>	<b>10500</b>

STATION 13			
Sampling Date: 18/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	2	100
2	<i>Thermocyclops hyalinus</i>	12	600
3	<i>Microsetella norvegica</i>	2	100
4	<i>Copepoda</i>	186	9300
		<b>202</b>	<b>10100</b>

STATION 14			
Sampling Date: 18/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	22	1100
2	<i>Oithona sp.</i>	7	350
3	<i>Microsetella norvegica</i>	1	50
4	<i>Copepoda</i>	12	600
		<b>42</b>	<b>2100</b>

STATION 15			
Sampling Date: 18/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus crassirostris</i>	12	600
2	<i>Oithona similis</i>	23	1150
3	<i>Macrosetella gracilis</i>	1	50
4	<i>Euterpina acutifront</i>	1	50
5	<i>Copepoda</i>	6	300
6	<i>Moinadaphnia nacleayii</i>	2	100
		<b>45</b>	<b>2250</b>

STATION 16			
Sampling Date: 18/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	2	100
2	<i>Calocalanus pavo</i>	1	50
3	<i>Pseudodiaptomus sp.</i>	1	50
4	<i>Scolecithrix sp.</i>	1	50
5	<i>Acartia pacifica</i>	3	150
6	<i>Acartia lausi</i>	1	50
7	<i>Oithona similis</i>	6	300
8	<i>Corycaeus sp.</i>	1	50
9	<i>Clytemnestra scutellata</i>	2	100
10	<i>Copepoda</i>	118	5900
		<b>136</b>	<b>6800</b>

STATION 17			
Sampling Date: 18/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	12	600

STATION 19			
Sampling Date: 19/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Harpacticoda</i>	1	50
2	<i>Paracalanus parvus</i>	4	200
3	<i>Labidocera kroeyeri</i>	3	150
4	<i>Acartia lausi</i>	10	500
5	<i>Thermocyclops hyalinus</i>	2	100
6	<i>Oncaea venusta</i>	2	100
7	<i>Clytemnestra scutellata</i>	2	100
8	<i>Copepoda</i>	6	300
		<b>30</b>	<b>1500</b>

STATION 20			
Sampling Date: 20/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	108	43200
2	<i>Temora turbinata Dana</i>	3	1200
3	<i>Acartia lausi</i>	62	24800
4	<i>Oithona similis</i>	6	2400
5	<i>Copepoda</i>	300	120000
		<b>479</b>	<b>191600</b>

STATION 21			
Sampling Date: 20/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus sp.</i>	96	4800
2	<i>Copepoda</i>	270	13500
		<b>366</b>	<b>18300</b>

STATION 22			
Sampling Date: 20/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Harpacticoda</i>	6	600
2	<i>Acartia sp.</i>	36	3600
3	<i>Oithona sp.</i>	72	7200
4	<i>Copepoda</i>	256	25600
		<b>370</b>	<b>37000</b>

STATION 24			
Sampling Date: 20/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus sp.</i>	54	10800
2	<i>Acartia sp.</i>	23	4600
3	<i>Oithona sp.</i>	36	7200
4	<i>Copepoda</i>	224	44800
		<b>337</b>	<b>67400</b>

STATION 26			
Sampling Date: 21/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Harpacticoda</i>	6	300
2	<i>Paracalanus sp.</i>	12	600
3	<i>Copepoda</i>	250	12500
		<b>268</b>	<b>13400</b>

STATION 27			
Sampling Date: 21/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus sp.</i>	17	850
2	<i>Acartia lausi</i>	2	100
3	<i>Oithona sp.</i>	12	600
4	<i>Copepoda</i>	18	900
		<b>49</b>	<b>2450</b>

STATION 28			
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2	<i>Oithona similis</i>	39	1950
3	<i>Thermocyclops hyalinus</i>	6	300
4	<i>Microcetella norvegica</i>	4	200
5	<i>Clytemnestra scutellata</i>	13	650
6	<i>Copepoda</i>	36	1800
7	<i>Microcyclops varicans</i>	1	50
		<b>111</b>	<b>5550</b>

STATION 8			
Sampling date: 08/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Evadne nordmani</i>	2	100
2	<i>Paracalanus parvus</i>	2	100
3	<i>Oithona similis</i>	15	750
4	<i>Euterpina cutifront</i>	2	100
5	<i>Copepoda</i>	60	3000
6	<i>Microcyclops varicans</i>	1	50
		<b>82</b>	<b>4100</b>

STATION 9			
Sampling date: 08/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	9	450
2	<i>Oithona sp.</i>	15	750
3	<i>Copepoda</i>	156	7800
		<b>180</b>	<b>9000</b>

STATION 10			
Sampling date: 09/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Oithona similis</i>	18	900
2	<i>Thermocyclops hyalinus</i>	1	50
3	<i>Clytemnestra scutellata</i>	1	50
4	<i>Copepoda</i>	1	50
		<b>21</b>	<b>1050</b>

STATION 11			
Sampling date: 09/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Pseudodiaptomus sp.</i>	11	550
2	<i>Acartia lausi</i>	12	600
3	<i>Oithona sp.</i>	60	3000
4	<i>Thermocyclops hyalinus</i>	6	300
5	<i>Copepoda</i>	60	3000
		<b>149</b>	<b>7450</b>

STATION 12			
Sampling date: 09/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	4	200
2	<i>Acartia lausi</i>	1	50
3	<i>Oithona sp.</i>	14	700
4	<i>Copepoda</i>	10	500
		<b>29</b>	<b>1450</b>

STATION 13			
Sampling date: 09/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	4	200
2	<i>Oithona sp.</i>	20	1000
3	<i>Thermocyclops hyalinus</i>	3	150
4	<i>Clytemnestra scutellata</i>	1	50
5	<i>Copepoda</i>	12	600
6	<i>ATM</i>	1	50
		<b>41</b>	<b>2050</b>

STATION 14			
Sampling date: 09/08/2006			
No	SPECIES	QUANTITY	INDS/M3

Sampling date: 01/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Conchoecia sp.</i>	1	50
2	<i>Paracalanus sp.</i>	12	600
3	<i>Pseudodiaptomus sp.</i>	10	500
4	<i>Acartia lausi</i>	2	100
5	<i>Oncaea venusta</i>	3	150
6	<i>Macrosetella gracilis</i>	2	100
		<b>30</b>	<b>1500</b>

STATION 29			
Sampling date: 01/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus crassirostris</i>	48	4800
2	<i>Pseudodiaptomus sp.</i>	2	200
3	<i>Acartia lausi</i>	6	600
4	<i>Oithona similis</i>	82	8200
5	<i>Oithona revicornis</i>	4	400
		<b>142</b>	<b>14200</b>

STATION 30			
Sampling date: 01/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Conchoecia imbricata</i>	6	600
2	<i>Paracalanus sp.</i>	6	600
3	<i>Pseudodiaptomus sp.</i>	6	600
4	<i>Oithona sp.</i>	15	1500
5	<i>Thermocyclops hyalinus</i>	3	300
6	<i>Copepoda</i>	76	7600
		<b>112</b>	<b>11200</b>

STATION 32			
Sampling date: 01/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	66	3300
2	<i>Acartia lausi</i>	42	2100
3	<i>Oithona similis</i>	180	9000
4	<i>Copepoda</i>	514	25700
5	<i>Moinadaphnia nacleayii</i>	2	100
		<b>804</b>	<b>40200</b>

STATION 33			
Sampling date: 02/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus sp.</i>	37	14800
2	<i>Acartia sp.</i>	18	7200
3	<i>Oithona sp.</i>	12	4800
4	<i>Copepoda</i>	558	223200
		<b>625</b>	<b>250000</b>

STATION 37			
Sampling date: 02/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus sp.</i>	6	600
2	<i>Oithona sp.</i>	1	100
3	<i>Oncaea venusta</i>	6	600
4	<i>Euterpina cutifront</i>	6	600
5	<i>Copepoda</i>	105	10500
		<b>124</b>	<b>12400</b>

STATION 34			
Sampling date: 02/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus sp.</i>	194	38800
2	<i>Acartia sp.</i>	36	7200
3	<i>Oithona sp.</i>	30	6000
		<b>260</b>	<b>52000</b>

NO	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> sp.	1	50
2	<i>Oithona</i> sp.	5	250
3	<i>Microsetella</i> sp.	1	50
4	<i>Copepoda</i>	6	300
		<b>13</b>	<b>650</b>

STATION 15			
Sampling Date: 09/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> sp.	40	4000
2	<i>Acartia</i> sp.	6	600
3	<i>Oithona</i> sp.	26	2600
4	<i>Thermocyclops</i> sp.	6	600
5	<i>Copepoda</i>	230	23000
		<b>308</b>	<b>30800</b>

STATION 16			
Sampling Date: 09/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> sp.	50	500
3	<i>Harpacticoda</i>	6	60
4	<i>Copepoda</i>	50	500
		<b>106</b>	<b>1060</b>

STATION 17			
Sampling Date: 09/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Conchoecia</i> sp.	1	50
2	<i>Harpacticoda</i>	8	400
3	<i>Paracalanus</i> sp.	3	150
4	<i>Acartia</i> sp.	10	500
5	<i>Oithona</i> sp.	2	100
6	<i>Copepoda</i>	60	3000
7	<i>Ambasiidae</i>	6	300
			<b>4500</b>

STATION 18			
Sampling Date: 09/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Pseudodiaptomus</i> sp.	2	200
2	<i>Acartia</i> sp.	18	1800
3	<i>Oithona</i> sp.	24	2400
4	<i>Copepoda</i>	330	33000
5	<i>Microcyclops</i> sp.	6	600
6	<i>Bosmina</i> sp.	4	400
		<b>384</b>	<b>38400</b>

STATION 35			
Sampling Date: 22/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> sp.	12	2400
2	<i>Paracalanus</i> sp.	180	36000
3	<i>Centropages</i> sp.	1	200
4	<i>Acartia</i> sp.	1	200
5	<i>Oithona</i> sp.	30	6000
		<b>224</b>	<b>44800</b>

STATION 36			
Sampling Date: 22/08/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> sp.	26	5200
2	<i>Pseudodiaptomus</i> sp.	4	800
3	<i>Acartia</i> sp.	2	400
4	<i>Oithona</i> sp.	2	400
		<b>34</b>	<b>6800</b>

Survey Institute of Marine Resources and Environment, November 2006

Site	Station	T/B	Depth(m)	Turbidity	Temp(°C)	Flow velocity(m/s)		BottomSedimentType	Total solids(SS)	Suspende d(solid)(SS)	pH	Salinity(ppt)	DO6(mg/l)	NO <sub>2</sub> (mg/l)	NO <sub>3</sub> (mg/l)	NH <sub>4</sub> (mg/l)	Alkalinity(kg)	TotalP	TotalN	Chlorophylla	Zooplankton	Phytoplankton	Microalgae	Toxic algae( cell/l)	Total coliforms	E. coli	Vibro		
						Tide( Flood)	Tide(Ebb)																						
A	1	T			30.9						7.5	<1	6.2	<0.05	<0.02		0.05	<0.05							7500				
		M	0.5					Mud																					
	2	T				31.1																					1100		
		M	1.0					Mud&Clay																					
	3	T				31.7																						1500	
		M	0.7					Mud&Clay&Sand																					
B	4	T	1.0		30.6																						1500		
		M	0.9					Mud&Clay																					
	5	T		1.2		31.0																						4300	
		M	1.5					Mud&Clay																					
	6	T		0.8		31.2																						2400	
		M	0.4					Mud&Clay&Sand																					
C	7	T		1.5		30.2																					4200		
		M	2.7					Mud&Clay																					
	8	T		1.2		30.2																						400	
		M	1.3					Mud&Clay																					
	9	T		1.0		30.9																						3500	
		M	0.7					Mud&Sand																					
D	10	T		1.2		29.9																					2000		
		M	1.0					Mud&Sand																					
	11	T				29.9																						2700	
		M	1.8					Mud&Sand																					
	12	T				30.9																						12000	
		M	1.4					Mud&Clay																					
E	13	T		1.5		28.9																					1500		
		M	11.0					Mud&Sand																					
	14	T		1.2		29.6																						3400	
		M	1.0					Sand																					
	15	T		1.3		29.0																						21000	
		M	0.8					Mud																					
F	16	T		1.6		28.8																					4400		
		M	1.0					Mud&Sand																					
	17	T		1.5		29.0																						2000	
		M	1.8					Mud																					
	18	T		1.0		29.3																						700	
		M	0.9					Mud&Clay&Sand																					
G	19	T		1.9		27.5																					300		
		M	1.3					Mud&Sand																					
	20	T		2.2		28.1																						2300	
		M	2.7					Mud&Sand																					
	21	T		1.6		27.3																						900	
		M	0.3					Mud&Sand																					
22	T		1.4		28.0																						1200		
	M	1.5					Mud&Sand																						
H	23	T		2.3		28.6																					4300		
		M	3.7					Mud&Sand																					
	24	T		1.6		28.1																						15000	
		M	2.5					Mud&Clay																					
	25	T		1.5		30.3																						2700	
		M	2.0					Mud&Sand																					
I	26	T		2.0		31.9																					1500		
		M	3.3					Mud&Clay																					
	27	T		2.3		32.3																						1400	
		M	3.1					Mud&Clay																					
	28	T		1.5		30.6																						900	
		M	0.8					Mud																					
J	29	T		1.6		29.7																					700		
		M	1.0					Mud&Clay																					
	30	T		1.5		28.5																						3900	
		M	2.5					Mud																					
	31	T		2.5		33.4																						7500	
		M																											
32	T		2.0		29.7																						2600		
	M	1.8					Mud&Clay																						
K	33	T		1.5		29.4																					1500		
		M	1.3					Mud&Clay																					
	34	T		1.6																								12000	
		M	2.1		2.3	34.0																							
	35	T		1.6		29.4																						2700	
		M	1.9					Clay&Sand																					
36	T		1.5		33.2																						1100		
	M	1.3					Mud&Sand																						

Survey Institute of Marine Resources and Environment, November 2006 Phytoplankton Species

STATION 21			
Sampling Date: 28/11/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira granulata</i>	169	3380
2	<i>Flagellaria triatula</i>	30	600
3	<i>Thalassionema nitzschoides</i>	1	20
5	<i>Navicula alpebralis</i>	1	20
6	<i>Diploneis mithi</i>	2	40
7	<i>Nitzschia sigma</i>	2	40
8	<i>Nitzschia</i> sp.	1	20
9	<i>Gymnodinium</i> sp.	9	180
10	<i>Protoperidinium</i> sp.	1	20
11	<i>Alexandrium</i> sp.	4	80
12	<i>Anabaena</i> sp.	1	20
13	<i>Scenedesmus</i> sp.	8	160
14	<i>Gloetila pelagicum</i>	127	2540
		<b>362</b>	<b>7240</b>

STATION 22			
Sampling Date: 28/11/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira</i> sp.	81	1620
2	<i>Leptocylindrus lanicus</i>	6	120
3	<i>Diatoma longatum</i>	4	80
4	<i>Synedra pulchella</i>	3	60
5	<i>Nitzschia</i> sp.	2	40
6	<i>Gymnodinium</i> sp.	2	40
7	<i>Alexandrium</i> sp.	1	20
8	<i>Oscillatoria</i> sp.2	1	20
9	<i>Anabaena</i> sp.	6	120
		<b>116</b>	<b>2320</b>

STATION 23			
Sampling Date: 28/11/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira granulata</i> , <i>angustissima</i>	21	420
2	<i>Synedra</i> sp.	16	320
3	<i>Gyrosigma</i> sp.	1	20
5	<i>Nitzschia</i> sp.	12	240
6	<i>Nitzschia lorenziana</i>	3	60
7	<i>Peridinium</i> sp.	16	320
8	<i>Oscillatoria</i> sp.1	3	60
9	<i>Oscillatoria</i> sp.2	1	20
10	<i>Pediastrum duplex</i>	1	20
11	<i>Spirogyra</i> sp.	465	9300
		<b>540</b>	<b>10800</b>

STATION 24			
Sampling Date: 20/11/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira granulata</i>	27	540
2	<i>Nitzschia lorenziana</i>	8	160
3	<i>Nitzschia sigma</i>	6	120
4	<i>Oscillatoria</i> sp.2	9	180
5	<i>Spirulina</i> sp.	1	20
6	<i>Anabaena</i> sp.	420	8400
		<b>471</b>	<b>9420</b>

STATION 26			
Sampling Date: 28/11/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira granulata</i> , <i>angustissima</i>	42	840
2	<i>Melosira varians</i>	1	20
3	<i>Coscinodiscus</i> sp.	1	20
4	<i>Actinocyclus undulatus</i>	2	40
5	<i>Synedra</i> sp.	2	40
6	<i>Grammatophora marina</i>	2	40
7	<i>Trachyneis aspera</i>	1	20
8	<i>Cymbella</i> sp.	2	40
9	<i>Diploneis</i> sp.1	23	460
10	<i>Gyrosigma</i> sp.1	4	80
11	<i>Gyrosigma</i> sp.	5	100
12	<i>Pleurosigma</i> sp.	53	1060
13	<i>Pleurosigma</i> sp. fasciola	1	20

STATION 21			
Sampling Date: 02/12/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira ummuloides</i>	2	40
2	<i>Melosira</i> sp.	14	280
4	<i>Leptocylindrus lanicus</i>	10	200
5	<i>Guinardia laccida</i>	1	20
6	<i>Chaetoceros affinis</i>	2	40
7	<i>Chaetoceros bonormis</i>	6	120
8	<i>Chaetoceros lorenzianus</i>	7	140
9	<i>Palmeria gardmaniana</i>	2	40
10	<i>Thalassionema nitzschoides</i>	3	60
11	<i>Pleurosigma affine</i>	1	20
12	<i>Pleurosigma</i> sp.	16	320
13	<i>Amphiprora alata</i>	1	20
14	<i>Amphiprora</i> sp.	1	20
15	<i>Nitzschia longissima</i>	2	40
16	<i>Bacillaria paxillifera</i>	6	120
17	<i>Pseudonitzschia</i> sp.1 (to)	2	40
18	<i>Campylodiscus</i> sp.	1	20
19	<i>Prorocentrum minimum</i>	30	600
20	<i>Gymnodinium</i> sp.	57	1140
21	<i>Polykrikos schwartzii</i>	7	140
22	<i>Ceratium furca</i>	3	60
23	<i>Gonyaulax</i> sp.	2	40
24	<i>Protoperidinium lucidum</i>	1	20
25	<i>Protoperidinium</i> sp.	4	80
26	<i>Peridinium quinquecorne</i>	1	20
27	<i>Peridinium</i> sp.	20	400
28	<i>Alexandrium</i> sp.	3	60
29	<i>Alexandrium pseudogonyaulax</i>	3	60
30	<i>Diplopsalis</i> sp.	2	40
31	<i>Oblea</i> sp.	11	220
		<b>227</b>	<b>4540</b>

STATION 23			
Sampling Date: 02/12/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Coscinodiscus</i> sp.	1	20
2	<i>Chaetoceros lorenzianus</i>	6	120
3	<i>Thalassionema nitzschoides</i>	12	240
4	<i>Bacillaria paxillifera</i>	3	60
5	<i>Pseudonitzschia</i> sp.2 (nho)	2	40
6	<i>Prorocentrum minimum</i>	104	2080
7	<i>Prorocentrum marginatum</i>	1	20
8	<i>Gymnodinium</i> sp.	182	3640
9	<i>Polykrikos schwartzii</i>	4	80
10	<i>Ceratium furca</i>	1	20
11	<i>Gonyaulax</i> sp.	14	280
12	<i>Protoperidinium</i> sp.	2	40
13	<i>Peridinium</i> sp.	33	660
14	<i>Alexandrium</i> sp.	2	40
15	<i>Alexandrium pseudogonyaulax</i>	32	640
16	<i>Diplopsalis</i> sp.	2	40
17	<i>Oblea</i> sp.	4	80
18	<i>Anabaena</i> sp.	6	120
19	<i>Microcystis</i> sp.	2	40
		<b>413</b>	<b>8260</b>

STATION 24			
Sampling Date: 02/12/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Coscinodiscus onesianus</i> , <i>commutata</i>	1	20
2	<i>Pleurosigma</i> sp.	2	40
3	<i>Nitzschia longissima</i> , <i>reversa</i>	1	20
4	<i>Prorocentrum minimum</i>	34	680
5	<i>Gymnodinium</i> sp.	79	1580
6	<i>Polykrikos schwartzii</i>	8	160
7	<i>Ceratium furca</i>	1	20
8	<i>Gonyaulax</i> sp.	27	540
9	<i>Protoperidinium lucidum</i>	2	40
10	<i>Protoperidinium</i> sp.	3	60
11	<i>Scrippsiella</i> sp.	2	40
12	<i>Alexandrium pseudogonyaulax</i>	65	1300

14	<i>Nitzschia lorenziana</i>	53	1060
15	<i>Nitzschia longissima</i> <i>reversa</i>	27	540
16	<i>Nitzschia sigma</i>	12	240
17	<i>Nitzschia sigma</i> <i>intercedens</i>	12	240
18	<i>Nitzschia</i> sp.1	5	100
19	<i>Nitzschia</i> sp.	92	1840
20	<i>Surirella cymma</i>	2	40
21	<i>Surirella tenera</i> <i>nervosa</i>	28	560
22	<i>Campylopus cheneis</i>	3	60
23	<i>Podolampas elegans</i>	1	20
24	<i>Oscillatoria</i> sp1	13	260
25	<i>Anabaena</i> sp	2	40
26	<i>Scenedesmus quadricauda</i>	4	80
27	<i>Spirogyra</i> sp	1	20
28	<i>Cosmarium</i> sp	1	20
29	<i>Closterium</i> sp	1	20
		<b>396</b>	<b>7920</b>

STATION E7			
Sampling Date: 28/11/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira granulata</i>	12	240
2	<i>Thalassiosira</i> sp.	2	40
3	<i>Amphiprora</i> <i>lata</i>	2	40
4	<i>Gymnodinium</i> sp	1	20
5	<i>Oscillatoria</i> sp2	1	20
6	<i>Anabaena</i> sp	1	20
7	<i>Gloeotila pelagicum</i>	4	80
		<b>23</b>	<b>460</b>

STATION E8			
Sampling Date: 28/11/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira</i> sp.	39	780
2	<i>Nitzschia longissima</i>	1	20
3	<i>Nitzschia sigma</i>	6	120
4	<i>Pseudonitzschia</i> sp.	1	20
5	<i>Alexandrium</i> sp	12	240
6	<i>Oscillatoria</i> sp1	1	20
7	<i>Anabaena</i> sp	25	500
		<b>85</b>	<b>1700</b>

STATION E9			
Sampling Date: 28/11/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira granulata</i> <i>angustissima</i>	44	880
2	<i>Chaetoceros</i> <i>abnormis</i>	4	80
3	<i>Trachyneis</i> <i>aspera</i>	1	20
4	<i>Cymbella</i> sp.	1	20
5	<i>Nitzschia lorenziana</i>	9	180
6	<i>Nitzschia longissima</i> <i>reversa</i>	63	1260
7	<i>Nitzschia sigma</i> <i>intercedens</i>	1	20
8	<i>Nitzschia</i> sp.	12	240
9	<i>Peridinium</i> sp.	33	660
10	<i>Oscillatoria</i> sp1	14	280
11	<i>Oscillatoria</i> sp2	9	180
12	<i>Anabaena</i> sp	1	20
13	<i>Scenedesmus quadricauda</i>	8	160
14	<i>Euglena</i> sp1.	2	40
15	<i>Euglenacus</i> <i>acus</i>	2	40
16	<i>Phacus</i> sp1.	1	20
		<b>205</b>	<b>4100</b>

STATION D10			
Sampling Date: 29/11/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassiosira</i> <i>lineata</i>	1	20
2	<i>Rhizosolenia</i> <i>setigera</i>	1	20
3	<i>Chaetoceros</i> <i>abnormis</i>	6	120
4	<i>Pleurosigma</i> <i>angulatum</i>	3	60
5	<i>Pleurosigma</i> sp	1	20
6	<i>Nitzschia lorenziana</i>	14	280
7	<i>Nitzschia longissima</i>	1	20
8	<i>Nitzschia sigma</i>	1	20
9	<i>Nitzschia sigma</i>	12	240
10	<i>Nitzschia</i> sp.	4	80
11	<i>Nitzschia</i> sp.	2	40
12	<i>Prorocentrum</i> <i>minimum</i>	1	20
13	<i>Gymnodinium</i> sp	1	20

13	<i>Diplopsalis</i> sp.	1	20
14	<i>Oblea</i> sp	3	60
15	<i>Pyrophacus</i> sp	3	60
16	<i>Oscillatoria</i> sp2	1	20
		<b>233</b>	<b>4660</b>

STATION E25			
Sampling Date: 01/12/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Pleurosigma</i> <i>affine</i>	1	20
2	<i>Pleurosigma</i> sp	1	20
3	<i>Nitzschia sigma</i>	1	20
4	<i>Nitzschia</i> sp.	2	40
5	<i>Prorocentrum</i> <i>minimum</i>	16	320
6	<i>Gymnodinium</i> sp	20	400
7	<i>Polykrikos</i> <i>schwartzii</i>	1	20
8	<i>Alexandrium</i> sp	40	800
9	<i>Alexandrium</i> <i>pseudogonyalax</i>	72	1440
10	<i>Oblea</i> sp	6	120
11	<i>Pyrophacus</i> sp	1	20
12	<i>Oscillatoria</i> sp2	32	640
13	<i>Staurastrum</i> sp.	1	20
		<b>156</b>	<b>3120</b>

STATION E29			
Sampling Date: 01/12/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Cyclotella</i> <i>coma</i>	1	20
2	<i>Thalassiosira</i> <i>lineata</i>	1	20
3	<i>Guinardia</i> <i>flaccida</i>	2	40
4	<i>Guinardia</i> <i>triatra</i>	1	20
5	<i>Chaetoceros</i> <i>urvisetus</i>	14	280
6	<i>Chaetoceros</i> <i>lorenzianus</i>	2	40
7	<i>Chaetoceros</i> sp.	5	100
8	<i>Palmeria</i> <i>ardmaniana</i>	1	20
9	<i>Navicula</i> <i>membranacea</i>	1	20
10	<i>Prorocentrum</i> <i>minimum</i>	20	400
11	<i>Polykrikos</i> <i>schwartzii</i>	4	80
12	<i>Gonyalax</i> sp	1	20
13	<i>Protoperdinium</i> sp.	18	360
14	<i>Peridinium</i> <i>quinquecone</i>	1	20
15	<i>Peridinium</i> sp.	17	340
16	<i>Alexandrium</i> <i>pseudogonyalax</i>	21	420
17	<i>Oblea</i> sp	1	20
18	<i>Pyrophacus</i> sp	4	80
19	<i>Hermesium</i> sp.	1	20
20	<i>Anabaena</i> sp	160	3200
		<b>278</b>	<b>5560</b>

STATION B30			
Sampling Date: 01/12/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira</i> sp.	4	80
2	<i>Coscinodiscus</i> <i>oculus-iridis</i>	2	40
3	<i>Asteromphalus</i> <i>leveanus</i>	1	20
4	<i>Lauderia</i> <i>arealis</i>	4	80
5	<i>Leptocylindrus</i> <i>lanicus</i>	93	1860
6	<i>Guinardia</i> <i>flaccida</i>	14	280
7	<i>Guinardia</i> <i>triatra</i>	38	760
8	<i>Bacteriostroma</i> <i>arians</i>	15	300
9	<i>Bacteriostroma</i> <i>longatum</i>	2	40
10	<i>Rhizosolenia</i> <i>cylindrus</i>	14	280
11	<i>Rhizosolenia</i> <i>robusta</i>	2	40
12	<i>Rhizosolenia</i> <i>bergonii</i>	1	20
13	<i>Rhizosolenia</i> sp.	16	320
14	<i>Chaetoceros</i> <i>affinis</i>	12	240
15	<i>Chaetoceros</i> <i>urvisetus</i>	78	1560
16	<i>Chaetoceros</i> <i>dentilatus</i>	2	40
17	<i>Chaetoceros</i> <i>delistans</i>	7	140
18	<i>Chaetoceros</i> <i>ibienii</i>	2	40
19	<i>Chaetoceros</i> <i>lorenzianus</i>	79	1580
20	<i>Biddulphia</i> <i>regia</i>	4	80
21	<i>Odontella</i> <i>nobilensis</i>	6	120
22	<i>Odontella</i> sp.	1	20
23	<i>Hemiaulus</i> <i>auackii</i>	1	20
24	<i>Ditylum</i> sp.	3	60
25	<i>Climacocum</i> <i>biconcavum</i>	8	160
26	<i>Thalassionema</i> <i>nitzschoides</i>	56	1120
27	<i>Navicula</i> <i>membranacea</i>	85	1700



14	<i>Peridinium</i> sp.	3	60
15	<i>Scrippsiella</i> sp.	2	40
16	<i>Oblea</i> sp.	1	20
17	<i>Oscillatoria</i> sp.2	3	60
		57	1140

STATION 11			
Sampling date: 29/11/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassiosira</i> sp.	5	100
2	<i>Nitzschia</i> longissima	3	60
3	<i>Nitzschia</i> sigma	2	40
4	<i>Gymnodinium</i> sanguineum	1	20
5	<i>Gonyaulax</i> polygramma	4	80
6	<i>Protoperidinium</i> sp.	6	120
7	<i>Peridinium</i> quinquecorne	1	20
8	<i>Alexandrium</i> sp.	4	80
9	<i>Alexandrium</i> sp.	3	60
10	<i>Anabaena</i> sp.	6	120
		35	700

STATION 12			
Sampling date: 29/11/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassiosira</i> neata	1	20
2	<i>Nitzschia</i> coreniana	1	20
3	<i>Nitzschia</i> longissima	1	20
4	<i>Nitzschia</i> longissima reversa	3	60
5	<i>Nitzschia</i> sigma	7	140
6	<i>Nitzschia</i> sp.	2	40
7	<i>Prorocentrum</i> minimum	5	100
8	<i>Peridinium</i> sp.	5	100
9	<i>Alexandrium</i> sp.	23	460
10	<i>Oblea</i> sp.	5	100
11	<i>Anabaena</i> sp.	4	80
		57	1140

STATION 13			
Sampling date: 29/11/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Asteromphalus</i> leveanus	1	20
2	<i>Thalassiosira</i> sp.	4	80
3	<i>Diatoma</i> longatum	2	40
4	<i>Pseudonitzschia</i> sp.	3	60
5	<i>Suriella</i> tenera nervosa	1	20
6	<i>Polykrikos</i> schwartzii	1	20
7	<i>Ceratium</i> furca	1	20
8	<i>Alexandrium</i> sp.	1	20
9	<i>Anabaena</i> sp.	4	80
10	<i>Pediastrum</i> sp.	1	20
11	<i>Gloeotila</i> pelagicum	9	180
		28	560

STATION 15			
Sampling date: 29/11/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassiosira</i> sp.	2	40
2	<i>Diatoma</i> longatum	2	40
3	<i>Thalassionema</i> itzschoides	7	140
4	<i>Thalassiothrix</i> frauenfeldii	1	20
5	<i>Diploneis</i> smithii	1	20
6	<i>Nitzschia</i> longissima	3	60
7	<i>Spirulina</i> sp.	1	20
8	<i>Pediastrum</i> sp.	1	20
9	<i>Scenedesmus</i> sp.	4	80
10	<i>Gloeotila</i> pelagicum	9	180
		31	620

STATION 17			
Sampling date: 29/11/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Rhizosolenia</i> cylindrus	3	60
2	<i>Rhizosolenia</i> setigera	1	20
3	<i>Chaetoceros</i> sp.	2	40
4	<i>Diploneis</i> sp. 1	1	20
5	<i>Pleurosigma</i> angulatum	1	20
6	<i>Pleurosigma</i> sp.	1	20
7	<i>Nitzschia</i> coreniana	1	20
8	<i>Nitzschia</i> longissima	2	40

28	<i>Pleurosigma</i> sp.	2	200
29	<i>Nitzschia</i> sigma	4	400
30	<i>Bacillaria</i> paxillifera	5	500
31	<i>Pseudonitzschia</i> sp. 1 (to)	11	1100
32	<i>Prorocentrum</i> minimum	2	200
33	<i>Prorocentrum</i> sp. 1 (thin coat)	1	100
34	<i>Protoperidinium</i> depressum	1	100
35	<i>Diplopsalis</i> sp.	1	100
		577	57700

STATION 32			
Sampling date: 30/11/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Nitzschia</i> sigma	1	20
2	<i>Prorocentrum</i> minimum	27	540
3	<i>Gymnodinium</i> sp.	1	20
4	<i>Gonyaulax</i> sp.	2	40
5	<i>Peridinium</i> sp.	1	20
6	<i>Alexandrium</i> sp.	1	20
7	<i>Alexandrium</i> pseudogonyaulax	163	3260
8	<i>Oblea</i> sp.	4	80
9	<i>Oscillatoria</i> sp. 1	2	40
10	<i>Anabaena</i> sp.	1	20
		244	4880

STATION 35			
Sampling date: 30/11/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Cyclotella</i> omta	1	20
2	<i>Coscinodiscus</i> oculus-iridis	12	240
3	<i>Actinopterychus</i> indulatus	1	20
4	<i>Thalassiothrix</i> longatum	1	20
5	<i>Thalassiosira</i> sp.	8	160
6	<i>Guinardia</i> laccida	3	60
7	<i>Guinardia</i> triata	2	40
8	<i>Chaetoceros</i> curvisetus	12	240
9	<i>Thalassionema</i> itzschoides	17	340
10	<i>Synedra</i> pulchella	1	20
11	<i>Grammatophora</i> marina	3	60
12	<i>Pleurosigma</i> aviculaceum	3	60
13	<i>Pseudonitzschia</i> sp.	2	40
14	<i>Ceratium</i> furca	1	20
15	<i>Gonyaulax</i> polygramma	1	20
16	<i>Alexandrium</i> sp.	524	10480
17	<i>Diplopsalis</i> sp.	1	20
18	<i>Oscillatoria</i> sp. 1	2	40
		584	11680

9	<i>Nitzschia longissima</i> var. <i>reversa</i>	1	20
10	<i>Nitzschia</i> sp.	1	20
11	<i>Pseudonitzschia</i> sp. 1 (to)	5	100
12	<i>Prorocentrum</i> <i>unicans</i>	2	40
13	<i>Prorocentrum</i> <i>minimum</i>	28	560
14	<i>Gymnodinium</i> sp	104	2080
15	<i>Polykrikos</i> <i>Schwartzii</i>	5	100
16	<i>Gonyaulax</i> sp	12	240
17	<i>Protoperidinium</i> sp.	19	380
18	<i>Peridinium</i> <i>quinquecorne</i>	2	40
19	<i>Peridinium</i> sp.	64	1280
20	<i>Alexandrium</i> <i>pseudogonyaulax</i>	2	40
21	<i>Obleata</i> sp	34	680
22	<i>Pyrophacus</i> sp	7	140
		<b>298</b>	<b>5960</b>

STATION F18			
Sampling Date: 29/11/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Melosira</i> <i>granulata</i> var. <i>angustissima</i>	26	520
2	<i>Prorocentrum</i> <i>minimum</i>	4	80
3	<i>Gonyaulax</i> sp	1	20
4	<i>Peridinium</i> sp.	10	200
5	<i>Oscillatoria</i> sp1	1	20
6	<i>Anabaena</i> sp	20	400
		<b>62</b>	<b>1240</b>

STATION G19			
Sampling Date: 02/12/2006			
No	SPECIES	QUANTITY	CELLS/L
1	<i>Thalassiosira</i> sp.	6	120
2	<i>Chaetoceros</i> sp.	25	500
3	<i>Grammatophora</i> <i>marina</i>	9	180
4	<i>Nitzschia</i> <i>longissima</i>	6	120
5	<i>Pseudonitzschia</i> sp.	1	20
6	<i>Gymnodinium</i> sp	17	340
7	<i>Polykrikos</i> <i>Schwartzii</i>	7	140
8	<i>Ceratium</i> <i>breve</i>	1	20
9	<i>Ceratium</i> <i>furca</i>	3	60
10	<i>Gonyaulax</i> <i>polygamma</i>	29	580
11	<i>Protoperidinium</i> sp.	20	400
12	<i>Peridinium</i> <i>quinquecorne</i>	1	20
13	<i>Alexandrium</i> sp.	23	460
14	<i>Anabaena</i> sp	2	40
		<b>150</b>	<b>3000</b>

**Survey of Marine Resources and Environment, November 2006 Zooplankton Species**

STATION 1			
Sampling date: 8/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	14	140
2	<i>Acartia lausi</i>	1	10
3	<i>Acartia</i>	1	10
		<b>16</b>	<b>160</b>

STATION 3			
Sampling date: 8/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	6	300
2	<i>Acartia lausi</i>	2	100
3	<i>Oithona</i> sp.	19	950
4	<i>Acartia</i>	1	50
5	<i>Copepoda</i>	24	1200
		<b>52</b>	<b>2600</b>

STATION 4			
Sampling date: 8/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	13	650
2	<i>Oncaea venusta</i>	2	100
3	<i>Corycaeus</i> sp.	1	50
4	<i>Acartia sinensis</i>	4	200
		<b>20</b>	<b>1000</b>

STATION 5			
Sampling date: 8/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	12	600
2	<i>Acartia lausi</i>	8	400
3	<i>Oithona</i> sp.	7	350
4	<i>Copepoda</i>	12	600
		<b>39</b>	<b>1950</b>

STATION 6			
Sampling date: 8/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Evadne cordmani</i>	1	250
2	<i>Paracalanus parvus</i>	30	7500
3	<i>Oithona</i> sp.	32	8000
		<b>63</b>	<b>15750</b>

STATION 7			
Sampling date: 8/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	4	400
2	<i>Acartia lausi</i>	14	1400
3	<i>Oithona</i> sp.	3	300
4	<i>Clytemnestra</i> sp.	1	100
		<b>22</b>	<b>2200</b>

STATION 8			
Sampling date: 8/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Evadne ergestina claus</i>	9	450
2	<i>Penilia schmackeri</i>	1	50
3	<i>Paracalanus parvus</i>	1	50
4	<i>Acartia lausi</i>	1	50
5	<i>Oithona</i> sp.	12	600
6	<i>Oncaea venusta</i>	7	350
7	<i>Oikopleura dioica</i>	1	50
		<b>32</b>	<b>1600</b>

STATION 9			
Sampling date: 8/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Acartia lausi</i>	11	2750
2	<i>Oithona</i> sp.	120	30000
3	<i>Copepoda</i>	18	4500
		<b>149</b>	<b>37250</b>

STATION 20			
Sampling date: 10/2/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Evadne ergestina claus</i>	4	80
2	<i>Conchoecia</i> sp.	1	20
3	<i>Paracalanus</i> sp.	3	60
4	<i>Pseudodiaptomus</i> sp.	1	20
5	<i>Acartia lausi</i>	9	180
6	<i>Oithona</i> sp.	3	60
7	<i>Euterpina acutifront</i>	1	20
8	<i>Copepoda</i>	80	1600
		<b>102</b>	<b>2040</b>

STATION 22			
Sampling date: 10/2/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Sagitta helicata</i>	1	10
2	<i>Penilia schmackeri</i>	1	10
3	<i>Paracalanus parvus</i>	18	180
4	<i>Oithona</i> sp.	13	130
5	<i>Corycaeus thalhi</i>	1	10
6	<i>Copepoda</i>	6	60
		<b>40</b>	<b>400</b>

STATION 23			
Sampling date: 10/2/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	5	50
2	<i>Oithona</i> sp.	9	90
3	<i>Copepoda</i>	6	60
		<b>20</b>	<b>200</b>

STATION 24			
Sampling date: 10/2/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	5	50
2	<i>Oithona similis</i>	6	60
3	<i>Copepoda</i>	48	480
		<b>59</b>	<b>590</b>

STATION 25			
Sampling date: 10/1/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	9	90
2	<i>Pseudodiaptomus</i> sp.	6	60
3	<i>Acartia lausi</i>	8	80
4	<i>Euterpina acutifront</i>	1	10
5	<i>Copepoda</i>	48	480
		<b>72</b>	<b>720</b>

STATION 26			
Sampling date: 10/1/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	15	150
2	<i>Acartia lausi</i>	14	140
3	<i>Oithona</i> sp.	1	10
4	<i>Oncaea venusta</i>	1	10
5	<i>Acartia</i>	1	10
6	<i>Copepoda</i>	60	600
		<b>92</b>	<b>920</b>

STATION 27			
Sampling date: 10/1/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	6	60
2	<i>Acartia lausi</i>	1	10
3	<i>Copepoda</i>	57	570
		<b>64</b>	<b>640</b>

STATION 28			
Sampling date: 10/1/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	5	50

STATION#10			
Sampling date: 09/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Evadnebergestina</i> Claus	1	10
2	<i>Penilia</i> schmackeri	1	10
3	<i>Conchocia</i> mbricata	1	10
4	<i>Oithona</i> sp.	5	50
5	<i>Oncaea</i> venusta	1	10
		9	90

STATION#11			
Sampling date: 09/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> parvus	6	60
2	<i>Pseudodiaptomus</i> sp.	2	20
3	A.T.T.K.m	3	30
		11	110

STATION#12			
Sampling date: 09/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> parvus	4	40
2	<i>Acartia</i> lausi	28	280
3	<i>Oithona</i> brevicornis	4	40
		36	360

STATION#13			
Sampling date: 09/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Evadnebergestina</i> Claus	24	240
2	<i>Penilia</i> schmackeri	2	20
3	<i>Paracalanus</i> parvus	9	90
4	<i>Centropages</i> furcatus	1	10
5	<i>Oithona</i> sp.	2	20
6	<i>Oncaea</i> venusta	8	80
7	<i>Microsetella</i> norvegica	2	20
8	<i>Oikopleura</i> dioica	4	40
9	A.T.T.K.m	3	30
10	<i>Cypridina</i> noctiluca	5	50
		60	600

STATION#14			
Sampling date: 09/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> parvus	14	140
2	<i>Acartia</i> sp.	2	20
3	<i>Oithona</i> sp.	5	50
4	<i>Euterpina</i> acutifront	1	10
		22	220

STATION#15			
Sampling date: 09/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> parvus	1	10
2	<i>Microsetella</i> norvegica	1	10
3	<i>Copepoda</i>	6	60
		8	80

STATION#16			
Sampling date: 11/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> parvus	9	90
2	<i>Oithona</i> sp.	2	20
3	<i>Acartia</i> sinensis	1	10
		12	120

STATION#17			
Sampling date: 09/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> parvus	2	20
2	<i>Oithona</i> sp.	1	10
3	<i>Copepoda</i>	12	120
		15	150

3	<i>Copepoda</i>	6	60
		13	130

STATION#29			
Sampling date: 10/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> sp.	28	280
2	<i>Acartia</i> lausi	4	40
3	<i>Oithona</i> sp.	2	20
4	<i>Microsetella</i> norvegica	3	30
		37	370

STATION#30			
Sampling date: 10/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Eucalanus</i> subcrassus Giesb.	2	200
2	<i>Paracalanus</i> parvus	36	3600
3	<i>Acartia</i> lausi	6	600
4	<i>Oithona</i> sp.	12	1200
5	<i>Corycaeus</i> thalhi	1	100
6	<i>Corycaeus</i> sp.	9	900
		66	6600

STATION#31			
Sampling date: 09/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> sp.	2	20
2	<i>Oithona</i> sp.	1	10
3	<i>Copepoda</i>	6	60
		9	90

STATION#32			
Sampling date: 09/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Penilia</i> schmackeri	1	10
2	<i>Paracalanus</i> parvus	7	70
3	<i>Oithona</i> sp.	2	20
4	<i>Oikopleura</i> rufescens	2	20
5	<i>Copepoda</i>	160	1600
		172	1720

STATION#33			
Sampling date: 09/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Sagitta</i> helicata	1	10
2	<i>Evadne</i> nordmani	2	20
3	<i>Paracalanus</i> parvus	23	230
4	<i>Acartia</i> pacifica	7	70
5	<i>Acartia</i> lausi	10	100
6	<i>Oithona</i> sp.	6	60
		49	490

STATION#37			
Sampling date: 09/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> parvus	9	450
2	<i>Acartia</i> lausi	1	50
3	<i>Oithona</i> sp.	3	150
4	<i>Oncaea</i> venusta	3	150
		16	800

STATION#34			
Sampling date: 09/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus</i> parvus	9	90
2	<i>Acartia</i> lausi	7	70
3	<i>Oithona</i> sp.	1	10
4	<i>Copepoda</i>	11	110
		28	280

STATION#35			
Sampling date: 09/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Harpacticoda</i>	1	10

STATION 18			
Sampling Date: 29/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	2	20
2	<i>Copepoda</i>	6	60
3	<i>AT&amp;T&lt;m</i>	3	30
		11	110

STATION 19			
Sampling Date: 02/12/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	48	480
2	<i>Acartia lausi</i>	70	700
3	<i>Oithona sp.</i>	18	180
4	<i>Microsetella norvegica</i>	6	60
5	<i>Copepoda</i>	86	860
		228	2280

2	<i>Paracalanus parvus</i>	6	60
3	<i>Acartia lausi</i>	1	10
4	<i>Copepoda</i>	35	350
		43	430

STATION 36			
Sampling Date: 30/11/2006			
No	SPECIES	QUANTITY	INDS/M3
1	<i>Paracalanus parvus</i>	14	140
2	<i>Acartia lausi</i>	1	10
3	<i>AT&amp;T&lt;m</i>	1	10
		16	160

Survey Institute of Marine Resources and Environment, May 2007

Site	Station	T/B	Depth(m)	Turbidity	Temp(°C)	FlowVelocity(m/s) Tide(Flood) Tide(Ebb)	BottomSedimentType	TotalSolid(TSS)	Suspende(dSolid)(SS)	pH	Salinity(ppt)	DO(mg/l)	NO <sub>2</sub> (mg/l)	NO <sub>3</sub> (mg/l)	NH <sub>4</sub> (mg/l)	Alkalinity(°K)	TotalP	TotalN	Chlorophylla(a)(mg/m <sup>3</sup> )	Zoobenthos(biomass)(unit/m <sup>2</sup> )	Phytoplankton(tonn)(cell/l)	Microalgae	ToxicAlgae(cell/l)	TotalColiforms	E.coli	Vibrio		
A	1	+			27.3					7.04	<1	6.10	<0.05	0.02		0.02	0.28	0.7										
		M	0.5				Mud							0.05	<0.02		0.02	0.29	1.6									
		+				28.5					6.09	<1	6.30															
B	2	+			30.2					7.95	<1	6.65	<0.05	0.02		0.05	<0.05	8.0										
		M	0.7				Mud&Clay&Sand						<0.05	<0.02		0.05	0.65	3.2										
		+				28.5					7.22	<1	6.20				0.07	<0.02	0.05	0.39	0.6							
C	4	+			28.5					6.60	<1	6.15				<0.05	<0.02	0.05	0.21	1.7								
		M	0.9				Mud&Clay																					
		+				30.0					8.42	<1	6.70				0.11	<0.02	0.02	0.37	3.7							
D	5	+			27.8					7.82	<1	6.65				<0.05	<0.02	0.02	0.21	1.1								
		M	2.7				Mud&Clay																					
		+				28.5					7.55	<1	6.35	<0.05	<0.02		0.05	0.49	4.2									
E	6	+			28.8					7.61	<1	6.15				<0.05	<0.02	0.05	0.52	3.0								
		M	1.3				Mud&Sand																					
		+				27.4					7.93	<1	6.70	0.06	0.02		0.05	0.52	3.0									
F	7	+			27.6					8.05	1.5	6.70				0.12	<0.02	0.05	0.65	2.3								
		M	1.8				Mud&Sand																					
		+				28.5					7.86	<1	6.60	0.10	<0.02		0.05	0.31	3.4									
G	8	+			27.7					8.23	2.3	7.25				0.06	0.02	0.05	0.39	1.7								
		M	1.0				Mud&Sand																					
		+				28.7					7.60	<1	6.80	0.12	0.05		0.02	0.60	1.1									
H	9	+			29.0					7.49	<1	7.10				0.09	0.04	0.02	0.35	1.2								
		M	0.8				Mud																					
		+				29.5					8.40	2.0	7.10	0.06	0.02		0.02	0.78	3.8									
I	10	+			29.0					8.31	1.8	7.30	0.06	0.02		0.02	1.05	2.7										
		M	1.8				Mud																					
		+				30.2					8.55	1.6	7.25	0.06	<0.02		0.05	1.28	3.6									
J	11	+			28.5					8.37	2.0	6.55	<0.05	0.02		0.02	1.53	2.5										
		M	1.3				Mud&Sand																					
		+				28.5					8.37	2.0	6.70				0.06	0.04	0.02	0.62								
K	12	+			28.9					8.39	2.0	6.60	<0.05	0.02		0.02	0.93											
		M	2.7				Mud&Sand																					
		+				28.9					8.39	2.0	6.60	<0.05	<0.02		0.02	0.91	2.8									
L	13	+			28.8					8.27	2.2	7.35	<0.05	<0.02		0.02	1.08											
		M	1.5				Mud&Sand																					
		+				28.8					8.34	2.2	7.55	<0.05	<0.02		0.02	1.08										
M	14	+			28.9					8.34	2.2	7.50	<0.05	<0.02		0.02	0.67											
		M	2.5				Mud&Clay																					
		+				29.0					8.24	2.0	7.40	<0.05	<0.02		0.02	0.78										
N	15	+			29.3					8.16	2.0	7.80	<0.05	<0.02		0.02	1.30	4.0										
		M	2.0				Mud&Sand																					
		+				29.1					8.26	2.0	7.55	<0.05	<0.02		0.02	0.96										
O	16	+			29.6					8.40	1.5	7.30	<0.05	0.02		0.02	0.69											
		M	0.8				Mud																					
		+				29.9					8.81	1.3	7.10	<0.05	<0.02		0.02	0.67	0.6									
P	17	+			28.3					8.25	4.0	7.50	<0.05	<0.02		0.02	0.67	0.6										
		M	1.0				Mud&Clay																					
		+				30.5					8.65	1.3	7.35	<0.05	<0.02		0.02	0.71	1.8									
Q	18	+			30.3					8.49	1.6	7.55	<0.05	<0.02		0.02	0.70											
		M	1.8				Mud&Clay																					
		+				29.3					8.30	1.5	7.25	<0.05	<0.02		<0.01	0.65	1.1									
R	19	+			30.3					8.83	1.3	7.20	<0.05	<0.02		0.02	0.68	0.8										
		M	1.6				Mud&Clay																					
		+				29.7					8.88	1.5	7.10	<0.05	<0.02		<0.01	1.45	1.1									
S	20	+			28.9					8.56	1.5	7.25	<0.05	<0.02		0.02	0.98	1.4										
		M	2.1				Mud&Sand																					
		+				29.5					8.70	1.3	7.35	<0.05	0.02		0.02	0.49	2.4									

Report by Chu Chi Thiet (2009)

Site	Station	T/B	Depth(m)	Turbidity	Temp(°C)	FlowVelocity(m/s) Tide(Flood) Tide(Ebb)	BottomSedimentType	TotalSolid(TSS)	Suspende(dSolid)(SS)	pH	Salinity(ppt)	DO(mg/l)	NO <sub>2</sub> (mg/l)	NO <sub>3</sub> (mg/l)	NH <sub>4</sub> (mg/l)	Alkalinity(°K)	TotalP	TotalN	Chlorophylla(a)(mg/m <sup>3</sup> )	Zoobenthos(biomass)(unit/m <sup>2</sup> )	Phytoplankton(tonn)(cell/l)	Microalgae	ToxicAlgae(cell/l)	TotalColiforms	E.coli	Vibrio	
1	1	+		0.65	32.0					7.6	5.0	5.00	0.00	0.13	0.00		0.00	0.28	2.4			80.500					
		M	1.1				Sand&Mud			8.4	16.0	5.00	0.00	0.07	0.00		0.00	0.00	1.2			83.333					
		+				32.0					8.3	17.0	5.33	0.00	0.00	0.00		0.00	0.00	0.0			57.501				
2	2	+		0.70	32.0					8.4	17.0	5.00	0.00	0.00	0.03		0.00	0.00	0.4			86.938					
		M	1.1				Sand&Mud			8.4	17.0	5.00	0.00	0.00	0.00		0.00	0.00	0.0			25.670					
		+				33.0					8.4	11.0	5.80	0.00	0.00	0.00		0.00	0.00	0.0			35.936				
3	3	+		0.60	33.0					8.5	11.0	5.70	0.00	0.00	0.00		0.00	0.00	1.5			13.900					
		M	1.2				Sand&Mud			8.5	11.0	5.70	0.00	0.00	0.00		0.00	0.00	0.6			8.833					
		+																									

**Survey Tran Dinh Minh, November 13th, 2011**

Site	Station	T/B	Depth(m)	Turbidity	Temp(°C)	Flow Velocity(m/s)	Tide(Flood)	Tide(Ebb)	Bottom Sediment Type	Total Solid (%)	Suspended Solids (SS)	pH	Salinity (ppt)	DO(mg/l)	NO <sub>2</sub> (mg/l)	NO <sub>3</sub> (mg/l)	NH <sub>4</sub> (mg/l)	Alkalinity (kh)	Total P	Total N	Chlorophyll a (mg/m <sup>3</sup> )	Zoobenthos Biomass (unit/m <sup>2</sup> )	Biomass (g/m <sup>2</sup> )	Phytoplankton (cells/l)	Microalgae (cell/l)	Total Coliforms	E. coli	Vibrio
A	T			0,65	28,5				Sand/Mud			8,20	27,0	6,00	<0,02	<0,05	<0,02	115			3,09					0	0	
	M	1,1																										
B	T			0,70	28,0				Sand/Mud			8,20	3,5	5,00	<0,02	<0,05	<0,02	80			2,89					0	0	
	M	1,1																										
C	T			0,60	28,0				Sand/Mud			7,50	1,0	6,00	<0,02	<0,05	<0,02	20			4,95					4000	43000	
	M	1,2																										
D	T			0,56	28,0				Mud/Sand			7,00	1,0	5,50	<0,02	<0,05	<0,02	20			7,01					4000	23000	
	M	0,9																										
E	T			0,46	28,0				Sand/Mud			7,90	2,0	5,50	<0,02	<0,05	<0,02	40			2,10					43000	43000	
	M	0,7																										

**Survey Tran Dinh Minh, November 13th, 2011 - Phytoplankton Species**

Station	Sampling Date	Analysis Date	No	SPECIES
A	3/11/2011	21/11/2011	<b>CYANOPHYTA</b>	
			1	<i>Aphanocapsa@elicatissima</i>
			2	<i>Aphanocapsa@hololica</i>
			3	<i>Microcystis@eruginosa</i>
			4	<i>Oscillatoria@mosa</i>
5	<i>Oscillatoria@tenus</i>			
<b>CHRYSOPHYTA</b>		6	<i>Dinobryon@setularia</i>	
<b>BACILLARIOPHYTA</b>		7	<i>Asteromphalus@leveanus</i>	
8	<i>Coconeis@cutellum</i>			
9	<i>Coscinodiscus@bulliens</i>			
10	<i>Coscinodiscus@onesianus</i>			
11	<i>Coscinodiscus@radiatus</i>			
12	<i>Cyclotella@triatra</i>			
13	<i>Chaetoceros@orenzianus</i>			
14	<i>Chaetoceros@parvodus</i>			
15	<i>Diploneis@fusca</i>			
16	<i>Gyrosigma@tenuatum</i>			
17	<i>Lauderia@nulata</i>			
18	<i>Lyrella@lyroides</i>			
19	<i>Melosira@granulata</i>			
20	<i>Melosira@granulata@var.@angussima</i>			
21	<i>Nitzschia@losterium</i>			
22	<i>Nitzschia@sigma</i>			
23	<i>Odontella@egida</i>			
24	<i>Odontella@inensis</i>			
25	<i>Petronella@marina</i>			
26	<i>Pleurosigma@bestuarii</i>			
27	<i>Pleurosigma@affine</i>			
28	<i>Rhizosolenia@sp.</i>			
29	<i>Surirella@gemma</i>			
30	<i>Surirella@minuta</i>			
31	<i>Synedra@lina</i>			
32	<i>Thalassionema@nitzschoides</i>			
33	<i>Tricarotium@fusus</i>			
<b>CHLOROPHYTA</b>		34	<i>Actinastrum@bantzchii@var.@bantzchii</i>	
35	<i>Arthrodesmus@convergens</i>			
36	<i>Coelastrum@amblicum</i>			
37	<i>Coelastrum@phaerocum</i>			
38	<i>Cosmarium@contractum</i>			
39	<i>Cosmarium@moniliforme</i>			
40	<i>Dictyosphaerium@pulchellum</i>			
41	<i>Eunotia@legans</i>			
42	<i>Micrasterium@borhemiense</i>			
43	<i>Micrasterium@quadrisetum@var.@quadrisetum</i>			
44	<i>Pandorina@horum</i>			
45	<i>Pediastrum@duplex@var.@duplex</i>			
46	<i>Pediastrum@duplex@var.@reticulatum</i>			
47	<i>Pediastrum@oryanum@var.@longicorne</i>			
48	<i>Scenedesmus@bliquus@var.@alternans</i>			
49	<i>Scenedesmus@cuminatus@var.@cuminatus</i>			
50	<i>Scenedesmus@cuminatus@var.@biseratus</i>			
51	<i>Scenedesmus@quadricauda@var.@quadricauda</i>			
52	<i>Schroederia@sp.</i>			
53	<i>Staurastrum@racile</i>			
54	<i>Staurastrum@odulosum</i>			
55	<i>Staurastrum@benarium</i>			
56	<i>Staurastrum@sp.</i>			
57	<i>Staurastrum@triatratum@var.@triatratum</i>			
58	<i>Staurastrum@wildemanii</i>			
59	<i>Tetraedron@racile</i>			
60	<i>Tetraedron@racile</i>			

Station	Sampling Date	Analysis Date	No	SPECIES
B	3/11/2011	24/11/2012	<b>CYANOPHYTA</b>	
			1	<i>Anabaena@f.rossa</i>
			2	<i>Aphanocapsa@elicatissima</i>
			3	<i>Cyanodictyon@perfectum</i>
			4	<i>Microcystis@eruginosa</i>
5	<i>Pseudonitzschia@moniliformis</i>			
<b>CHRYSOPHYTA</b>		6	<i>Dinobryon@bavaricum</i>	
<b>BACILLARIOPHYTA</b>		7	<i>Asteromphalus@leveanus</i>	
8	<i>Biddulphia@sp.</i>			
9	<i>Coconeis@cutellum</i>			
10	<i>Coscinodiscus@radiatus</i>			
11	<i>Cyclotella@triatra</i>			
12	<i>Diploneis@bombus</i>			
13	<i>Ditylum@rightwellii</i>			
14	<i>Guinardia@triatra</i>			
15	<i>Gyrosigma@fasciola</i>			
16	<i>Gyrosigma@tenuatum</i>			
17	<i>Lyrella@poides</i>			
18	<i>Melosira@granulata</i>			
19	<i>Melosira@granulata@var.@angussima</i>			
20	<i>Nitzschia@sigma</i>			
21	<i>Odontella@inensis</i>			
22	<i>Petronella@marina</i>			
23	<i>Pleurosigma@bestuarii</i>			
24	<i>Surirella@obustata@var.@splendida</i>			
25	<i>Thalassionema@nitzschoides</i>			
<b>CHLOROPHYTA</b>		26	<i>Actinastrum@bantzchii@var.@bantzchii</i>	
27	<i>Closterium@juncidum</i>			
28	<i>Cosmarium@contractum</i>			
29	<i>Dictyosphaerium@pulchellum</i>			
30	<i>Eunotia@legans</i>			
31	<i>Micrasterium@borhemiense</i>			
32	<i>Pandorina@horum</i>			
33	<i>Pediastrum@oryanum@var.@oryanum</i>			
34	<i>Scenedesmus@cuminatus@var.@biseratus</i>			
35	<i>Scenedesmus@benticulatus@var.@biseratus</i>			
36	<i>Scenedesmus@quadricauda@var.@quadricauda</i>			
37	<i>Staurastrum@racile</i>			
38	<i>Staurastrum@triangularis@var.@imneticus</i>			
39	<i>Tetraedron@racile</i>			
40	<i>Xanthidium@f. scotti</i>			
<b>EUGLENOPHYTA</b>		41	<i>Strombomonas@biformis@var.@brevicollis</i>	
<b>DINOPHYTA</b>		42	<i>Glenodinium@inctum</i>	
43	<i>Peridinium@inctum</i>			
<b>TOTAL</b>		<b>43</b>		

Station	Sampling Date	Analysis Date	No	SPECIES
C	3/11/2011	24/11/2011	<b>CYANOPHYTA</b>	
			1	<i>Aphanocapsa@elicatissima</i>
			2	<i>Arthrospira@assortii</i>
			3	<i>Microcystis@eruginosa</i>
			4	<i>Oscillatoria@mosa</i>
5	<i>Oscillatoria@tenus</i>			
<b>CHRYSOPHYTA</b>		6	<i>Dinobryon@setularia</i>	
<b>BACILLARIOPHYTA</b>		7	<i>Amphiprorata@lata</i>	
8	<i>Biddulphia@sp.</i>			
9	<i>Campylodiscus@oemelianus</i>			
10	<i>Coscinodiscus@subtilis</i>			
11	<i>Cyclotella@triatra</i>			
12	<i>Cyclotella@omta</i>			
13	<i>Chaetoceros@orenzianus</i>			
14	<i>Diploneis@bombus</i>			
15	<i>Diploneis@mithii</i>			
16	<i>Gyrosigma@fasciola</i>			
17	<i>Gyrosigma@inensis</i>			
18	<i>Melosira@granulata</i>			
19	<i>Melosira@granulata@var.@angussima</i>			
20	<i>Nitzschia@losterium</i>			
21	<i>Nitzschia@sigma</i>			
22	<i>Pleurosigma@bestuarii</i>			
23	<i>Pleurosigma@affine</i>			
24	<i>Surirella@gemma</i>			
25	<i>Surirella@obustata@var.@splendida</i>			
26	<i>Synedra@lina</i>			
<b>CHLOROPHYTA</b>		27	<i>Actinastrum@bantzchii@var.@bantzchii</i>	
28	<i>Ankistrodesmus@spiralis</i>			
29	<i>Closterium@juncidum</i>			
30	<i>Closterium@kuetzingii</i>			
31	<i>Closterium@naciculum</i>			
32	<i>Coelastrum@amblicum</i>			
33	<i>Cosmarium@contractum</i>			
34	<i>Cosmarium@moniliforme</i>			
35	<i>Dictyosphaerium@pulchellum</i>			
36	<i>Eunotia@legans</i>			
37	<i>Hyalothece@quacosa</i>			
38	<i>Micrasterium@borhemiense</i>			
39	<i>Micrasterium@quadrisetum@var.@quadrisetum</i>			
40	<i>Pandorina@horum</i>			
41	<i>Pediastrum@duplex@var.@duplex</i>			
42	<i>Pediastrum@duplex@var.@reticulatum</i>			
43	<i>Pediastrum@oryanum@var.@longicorne</i>			
44	<i>Scenedesmus@bliquus@var.@alternans</i>			
45	<i>Scenedesmus@cuminatus@var.@cuminatus</i>			
46	<i>Scenedesmus@quadricauda@var.@quadricauda</i>			
47	<i>Scenedesmus@quadricauda@var.@quadricauda</i>			
48	<i>Staurastrum@triangularis@var.@imneticus</i>			
49	<i>Staurastrum@racile</i>			
<b>EUGLENOPHYTA</b>		50	<i>Spondyliolum@planum</i>	
51	<i>Staurastrum@jejectum</i>			
52	<i>Staurastrum@racile</i>			
53	<i>Staurastrum@sp.</i>			
54	<i>Staurastrum@triatratum@var.@triatratum</i>			
55	<i>Tetraedron@racile</i>			
56	<i>Xanthidium@f. scotti</i>			
<b>EUGLENOPHYTA</b>		57	<i>Euglenia@pirogyra</i>	
58	<i>Lepocinclis@bium</i>			

Station	Sampling Date	Analysis Date	No	SPECIES
D	3/11/2012	25/11/2012	<b>CYANOPHYTA</b>	
			1	<i>Aphanocapsa@elicatissima</i>
			2	<i>Merismopedial@tenuissima</i>
			3	<i>Microcystis@eruginosa</i>
			4	<i>Oscillatoria@perornata</i>
5	<i>Pseudonitzschia@moniliformis</i>			
<b>CHRYSOPHYTA</b>		6	<i>Dinobryon@setularia</i>	
<b>BACILLARIOPHYTA</b>		7	<i>Amphiprorata@lata</i>	
8	<i>Biddulphia@sp.</i>			
9	<i>Coscinodiscus@radiatus</i>			
10	<i>Cyclotella@omta</i>			
11	<i>Lauderia@nulata</i>			
12	<i>Melosira@granulata</i>			
13	<i>Melosira@granulata@var.@angussima</i>			
14	<i>Nitzschia@losterium</i>			
15	<i>Nitzschia@sigma</i>			
16	<i>Pleurosigma@bestuarii</i>			
17	<i>Surirella@gemma</i>			
18	<i>Surirella@obustata@var.@splendida</i>			
19	<i>Synedra@lina</i>			
<b>CHLOROPHYTA</b>		20	<i>Actinastrum@bantzchii@var.@bantzchii</i>	
21	<i>Ankistrodesmus@spiralis</i>			
22	<i>Ankistrodesmus@spiralis</i>			
23	<i>Arthrodesmus@convergens</i>			
24	<i>Closterium@racile</i>			
25	<i>Closterium@kuetzingii</i>			
26	<i>Closterium@setocum</i>			
27	<i>Coelastrum@amblicum</i>			
28	<i>Cosmarium@moniliforme</i>			
29	<i>Crucigeldium@quadrata</i>			
30	<i>Desmidiium@ballayi</i>			
31	<i>Dictyosphaerium@pulchellum</i>			
32	<i>Eunotia@legans</i>			
33	<i>Hyalothece@quacosa</i>			
34	<i>Micrasterium@borhemiense</i>			
35	<i>Micrasterium@quadrisetum@var.@quadrisetum</i>			
36	<i>Oryctonema@laeve</i>			
37	<i>Pandorina@horum</i>			
38	<i>Pediastrum@duplex@var.@duplex</i>			
39	<i>Pediastrum@duplex@var.@reticulatum</i>			
40	<i>Pediastrum@oryanum@var.@longicorne</i>			
41	<i>Scenedesmus@cuminatus@var.@longatus</i>			
42	<i>Scenedesmus@cuminatus@var.@longatus</i>			
43	<i>Scenedesmus@quadricauda@var.@quadricauda</i>			
44	<i>Staurastrum@triangularis@var.@imneticus</i>			
45	<i>Staurastrum@jejectum</i>			
46	<i>Staurastrum@racile</i>			
47	<i>Staurastrum@odulosum</i>			
48	<i>Staurastrum@sp.</i>			
49	<i>Staurastrum@triangularis@var.@imneticus</i>			
<b>EUGLENOPHYTA</b>		50	<i>Euglenia@pirogyra</i>	
51	<i>Phacus@longicauda</i>			
52	<i>Phacus@leuconectes@var.@leuconectes</i>			
53	<i>Phacus@leuconectes@var.@leuconectes</i>			
54	<i>Strombomonas@biformis@var.@brevicollis</i>			
<b>DINOPHYTA</b>		55	<i>Ceratium@irunidella</i>	
56	<i>Peridinium@inctum</i>			
<b>TOTAL</b>		<b>56</b>		

61	<i>Xanthidium@f. scotti</i>
<b>DINOPHYTA</b>	
62	<i>Alexandrium@f. affine</i>
63	<i>Peridinium@inctum</i>
<b>DICTYOCOPHYTA</b>	
64	<i>Dictyocha@fibula</i>
<b>TOTAL</b>	
	<b>64</b>

59	<i>Phacus@longicauda</i>
60	<i>Phacus@leuconectes@var.@leuconectes</i>
61	<i>Strombomonas@biformis@var.@brevicollis</i>
62	<i>Trachelomonas@rmata</i>
63	<i>Trachelomonas@volvocina</i>
<b>DINOPHYTA</b>	
64	<i>Alexandrium@f. affine</i>
65	<i>Ceratium@irunidella</i>
66	<i>Peridinium@inctum</i>
67	<i>Peridinium@sp.</i>
<b>TOTAL</b>	
	<b>67</b>

Station	E	Sampling Date	13/11/2011
		Analysis Date	25/11/2011
No	SPECIES		
CYANOPHYTA			
1	<i>Microcystis aeruginosa</i>		
2	<i>Oscillatoria limosa</i>		
3	<i>Pseudonitzschia f. moniliformis</i>		
CHRYSOPHYTA			
4	<i>Dinobryon salutaria</i>		
BACILLARIOPHYTA			
5	<i>Amphiprora lata</i>		
6	<i>Asteromphalus leveanus</i>		
7	<i>Biddulphia</i> sp.		
8	<i>Cocconeis scutellum</i>		
9	<i>Coscinodiscus radiatus</i>		
10	<i>Cyclotella striata</i>		
11	<i>Chaetoceros lorenzianus</i>		
12	<i>Chaetoceros parvodus</i>		
13	<i>Chaetoceros pseudo-curveletus</i>		
14	<i>Chaetoceros</i> sp.		
15	<i>Diploneis mithii</i>		
16	<i>Ditylum brightwellii</i>		
17	<i>Gyrodinium aureolum</i>		
18	<i>Melosira granulata</i>		
19	<i>Melosira granulata</i> var. <i>ingussima</i>		
20	<i>Nitzschia oosterium</i>		
21	<i>Nitzschia sigma</i>		
22	<i>Odontella regia</i>		
23	<i>Odontella sinensis</i>		
24	<i>Petrolia granulata</i>		
25	<i>Pleurosigma bestuarii</i>		
26	<i>Rhizosolenia imbricata</i>		
27	<i>Suirella robusta</i> var. <i>splendida</i>		
28	<i>Synedra lina</i>		
29	<i>Thalassionema frauenfeldii</i>		
30	<i>Thalassionema nitzschoides</i>		
CHLOROPHYTA			
31	<i>Actinastrum hantzschii</i> var. <i>hantzschii</i>		
32	<i>Kirchneriella lunaris</i>		
33	<i>Coelastrum amblicum</i>		
34	<i>Cosmarium</i> sp.		
35	<i>Eunotia legans</i>		
36	<i>Micrasterium borhemiense</i>		
37	<i>Micrasterium quadrisetum</i> var. <i>quadrisetum</i>		
38	<i>Oocystis borgei</i>		
39	<i>Pediastrum duplex</i> var. <i>reticulatum</i>		
40	<i>Pediastrum tetras</i> var. <i>tetras</i>		
41	<i>Scenedesmus cuminatus</i> var. <i>biseratus</i>		
42	<i>Scenedesmus arcuatus</i> var. <i>arcuatus</i>		
43	<i>Scenedesmus quadricauda</i> var. <i>quadricauda</i>		
44	<i>Scenedesmus</i> sp.		
45	<i>Staurastrum lejeunei</i>		
46	<i>Staurastrum gracile</i>		
47	<i>Staurastrum nodulosum</i>		
48	<i>Staurastrum senarium</i>		
49	<i>Staurastrum striolatum</i> var. <i>striolatum</i>		
50	<i>Staurastrum wildemanii</i>		
51	<i>Staurastrum triangularis</i> var. <i>imneticus</i>		
52	<i>Tetraedron gracile</i>		
53	<i>Tetrastrum heterocanthum</i>		
EUGLENOPHYTA			
54	<i>Euglena acus</i>		
55	<i>Phacus longicauda</i>		
56	<i>Phacus leuconectes</i> var. <i>leuconectes</i>		
57	<i>Strombomonas papiformis</i> var. <i>brevicollis</i>		
DINOPHYTA			
58	<i>Ceratium hirundella</i>		
59	<i>Dictyochaeta bula</i>		
TOTAL	59		

Survey Tran Dinh Minh, November 18th, 2011

Site	Station	T/B	Depth (m)	Turbidity	Temp (°C)	Flow Velocity (m/s)		Bottom Sediment Type	Total Solids (TSS)	Suspended Solids (SS)	pH	Salinity (ppt)	DO (mg/l)	NO <sub>2</sub> (mg/l)	NO <sub>3</sub> (mg/l)	NH <sub>4</sub> (mg/l)	Alkalinity (KH)	Total P	Total N	Chlorophyll a (mg/m <sup>3</sup> / µg/l)	Zooplankton Biomass		Phytoplankton (cells/l)	Microalgae	Toxic Algae (cell/l)	Total Coliforms	E. coli	Vibrio		
						Tide (Flood)	Tide (Ebb)														unit/m <sup>3</sup>	g/m <sup>3</sup>								
A	T			0.70	29.0						7.20	1.0	6.00																	
	B	1.1						Sand						<0.02	<0.05	<0.02	40			1.20						43000	0			
B	T			0.76	29.0						7.40	0.5	5.50																	
	B	1.5						Mud/Sand						<0.02	<0.05	<0.02	30			3.68						15000	93000			
C	T			0.00	29.0						7.20	0.5	6.00																	
	B	0.7						Sand/Mud						<0.02	<0.05	<0.02	40			3.10						9000	93000			
D	T			0.72	29.0						7.20	0.5	6.00																	
	B	1.8						Mud/Sand						<0.02	<0.05	<0.02	20/30			4.90						7000	0			
E	T			0.84	29.0						7.40	1.5	5.50																	
	B	0.9						Sand/Mud						<0.02	<0.05	<0.02	30			2.29						0	0			



Survey Tran Dinh Minh, November 28th 2011 Phytoplankton Species

Station A	Sampling Date: 8/11/2011
No	SPECIES
<b>CYANOPHYTA</b>	
1	<i>Arthrospira</i> sp.
2	<i>Microcystis</i> sp.
3	<i>Microcystis</i> sp.
4	<i>Oscillatoria</i> sp.
5	<i>Pseudonitzschia</i> sp.
<b>BACILLARIOPHYTA</b>	
6	<i>Amphirostralella</i>
7	<i>Bacillaria</i> sp.
8	<i>Cocconeis</i> sp.
9	<i>Cocconeis</i> sp.
10	<i>Cocconeis</i> sp.
11	<i>Diploneis</i> sp.
12	<i>Lyrella</i> sp.
13	<i>Mastogloia</i> sp.
14	<i>Melosira</i> sp.
15	<i>Melosira</i> sp.
16	<i>Nitzschia</i> sp.
17	<i>Nitzschia</i> sp.
18	<i>Nitzschia</i> sp.
19	<i>Odontella</i> sp.
20	<i>Pleurosigma</i> sp.
21	<i>Pleurosigma</i> sp.
22	<i>Pleurosigma</i> sp.
23	<i>Rhizosolenia</i> sp.
24	<i>Surirella</i> sp.
25	<i>Synedra</i> sp.
26	<i>Thalassionema</i> sp.
<b>CHLOROPHYTA</b>	
27	<i>Actinastrum</i> sp.
28	<i>Closterium</i> sp.
29	<i>Closterium</i> sp.
30	<i>Coelastrum</i> sp.
31	<i>Crucigenia</i> sp.
32	<i>Eunotia</i> sp.
33	<i>Oocystis</i> sp.
34	<i>Pandorina</i> sp.
35	<i>Pediastrum</i> sp.
36	<i>Pediastrum</i> sp.
37	<i>Pediastrum</i> sp.
38	<i>Scenedesmus</i> sp.
39	<i>Staurastrum</i> sp.
40	<i>Staurastrum</i> sp.
41	<i>Staurastrum</i> sp.
42	<i>Staurastrum</i> sp.
43	<i>Tetrastrum</i> sp.
44	<i>Xanthium</i> sp.
<b>DINOPHYTA</b>	
45	<i>Ceratium</i> sp.
<b>TOTAL</b>	<b>45</b>

Station B	Sampling Date: 8/11/2012
No	SPECIES
<b>CYANOPHYTA</b>	
1	<i>Aphanocapsa</i> sp.
2	<i>Aphanocapsa</i> sp.
3	<i>Microcystis</i> sp.
4	<i>Microcystis</i> sp.
5	<i>Oscillatoria</i> sp.
<b>BACILLARIOPHYTA</b>	
6	<i>Amphirostralella</i>
7	<i>Asteromphalus</i> sp.
8	<i>Bellerophon</i> sp.
9	<i>Biddulphia</i> sp.
10	<i>Coloneis</i> sp.
11	<i>Campylodiscus</i> sp.
12	<i>Cocconeis</i> sp.
13	<i>Cocconeis</i> sp.
14	<i>Cocconeis</i> sp.
15	<i>Cocconeis</i> sp.
16	<i>Cocconeis</i> sp.
17	<i>Cyclotella</i> sp.
18	<i>Gyrosigma</i> sp.
19	<i>Gyrosigma</i> sp.
20	<i>Lyrella</i> sp.
21	<i>Melosira</i> sp.
22	<i>Melosira</i> sp.
23	<i>Nitzschia</i> sp.
24	<i>Nitzschia</i> sp.
25	<i>Odontella</i> sp.
26	<i>Pleurosigma</i> sp.
27	<i>Pleurosigma</i> sp.
28	<i>Stauroneis</i> sp.
29	<i>Surirella</i> sp.
30	<i>Surirella</i> sp.
31	<i>Synedra</i> sp.
32	<i>Trachinesis</i> sp.
<b>CHLOROPHYTA</b>	
33	<i>Actinastrum</i> sp.
34	<i>Closterium</i> sp.
35	<i>Closterium</i> sp.
36	<i>Closterium</i> sp.
37	<i>Closterium</i> sp.
38	<i>Coelastrum</i> sp.
39	<i>Cosmarium</i> sp.
40	<i>Coelastrum</i> sp.
41	<i>Eunotia</i> sp.
42	<i>Oocystis</i> sp.
43	<i>Oocystis</i> sp.
44	<i>Pandorina</i> sp.
45	<i>Pediastrum</i> sp.
46	<i>Pediastrum</i> sp.
47	<i>Scenedesmus</i> sp.
48	<i>Scenedesmus</i> sp.
49	<i>Scenedesmus</i> sp.
50	<i>Scenedesmus</i> sp.
51	<i>Staurastrum</i> sp.
52	<i>Staurastrum</i> sp.
53	<i>Staurastrum</i> sp.
54	<i>Staurastrum</i> sp.
<b>DINOPHYTA</b>	
55	<i>Alexandrium</i> sp.
56	<i>Ceratium</i> sp.
57	<i>Dinophysis</i> sp.
<b>DICTYOCOPHYTA</b>	
58	<i>Dictyocha</i> sp.
<b>TOTAL</b>	<b>58</b>

Station C	Sampling Date: 8/11/2011
No	SPECIES
<b>CYANOPHYTA</b>	
1	<i>Aphanocapsa</i> sp.
2	<i>Aphanocapsa</i> sp.
3	<i>Microcystis</i> sp.
4	<i>Oscillatoria</i> sp.
5	<i>Oscillatoria</i> sp.
<b>BACILLARIOPHYTA</b>	
6	<i>Amphirostralella</i>
7	<i>Asteromphalus</i> sp.
8	<i>Campylodiscus</i> sp.
9	<i>Cocconeis</i> sp.
10	<i>Cocconeis</i> sp.
11	<i>Cyclotella</i> sp.
12	<i>Chaetoceros</i> sp.
13	<i>Diploneis</i> sp.
14	<i>Diploneis</i> sp.
15	<i>Gyrosigma</i> sp.
16	<i>Gyrosigma</i> sp.
17	<i>Gyrosigma</i> sp.
18	<i>Gyrosigma</i> sp.
19	<i>Lauderia</i> sp.
20	<i>Lyrella</i> sp.
21	<i>Mastogloia</i> sp.
22	<i>Melosira</i> sp.
23	<i>Melosira</i> sp.
24	<i>Navicula</i> sp.
25	<i>Nitzschia</i> sp.
26	<i>Nitzschia</i> sp.
27	<i>Surirella</i> sp.
28	<i>Odontella</i> sp.
29	<i>Petronella</i> sp.
30	<i>Petronella</i> sp.
31	<i>Petronella</i> sp.
32	<i>Pleurosigma</i> sp.
33	<i>Pleurosigma</i> sp.
34	<i>Rhizosolenia</i> sp.
35	<i>Surirella</i> sp.
36	<i>Surirella</i> sp.
37	<i>Synedra</i> sp.
38	<i>Thalassionema</i> sp.
39	<i>Thalassionema</i> sp.
<b>CHLOROPHYTA</b>	
40	<i>Actinastrum</i> sp.
41	<i>Ankistrodesmus</i> sp.
42	<i>Closterium</i> sp.
43	<i>Closterium</i> sp.
44	<i>Coelastrum</i> sp.
45	<i>Cosmarium</i> sp.
46	<i>Eunotia</i> sp.
47	<i>Pediastrum</i> sp.
48	<i>Pediastrum</i> sp.
49	<i>Pediastrum</i> sp.
50	<i>Scenedesmus</i> sp.
51	<i>Scenedesmus</i> sp.
52	<i>Scenedesmus</i> sp.
53	<i>Staurastrum</i> sp.
54	<i>Staurastrum</i> sp.
55	<i>Staurastrum</i> sp.
56	<i>Tetraedron</i> sp.
57	<i>Tetrastrum</i> sp.
<b>EUGLENOPHYTA</b>	
58	<i>Euglena</i> sp.
59	<i>Phacus</i> sp.
<b>DINOPHYTA</b>	
60	<i>Ceratium</i> sp.
61	<i>Pratoperidium</i> sp.
<b>TOTAL</b>	<b>61</b>

Station D	Sampling Date: 8/11/2011
No	SPECIES
<b>CYANOPHYTA</b>	
1	<i>Aphanocapsa</i> sp.
2	<i>Microcystis</i> sp.
3	<i>Pseudonitzschia</i> sp.
4	<i>Pseudonitzschia</i> sp.
<b>BACILLARIOPHYTA</b>	
5	<i>Amphirostralella</i>
6	<i>Asteromphalus</i> sp.
7	<i>Cocconeis</i> sp.
8	<i>Cocconeis</i> sp.
9	<i>Cocconeis</i> sp.
10	<i>Cyclotella</i> sp.
11	<i>Chaetoceros</i> sp.
12	<i>Diploneis</i> sp.
13	<i>Grammatophora</i> sp.
14	<i>Gyrosigma</i> sp.
15	<i>Lyrella</i> sp.
16	<i>Mastogloia</i> sp.
17	<i>Melosira</i> sp.
18	<i>Navicula</i> sp.
19	<i>Navicula</i> sp.
20	<i>Nitzschia</i> sp.
21	<i>Nitzschia</i> sp.
22	<i>Odontella</i> sp.
23	<i>Pleurosigma</i> sp.
24	<i>Pleurosigma</i> sp.
25	<i>Rhizosolenia</i> sp.
26	<i>Surirella</i> sp.
27	<i>Synedra</i> sp.
28	<i>Thalassionema</i> sp.
29	<i>Thalassionema</i> sp.
<b>CHLOROPHYTA</b>	
30	<i>Actinastrum</i> sp.
31	<i>Arthrodesmus</i> sp.
32	<i>Closterium</i> sp.
33	<i>Closterium</i> sp.
34	<i>Closterium</i> sp.
35	<i>Coelastrum</i> sp.
36	<i>Coelastrum</i> sp.
37	<i>Cosmarium</i> sp.
38	<i>Crucigenia</i> sp.
39	<i>Dictyosphaerium</i> sp.
40	<i>Hyalotheca</i> sp.
41	<i>Orychonema</i> sp.
42	<i>Pandorina</i> sp.
43	<i>Pediastrum</i> sp.
44	<i>Pediastrum</i> sp.
45	<i>Scenedesmus</i> sp.
46	<i>Scenedesmus</i> sp.
47	<i>Scenedesmus</i> sp.
48	<i>Scenedesmus</i> sp.
49	<i>Scenedesmus</i> sp.
50	<i>Scenedesmus</i> sp.
51	<i>Spondylolum</i> sp.
52	<i>Staurastrum</i> sp.
53	<i>Staurastrum</i> sp.
54	<i>Staurastrum</i> sp.
55	<i>Staurastrum</i> sp.
56	<i>Staurastrum</i> sp.
57	<i>Staurastrum</i> sp.
58	<i>Tetraedron</i> sp.
<b>EUGLENOPHYTA</b>	
59	<i>Phacus</i> sp.
<b>DINOPHYTA</b>	
61	<i>Ceratium</i> sp.
62	<i>Peridinium</i> sp.
<b>DICTYOCOPHYTA</b>	
63	<i>Dictyocha</i> sp.
<b>TOTAL</b>	<b>63</b>

Station#	Sampling#	18/11/2011
	Analysis#	21/11/2011
No	SPECIES	
CYANOPHYTA		
1	<i>Aphanocapsa@elicatissima</i>	
2	<i>Microcystis@berghiana</i>	
3	<i>Microcystis@botrys</i>	
4	<i>Microcystis@bairdii</i>	
5	<i>Microcystis@essenbergii</i>	
6	<i>Pseudonitzschia@nonifliformis</i>	
BACILLARIOPHYTA		
7	<i>Amphiroa@lata</i>	
8	<i>Bacillaria@paxillifera</i>	
9	<i>Biddulphia@sp.</i>	
10	<i>Caloneis@fenzlii</i>	
11	<i>Coscinodiscus@pennsylvanicus</i>	
12	<i>Coscinodiscus@radiatus</i>	
13	<i>Cyclotella@koma</i>	
15	<i>Gyrodinium@fasciola</i>	
16	<i>Gyrodinium@inensis</i>	
17	<i>Melosira@granulata</i>	
18	<i>Melosira@granulata@var.@angustissima</i>	
19	<i>Nitzschia@lasterium</i>	
20	<i>Nitzschia@bozzaniana</i>	
21	<i>Nitzschia@sigma</i>	
22	<i>Odotella@ongicruric</i>	
23	<i>Pleurosigma@bestuarii</i>	
24	<i>Pleurosigma@angulatum</i>	
25	<i>Thalassionema@nitzschoides</i>	
CHLOROPHYTA		
26	<i>Actinastrum@bantzchii@var.@bantzchii</i>	
27	<i>Coelastrum@amblicum</i>	
28	<i>Coelastrum@phaenicum</i>	
29	<i>Crucigenia@quadrata</i>	
30	<i>Dictyosphaerium@bulbellum</i>	
31	<i>Hyalotheca@nucosa</i>	
32	<i>Micrasterium@barthemiense</i>	
33	<i>Pediastrum@duplex@var.@duplex</i>	
34	<i>Pediastrum@duplex@var.@reticulatum</i>	
35	<i>Scenedesmus@cuminatus@var.@cuminatus</i>	
36	<i>Spondylusium@planum</i>	
37	<i>Staurastrum@ejectum</i>	
38	<i>Staurastrum@gracile</i>	
39	<i>Staurastrum@odulosum</i>	
40	<i>Staurastrum@wildemanii</i>	
41	<i>Stauradesmus@triangularis@var.@imneticus</i>	
EUGLENOPHYTA		
42	<i>Trachelomonas@volvocina</i>	
DINOPHYTA		
43	<i>Ceratium@irunidella</i>	
DICTYOCOPHYTA		
44	<i>Dictyocha@bula</i>	
TOTAL	44	

Survey Tran Dinh Minh, November 23rd, 2011

Site	Station	T/B	Depth(m)	Turbidity	Temp(°C)	Flow velocity(m/s)	Tide	Bottom type	Total solids(TSS)	Suspended (SS)	pH	Salinity (ppt)	DO(mg/l)	NO <sub>2</sub> (mg/l)	NO <sub>3</sub> (mg/l)	NH <sub>4</sub> (mg/l)	Alkalinity (kh)	Total P	Total N	Chlorophyll a (mg/m <sup>3</sup> = ug/l)	Zoobenthos biomass (unit/m <sup>2</sup> g/m <sup>2</sup> )	Phytoplankton (cells/l)	Microalgae	Toxic algae (cell/l)	Total coliforms	E.coli	Vibrio																										
A	T	M	1.6	0.54	27.0		Flood	Sand			8.70	32.0	6.00	<0.02	>0.05	<0.02	600000			4.56						0	0																										
																													M	1.6	1.39	27.0		Sand			8.40	32.0	5.00	<0.02	>0.05	<0.02	600000			3.74					0	0	
	M	2.5	0.66	27.0		Mud&Sand			8.30	32.0	6.00	<0.02	>0.05	<0.02	700010			3.60					0	0																													
																										B	0.8	0.58	27.0		Sand&Mud			8.30	32.0	5.50	<0.02	>0.05	<0.02	600000			4.94						0	0			

Survey of Tran Dinh Minh, November 23th 2011 Phytoplankton Species

Station A	Sampling Date: 23/11/2011
No	SPECIES
CYANOPHYTA	
1	Trichodesmium erythraeum
BACILLARIOPHYTA	
2	Amphiproralata
3	Amphora strearia var. strearia
4	Asteromphalus leveanus
5	Asterionellopsis facialis
6	Bacteriastrium tomosum
7	Bacteriastrium tomosum var. hispida
8	Biddulphia sp.
9	Campylodiscus titanianus var. anibaricus
10	Campylodiscus lecorus var. innatus
11	Chaetoceros leciapiens
12	Chaetoceros leuciculus
13	Chaetoceros leucivertus
14	Chaetoceros indicus
15	Chaetoceros brentianus
16	Chaetoceros paradoxus
17	Chaetoceros peruvianus
18	Chaetoceros pseudo-curvisetus
19	Chaetoceros sp.
20	Cyclotella triata
21	Coscinodiscus bulliens
22	Coconeis scutellum
23	Corethron pennatum
24	Coscinodiscus entralis
25	Coscinodiscus agos
26	Diplois fusca
27	Dactylosolenia bayanus
28	Dityllum brightwellii
29	Dityllum sol
30	Eucampia odacus
31	Guinardia triata
32	Lauderia nulata
33	Lyrella lavalae var. indica
34	Navicula garrensis var. americana
35	Navicula sp.
36	Nitzschia sigma
37	Odontella nobilensis
38	Odontella regia
39	Odontella longicurric
40	Paralia sulcata
41	Pleurosigma bestuarii
42	Pleurosigma affine
43	Rhizosolenia tomosa
44	Rhizosolenia sp.
45	Podosira telligera
46	Skeletonema costatum
47	Suriella bonanica
48	Thalassionema frauenfeldii
49	Thalassionema itzschoides
50	Triceratium foveum
51	Thalassiosira concentrica
DINOPHYTA	
52	Ceratium furca
53	Ceratium richoceros
54	Diplopelta symmetrica
55	Dinophysis laudata
56	Dinophysis billes
57	Protoperidium bolognum
58	Protoperidium pentagonum
DICTYOCOPHYTA	
59	Dictyochaeta bula
TOTAL	59

Station B	Sampling Date: 23/11/2012
No	SPECIES
CYANOPHYTA	
1	Trichodesmium erythraeum
BACILLARIOPHYTA	
2	Amphiproralata
3	Amphora spectabilis
4	Asteromphalus leveanus
5	Asterionellopsis facialis
6	Bacteriastrium furcatum
7	Bacteriastrium yalinum var. princeps
8	Biddulphia sp.
9	Campylodiscus lecorus var. innatus
10	Coloneis nearis
11	Chaetoceros affine
12	Chaetoceros leucivertus
13	Chaetoceros leciapiens
14	Chaetoceros diversus
15	Chaetoceros brentianus
16	Chaetoceros peruvianus
17	Chaetoceros pseudo-curvisetus
18	Chaetoceros sp.
19	Cyclotella triata
20	Coscinodiscus bulliens
21	Coconeis scutellum
22	Coscinodiscus gonesianus
23	Coscinodiscus sp.
24	Diplois fusca
25	Diplois smithii
26	Diplois saba
27	Dactylosolenia bayanus
28	Dityllum brightwellii
29	Dityllum sol
30	Guinardia triata
31	Hemiaulus sinensis
32	Lauderia nulata
33	Lithodesmium undulatum
34	Menieria membranacea
35	Navicula garrensis var. americana
36	Navicula cf. breta
37	Odontella nobilensis
38	Odontella regia
39	Odontella longicurric
40	Odontella longicurric
41	Petronia granulata
42	Pleurosigma bestuarii
43	Pleurosigma salinarum
44	Pleurosigma sp.
45	Proboscia lata
46	Pseudonitzschia sp.
47	Rhizosolenia bergonii
48	Rhizosolenia tomosa
49	Podosira telligera
50	Skeletonema costatum
51	Thalassionema frauenfeldii
52	Thalassionema itzschoides
53	Thalassiosira concentrica
DINOPHYTA	
54	Ceratium furca
55	Ceratium furca
56	Ceratium richoceros
57	Dinophysis billes
58	Protoperidium bolognum
59	Protoperidium bolognum
DICTYOCOPHYTA	
60	Dictyochaeta bula
CHLOROPHYTA	
61	Pediastrum duplex var. duplex
TOTAL	61

Station C	Sampling Date: 23/11/2011
No	SPECIES
CYANOPHYTA	
1	Trichodesmium erythraeum
BACILLARIOPHYTA	
2	Amphiproralata
3	Asteromphalus leveanus
4	Asterionellopsis facialis
5	Bacteriastrium furcatum
6	Bacteriastrium yalinum var. princeps
7	Bellerophon orotogicalis
8	Biddulphia sp.
9	Campylodiscus lecorus var. innatus
10	Campylodiscus brightwellii
11	Chaetoceros leucivertus
12	Chaetoceros leciapiens
13	Chaetoceros leuciculus
14	Chaetoceros bidymus
15	Chaetoceros diversus
16	Chaetoceros indicus
17	Chaetoceros brentianus
18	Chaetoceros paradoxus
19	Chaetoceros peruvianus
20	Chaetoceros pseudo-curvisetus
21	Cyclotella triata
22	Coscinodiscus bulliens
23	Coconeis scutellum
24	Coscinodiscus entralis
25	Coscinodiscus gonesianus
26	Diplois smithii
27	Dactylosolenia bayanus
28	Dityllum brightwellii
29	Dityllum sol
30	Guinardia triata
31	Gyrosigma balticum
32	Gyrosigma fasciola
33	Hemiaulus membranaceus
34	Hemiaulus sinensis
35	Hemiaulus sp.
36	Lauderia nulata
37	Lyrella lavalae
38	Melosira nonifliformes
39	Melosira granulata
40	Nitzschia brentiana
41	Odontella nobilensis
42	Odontella regia
43	Odontella sinensis
44	Petronia granulata
45	Pleurosigma bestuarii
46	Pleurosigma salinarum
47	Pleurosigma angulatum
48	Pleurosigma lectum
49	Pleurosigma sp.
50	Proboscia lata
51	Pseudonitzschia sp.
52	Pseudosolenia talcaensis
53	Rhizosolenia sp.
54	Rhizosolenia styliiformis
55	Podosira telligera
56	Skeletonema costatum
57	Thalassionema frauenfeldii
58	Thalassionema itzschoides
59	Thalassiosira concentrica
DINOPHYTA	
60	Ceratium furca
61	Dinophysis laudata
DICTYOCOPHYTA	
62	Dictyochaeta bula
TOTAL	62

Station D	Sampling Date: 23/11/2011
No	SPECIES
CYANOPHYTA	
1	Trichodesmium erythraeum
BACILLARIOPHYTA	
2	Amphiproralata
3	Amphora spectabilis
4	Asteromphalus leveanus
5	Asterionellopsis facialis
6	Bacteriastrium tomosum var. hispida
7	Bellerophon orotogicalis
8	Chaetoceros affine
9	Chaetoceros leucivertus
10	Chaetoceros debilis
11	Chaetoceros leciapiens
12	Chaetoceros leuciculus
13	Chaetoceros diversus
14	Chaetoceros indicus
15	Chaetoceros brentianus
16	Chaetoceros paradoxus
17	Chaetoceros peruvianus
18	Chaetoceros pseudo-curvisetus
19	Climacodinium concavum
20	Cyclotella triata
21	Coscinodiscus bulliens
22	Coscinodiscus entralis
23	Coscinodiscus gonesianus
24	Coscinodiscus sp.
25	Diplois tomus
26	Dactylosolenia bayanus
27	Dityllum brightwellii
28	Dityllum sol
29	Gramatophora marina
30	Guinardia triata
31	Gyrosigma fasciola
32	Hemiaulus membranaceus
33	Hemiaulus sinensis
34	Lauderia nulata
35	Melosira granulata
36	Navicula garrensis var. americana
37	Nitzschia sigma
38	Odontella regia
39	Odontella sinensis
40	Petronia granulata
41	Pleurosigma bestuarii
42	Pleurosigma salinarum
43	Pleurosigma sp.
44	Proboscia lata
45	Pseudonitzschia sp.
46	Pseudosolenia talcaensis
47	Rhizosolenia bergonii
48	Rhizosolenia tomosa
49	Podosira telligera
50	Skeletonema costatum
51	Suriella robusta var. splendida
52	Thalassionema frauenfeldii
53	Thalassionema itzschoides
DINOPHYTA	
54	Ceratium furca
55	Ceratium furca
56	Ceratium richoceros
57	Dinophysis laudata
58	Protoperidium bolognum
59	Protoperidium bolognum
60	Protoperidium pentagonum
61	Protoperidium bolognum
DICTYOCOPHYTA	
62	Dictyochaeta bula
CHLOROPHYTA	
63	Pediastrum duplex var. duplex
TOTAL	63

Station#	Sampling Date: 23/11/2012
	Analysis Date: 28/11/2012
No	SPECIES
CYANOPHYTA	
1	<i>Trichodesmium erythraeum</i>
BACILLARIOPHYTA	
2	<i>Amphiprora bilata</i>
3	<i>Asteromphalus leuevenus</i>
4	<i>Asteromphalus bellatus</i>
5	<i>Asterionellopsis lacliale</i>
6	<i>Bacteriastrum omosum</i>
7	<i>Bacteriastrum hyalinum</i> var. <i>princeps</i>
8	<i>Campylodiscus wecorus</i> var. <i>pinnatus</i>
9	<i>Campylodiscus brightwellii</i>
10	<i>Campylodiscus haemelianus</i>
11	<i>Coloneis linearis</i>
12	<i>Chaetoceros oarctatus</i>
13	<i>Chaetoceros luvisetus</i>
14	<i>Chaetoceros debilis</i>
15	<i>Chaetoceros lecipiens</i>
16	<i>Chaetoceros lenticulatus</i>
17	<i>Chaetoceros liversus</i>
18	<i>Chaetoceros korenzianus</i>
19	<i>Chaetoceros paradoxus</i>
20	<i>Chaetoceros peruvianus</i>
21	<i>Chaetoceros pseudo-curvisetus</i>
22	<i>Chaetoceros</i> sp.
23	<i>Cyclotella triata</i>
24	<i>Coconeis scutellum</i>
25	<i>Coscinodiscus breslianus</i>
26	<i>Coscinodiscus</i> sp.
27	<i>Diploneis fusca</i>
28	<i>Dactylosolenia lavyanus</i>
29	<i>Ditylum sal</i>
30	<i>Gyrosigma sinensis</i>
31	<i>Hemialus</i> sp.
32	<i>Lauderia annulata</i>
33	<i>Lyrella yroides</i>
34	<i>Mastogloia quinquecostata</i>
35	<i>Melosira nonifliformes</i>
36	<i>Melosira granulata</i>
37	<i>Odontella regia</i>
38	<i>Odontella sinensis</i>
39	<i>Odontella longicurric</i>
40	<i>Paralia sulcata</i>
41	<i>Petronella granulata</i>
42	<i>Pleurosigma bestuarii</i>
43	<i>Pleurosigma angulatum</i>
44	<i>Pleurosigma tectum</i>
45	<i>Pleurosigma</i> sp.
46	<i>Pseudonitzschia</i> sp.
47	<i>Rhizosolenia bergonii</i>
48	<i>Rhizosolenia choctea</i>
49	<i>Podocira stelligera</i>
50	<i>Skeletonema costatum</i>
51	<i>Surirella robusta</i> var. <i>splendida</i>
52	<i>Thalassionema frauenfeldii</i>
53	<i>Thalassionema tzschoioides</i>
54	<i>Thalassiosira leptopus</i>
55	<i>Trachinella bebyi</i>
DINOPHYTA	
56	<i>Ceratium furca</i>
57	<i>Ceratium fusus</i>
58	<i>Dinophysis audata</i>
59	<i>Dinophysis niles</i>
60	<i>Protoperidinium divergens</i>
61	<i>Protoperidinium oceanicum</i>
62	<i>Protoperidinium roseum</i>
DICTYOCOPHYTA	
63	<i>Dictyochaeta bula</i>
CHLOROPHYTA	
64	<i>Staurastrum gracile</i>
TOTAL	64

Environmental Survey - first quarter 2015 (spring, beginning of the production season) (analyses carried out in the Hue Centre for Environmental Monitoring)																																		
Site	Station	T/R	Depth(m)	Turbidity	Temp(°C)	Flow velocity (m/s)	Tides (Flood)	Tides (Ebb)	Bottom sediment type	Total (TS) (SS)	Suspended (SS)	pH	Salinity (ppt)	DO2 (mg/l)	NO3- (mg/l)	NO2- (mg/l)	NH4+ (mg/l)	Alkalinity (KH)	Total P	Total N	Chlorophyll a (mg/L)	Zooplankton biomass (unit/m <sup>3</sup> )	g/m <sup>3</sup>	Phytoplankton (cell/L)	Microalgae	Toxic (cell/L)	Total coliforms (MPN/100mL)	E. coli	Vibrio					
Tam Giang	Ntq1	T		4.4						12.8		7.30	5.40			0.280																		
		M																																
	Ntq2	T		6.0							4.6		7.30	5.70			0.530																	
		M																																
	Ntq3	T		12.6							3.8		7.40	5.60			0.420																	
		M																																
	Ntq4	T		6.1							2.8		7.40	5.20			0.400																	
		M																																
	Ntq5	T		3.3							6.4		7.90	5.70			0.260																	
		M																																
Ntq6	T		7.1							7.8		7.90	5.70			0.080																		
	M																																	
Ngien Binh	Ndb1	T		9.4						7.8		8.70	7.30			0.040																		
		M																																
	Ndb2	T		13.0							22		8.60	8.70			0.037																	
		M																																
	Ndb3	T		26.5							14.4		7.60	7.00			0.070																	
Ndb4	T		20.6							12.8		7.40	7.10			0.120																		
	M																																	
Cau Hai	Ndc1	T		36.1						10.2		9.20	7.20			0.120																		
		M																																
	Ndc2	T		26.0							14		8.90	6.50			0.037																	
		M																																
Ndc3	T		8.6							5		9.10	9.30			0.310																		
	M																																	
Ndc4	T		24.2							9		7.90	6.40			0.070																		



Environmental Survey Average Values 2015 (analyses carried out in the Hue Centre for Environmental Monitoring)																																
Site	Station	T/R	Depth(m)	Turbidity	Temp(°C)	Flow Velocity(m/s)	Tide2 (Flood)	Tide3 (Ebb)	Bottom Sediment Type	Total Solids (TSS)	Suspended Solids (SS)	pH	Salinity (ppt)	DO2 (mg/l)	NO2 (mg/l)	NO3 (mg/l)	NH4 (mg/l)	Alkalinity (kh)	Total P	Total N	Chlorophyll (a) (mg/m <sup>3</sup> - WPL)	ZooBenthos/Biomass	Phytoplankton (cells/l)	Microalgae	Toxicity (µg/l)	Total Coliforms (MPN/100ml)	E. coli	Vibrio				
Tam Giang	Nptg1	T		20.6								7.10	5.30																			
		M								20.4																					297.5	
		B																														
	Nptg2	T		34.4									7.10	5.20				0.21													625	
		M								11.9																						
		B																														
	Nptg3	T		22.2									7.10	4.70																	423.8	
		M								15.2																						
		B																														
	Nptg4	T		15.0									7.10	5.00																	389.5	
		M								7.9																						
		B																														
Nptg5	T		13.2									7.80	5.30																	66		
	M								6.9																							
	B																															
Nptg6	T		11.3									7.80	5.40																	92.8		
	M								8.6																							
	B																															
Huyen Th	Ndh1	T		34.0								8.10	5.90																	527.5		
		M								9.1																						
		B																														
	Ndh2	T		27.5									8.90	8.30																		622
		M								12.8																						
		B																														
Ndh3	T		21.4									7.90	6.70																	3216		
	M								8.9																							
	B																															
Ndh4	T		26.5									7.60	6.30																	1890		
	M								8.6																							
	B																															
Cau Hai	Ndh1	T		29.6								9.00	8.50																	781.5		
		M								7.2																						
		B																														
	Ndh2	T		27.1									8.40	6.90																	265.3	
		M								13.3																						
		B																														
Ndh3	T		14.6									8.50	8.50																	421.5		
	M								6.8																							
	B																															
Ndh4	T		23.0									8.20	6.90																	970.3		
	M																															
	B									9.7																						

## Annex 2 – Comprehensive datasets of surveys 2015-2018, primary data collected during the implementation of this PhD project.

Environmental survey, January 3-4th 2016 (analyses carried out in the Hue Centre for Environmental Monitoring)																												
Site	Station	T/B	Depth(m)	Turbidity	Temp(°C)	FlowVelocity(m/s) TideH TideL	TideH TideL	BottomSedimentType	TotalSolid (TSS)	Suspende dSolid(s)	pH	Salinity (ppt)	DOE (mg/l)	NO <sub>3</sub> (mg/l)	NO <sub>2</sub> (mg/l)	NH <sub>4</sub> (mg/l)	Alkalinity (KH)	TotalP	TotalN	Chlorophylla (mg/m <sup>3</sup> · µg/l)	ZooBenthosBiomass unit/m <sup>2</sup> g/m <sup>3</sup>	Phytoplank tonCell(s/l)	Microalgae	ToxicAlgae (cell/l)	Total coliforms (MPN/100ml)	E.Coli	Vibrio	
Phuoc Phuoc Phuoc Phuoc Phuoc Phuoc	D1	T						silt-sand(?)	192.0	192.0	7.86	7.9	8.75	0.00	0.00	0.00	3000ppm	0.24	<1.4	4.02		205792		125758	1100000	11.0	41.0	
		M	155	1.35	25.78																							
	D2	T							sand	283.0	283.0	8.29	16.3	9.74	0.00	0.00	0.00	400ppm	0.08	<1.4	2.15		127050		51638	460000	2.4	<1
		M	180	1.3	25.40																							
	D3	T							sand(?)	58.0	58.0	8.81	6.9	8.84	0.00	0.00	0.00	300ppm	0.36	2.8	2.80		79354		76978	210000	2.4	>1
		M	55	0.55	25.50																							
D5	T							(?)	145.0	145.0	9.38	10.0	14.28	0.25	0.25	0.25	400ppm	0.23	<1.4	4.18		91664		28694	1500000	110.0	20.0	
	M	140	1	24.00																								
D6	T							(?)	23.0	23.0	9.26	8.6	14.80	0.00	0.00	0.00	400ppm	0.11	<1.4	3.72		54940		28289	430000	460.0	23.0	
	M	135	1.35	25.36																								
D7	T							(?)	197.5	197.5	6.7	9.6	12.24	0.00	0.00	0.00	400ppm	0.34	<1.4	4.20		56971		55592	2900000	1100.0	44.0	
	M	130	1.3	24.13																								
Phuoc Phuoc Phuoc	D8	T						silt-mud(?)	49.0	49.0	8.5	7.0	9.71	0.00	0.00	0.25	400ppm	0.11	<1.4	3.1		58088		56167	43000	4.6	11.0	
		M	120	1.2	25.23																							
Phuoc Phuoc Phuoc	D9	T						silt-mud(?)	64.5	64.5	7.94	6.9	9.61	0.00	0.00	0.00	300ppm	0.59	7.0	3.60		142042		98634	240000	4.6	>1	
		M	130	1.2	25.63																							

Environmental survey, first quarter 2016 (spring, beginning of the production season) (analyses carried out in the Hue Centre for Environmental Monitoring)																														
Site	Station	T/B	Depth(m)	Turbidity	Temp(°C)	FlowVelocity(m/s) TideH TideL	TideH TideL	BottomSedimentType	TotalSolid (TSS)	Suspende dSolid(s)	pH	Salinity (ppt)	DOE (mg/l)	NO <sub>3</sub> (mg/l)	NO <sub>2</sub> (mg/l)	NH <sub>4</sub> (mg/l)	Alkalinity (KH)	TotalP	TotalN	Chlorophylla (mg/m <sup>3</sup> · µg/l)	ZooBenthosBiomass unit/m <sup>2</sup> g/m <sup>3</sup>	Phytoplank tonCell(s/l)	Microalgae	ToxicAlgae (cell/l)	Total coliforms (MPN/100ml)	E.Coli	Vibrio			
TamEiang	Np1g1	T																												
		M		13.2	20.9					10.8		5.50	4.60		0.380												93			
	Np1g2	T																												
		M		22.2	20.8					8.4		6.10	4.50		0.296													150		
	Np1g3	T																												
		M		23.9	20.7					14		5.30	5.30		0.564													290		
Np1g4	T																													
	M		27.1	20.8					12.4		5.80	5.90		0.170													28			
Np1g5	T																													
	M		14.7	21.7					10.4		7.60	5.60		0.060													21			
Np1g6	T																													
	M		10.5	21.2					8.2		7.70	5.20		<0.017													43			
NuyenBinh	Np2h1	T																												
		M		3.9	21.6					17.6		8.20	4.10		0.420													75		
	Np2h2	T																												
		M		0.1	21.8					5		8.00	5.00		0.840													2100		
	Np2h3	T																												
		M		6.5	21.3					8.8		7.60	6.90		0.220														240	
Np2h4	T																													
	M		5.2	22.2					6.6		7.50	4.50		0.400														150		
CauBinh	Np3h1	T																												
		M		6.4	20.8					5		7.80	4.00		0.220													460		
	Np3h2	T																												
		M		5.5	20.5					6		7.70	4.20		0.620														240	
	Np3h3	T																												
		M		8.5	21.4					11.4		7.60	4.00		0.210														2100	
Np3h4	T																													
	M		5.0	21.5					4.6		7.70	4.20		0.140														460		

Environmental survey, second quarter 2016 (summer, during the production season) (analyses carried out in the Hue Centre for Environmental Monitoring)																														
Site	Station	T/B	Depth(m)	Turbidity	Temp(°C)	FlowVelocity(m/s) TideH TideL	TideH TideL	BottomSedimentType	TotalSolid (TSS)	Suspende dSolid(s)	pH	Salinity (ppt)	DOE (mg/l)	NO <sub>3</sub> (mg/l)	NO <sub>2</sub> (mg/l)	NH <sub>4</sub> (mg/l)	Alkalinity (KH)	TotalP	TotalN	Chlorophylla (mg/m <sup>3</sup> · µg/l)	ZooBenthosBiomass unit/m <sup>2</sup> g/m <sup>3</sup>	Phytoplank tonCell(s/l)	Microalgae	ToxicAlgae (cell/l)	Total coliforms (MPN/100ml)	E.Coli	Vibrio			
TamEiang	Np1g1	T																												
		M		28.5						2.3		7.10	5.30		<0.009	0.050												290		
	Np1g2	T																												
		M		28.7						2		6.90	5.50		<0.009	0.075													93	
	Np1g3	T																												
		M		28.9						4.7		7.00	4.80		<0.009	<0.017													460	
Np1g4	T																													
	M		29.0						4.7		7.00	5.10		<0.009	<0.017													240		
Np1g5	T																													
	M		28.6						2.7		7.10	4.70		<0.009	<0.017													150		
Np1g6	T																													
	M		28.4						10		7.00	4.60		<0.009	<0.017													240		
NuyenBinh	Np2h1	T																												
		M		33.2						6.5		7.90	5.50		<0.009	0.540													23	
	Np2h2	T																												
		M		32.1						11		8.60																		

