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Soil bacterial communities under slash and burn in Mozambique as revealed by a metataxonomic approach

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(Article begins on next page)

1 Running Title: SOIL BACTERIA COMMUNITIES UNDER SLASH AND BURN

2

3 **Soil Bacteria Communities under Slash and Burn in Mozambique as Revealed**  
4 **by Metataxonomic Approach**

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12 ABSTRACT

13 The “slash and burn” system is a subsistence agronomical practice widely spread in tropical areas  
14 all over the world. This system has been deeply studied, especially for its impacts on agronomical  
15 aspects and soil physicochemical properties, while the knowledge on their microbial diversity is  
16 scarce. In the present study, for the first time the soil bacterial diversity of three locations from central  
17 Mozambique where “slash and burn” has been practiced with different duration of the forest fallow  
18 period ( $\approx 25$ , 35, and  $\approx 50$  years) has been elucidated through a metataxonomic approach. Bacterial  
19 communities were evaluated on genetic horizons of soils under charcoal kiln, crop field, and forest.  
20 The aim of this study was to examine the influence of spatial (location and land use), temporal (forest  
21 fallow period), and vertical (horizons) variations in selecting bacterial populations in relation to the  
22 physico-chemical properties of the soil. Metataxonomic analysis detected 25 different phyla whose  
23 distribution varied horizontally and vertically in relation to soil properties: pH, easily oxidizable  
24 organic carbon, total nitrogen, and available phosphorous, but also particle-size distribution and  
25 mineralogical composition. Such properties were strongly affected and altered by land use  
26 management; in particular, charcoal kilns showed better soil properties and the greatest differences  
27 in microbial community with respect to crop field and forest, which were quite similar. This might  
28 suggest the inability of a forest fallow period shorter than 50 years to improve soil fertility and induce  
29 changes in microbial community. The uncommon application of the pedologic approach for microbial  
30 evaluation has allowed detecting a clear separation in microbiota composition along the soil profile,  
31 with eutrophic bacteria mainly located in the A horizons, while oligotrophic bacteria abounded in the  
32 Bo horizons. Considering horizontal and vertical heterogeneity in the same study represent a novelty  
33 for bacteria metataxonomic analysis.

34

35 *Key Words:* 16S rRNA gene sequencing, agroforestry, land-use change, soil microbiota, soil  
36 physicochemical properties

## 37 INTRODUCTION

38 The agroforestry system known as “slash and burn” is largely practiced by smallholder farmers  
39 in tropical and subtropical regions (Mertz *et al.*, 2009a; FAO, 2015; Kukla *et al.*, 2018) and consists  
40 of occupying a piece of land, slashing, and burning vegetation in order to convert forest into  
41 agricultural fields (Gay-des-Combes, 2017b). During the conversion, some ephemeral charcoal kilns  
42 (3-6 per hectare) are arranged to produce charcoal for the family; thus, after 2-4 years of cultivation,  
43 the area is abandoned to natural reforestation until it will be slashed and burnt again after decades  
44 (Kabisa and Ncheengamwa, 2020). The forest fallow period is requested to allow soil fertility to  
45 recover before being further cultivated since no fertilizer is used in this farming system (Drexler,  
46 2020). Once the fallow period lasted 50-100 years but, because of the demographic and economic  
47 changes over the last 4-5 decades, the cycle has been progressively shortened to one to a few decades  
48 (Jakovac *et al.*, 2016; Gay-des-Combes *et al.*, 2017a).

49 The slash and burn system often occurs on scarcely fertile soils (mainly Oxisols) and contributes  
50 accelerating their degradation (Styger *et al.*, 2007; Xu *et al.*, 2019). Therefore, slash and burn has  
51 been considered unsustainable since it favours deforestation, loss of biodiversity, soil depletion, and  
52 erosion (Kleinman *et al.*, 1995; Nath *et al.*, 2016; Gay-des-Combes *et al.*, 2017b). Soils subjected to  
53 this practice have been widely studied for their variable physicochemical properties and fertility levels  
54 (Juo and Manu, 1996; Thomaz *et al.*, 2014; Thomaz, 2018), whereas their microbial communities  
55 have been scarcely investigated (Nourou Sall *et al.*, 2006; Sul *et al.*, 2013; Saliou Sarr *et al.*, 2019).  
56 Microbial diversity and activity are very susceptible to ecosystem variations due to natural factors  
57 and/or anthropic activity, but the biotic functionality of the system is still hard to assess and  
58 understand (Nannipieri *et al.*, 2017). In detail, bacterial community diversity is strongly correlated  
59 with the nature of the parent material and soil physicochemical properties such as structure, texture,  
60 water holding capacity, nutrient availability, and organic matter content (Ulrich and Becker, 2006;  
61 Lauber *et al.*, 2008; Sofo *et al.*, 2019). Thus, to assess and understand the soil biotic functionality of  
62 the slash and burn system, it is mandatory to consider soil physicochemical properties and microbial

63 diversity according to land use (spatial variation), duration of the forest fallow (temporal variation)  
64 and, within each soil, the nature of genetic horizons (vertical variation).

65 Based on these premises, we hypothesised that, notwithstanding centuries of slash and burn that could  
66 have homogenized all the system, bacterial community can differentiate horizontally (location and  
67 land use) or vertically (horizons). Therefore, the aims of the study were to evaluate the bacterial  
68 diversity through a metataxonomic approach in soils subjected to slash and burn and influenced by  
69 spatial (location and land use), temporal (forest fallow period), and vertical (horizons) variations  
70 correlated to the physicochemical properties of the soil. For testing this, we selected three locations  
71 of central Mozambique submitted to slash and burn where soil samples were collected under charcoal  
72 kiln, agricultural field, and forest (spatial variations). The locations were selected on the basis of the  
73 forests age, so to obtain a chronosequence driven by the duration of the forest fallow (temporal  
74 variation). The novelty of this research is that both horizontal and vertical heterogeneity was  
75 considered at once in the same study.

76

## 77 MATERIALS AND METHODS

### 78 *Study areas*

#### 79 *Agro-ecological and vegetation characterization*

80 The zone selected for the study is part of the Manica Province, central Mozambique (Fig. S1,  
81 see Supplementary Material for Figure S1). Here, we selected three locations with high agricultural  
82 potential where slash and burn is very common and going on **for** centuries: Vanduzi, Sussundenga,  
83 and Macate (Fig. S1). Based on climatic conditions, soil type, elevation, and farming system, these  
84 districts are located in the Agro-Ecological Zone R4, which includes lands between 200 and 1000 m  
85 above sea level (Maria and Yost, 2006). The mean annual rainfall of the zone ranges from 1000 to  
86 1200 mm, while the mean annual air temperature is  $\approx 21$  °C, with February as the warmest month  
87 (24.2 °C) and July as the coldest one (16.0 °C) (Climate-Data, 2019). The soil moisture regime is  
88 *aridic*, and the soil temperature regime is *thermic* (Soil Survey Staff, 2014). Following the Köppen-

89 Geiger updated climate classification, the climate of the zone is humid subtropical with a cool to mild  
90 season from April to September and a hot and humid season from October to March (Kottek *et al.*,  
91 2006; Belda *et al.*, 2014). The geology of the zone is dominated by metamorphic rocks of the  
92 Mesoproterozoic Southern Irumide Belt (950--1060 Ma) litho-tectonic unit (Cháuque *et al.*, 2019).  
93 The predominant soil type of the zone belongs to the order of Oxisols, with low fertility and a strong  
94 erosion due to the topography of the terrain (Maria and Yost, 2006). The main food crops are maize  
95 (*Zea mais* L.), sorghum (*Sorghum vulgare* Pers), millet (*Panicum miliaceum* L.), and beans. At the  
96 three locations, the forest conditions were generally poor in terms of plant biodiversity. As witnessed  
97 by the presence of several charcoal kiln rests (even more than 20 per hectare), the forests have been  
98 growing up on abandoned crop fields forming the so called *miombo* biome. This latter is typical of  
99 tropical woodland (open forest) comprising savannas and shrublands made of sparse trees with a more  
100 or less thick grass understorey (Siteo, 2004). The *miombo* was made of an upper stratum mainly  
101 composed of the leguminous trees *Brachystegia spiciformis* Benth., *Brachystegia tamarindoides*  
102 Benth., and *Julbernardia globiflora* (Benth.) Troupin, with an understorey composed of herbaceous  
103 species like *Themeda triandra* Forssk., *Panicum maximum* Jacq., *Hyparrhenia filipendula* (Hochst.)  
104 Stapf, and *Andropogon gayanus* Kunth. At Vanduzi there were also a few old mango trees (*Mangifera*  
105 *indica* L.), remainders of an abandoned mango orchard. After abandonment of the fields, a slight  
106 exploitation of the reforesting ranges was maintained because they represent the source of subsistence  
107 goods like timber, poles, firewood, foods, medicines, grazing, leaf litter, and game (Chidumayo *et*  
108 *al.*, 1996; Dewees *et al.*, 2011).

109

#### 110 *The studied slash and burn systems*

- 111 ● Vanduzi

112 Information on Vanduzi was obtained by interviewing local leaders and field owners.  
113 According to them, the charcoal kiln had been arranged four years before the survey The crop field  
114 was settled one year before the survey with an intercropping system of different varieties of banana

115 tree (*Musa paradisiaca* L.), horse radish tree (*Moringa oleifera* Lam.), and sorghum. On the basis of  
116 the information gathered, the forest was  $\approx 25$  years old.

117         • Sussundenga

118             Information about Sussundenga was also obtained by interviewing the landowner. The charcoal  
119 kiln had been used in the year of survey, while the crop field had been cultivated with maize for two  
120 years. Detailed information about the age of the actual forests was obtained from the Sussundenga  
121 Research Station at the *Instituto de Investigação Agrária de Moçambique* (IIAM/CZC). Here,  
122 documents attest the field-adjacent forest was cut in 1982, consequently in 2017 it was 35 years old  
123 and a portion of this forest was cut again in February 2017 to be cultivated.

124         • Macate

125             Information about Macate was also obtained by interviewing local leaders and field owners.  
126 The charcoal kiln was 16 years old; the crop field had been consecutively cultivated with maize for  
127 16 years, and the field-adjacent forest was  $\approx 50$  years old.

128 To resume, the land use chronosequence followed the order: at Vanduzi the field was 1-year old and  
129 the forest was  $\approx 25$  years old; at Sussundenga, the field was 2 years old and the forest was 35 years  
130 old; at Macate, the field was 16 years old and the forest was  $\approx 50$  years old. For charcoal kilns it was  
131 not possible to obtain an increasing order of age for the same sequence of locations being the kiln 4  
132 years old at Vanduzi, less than 1 year at Sussundenga, and 16 years at Macate. To prove the age of  
133 the forests, being useless the counting of tree rings, we ascertained that the average tree diameters of  
134 the ubiquitous *Brachystegia spiciformis* trees of Macate (33 cm) was higher compared with that of  
135 Sussundenga (26 cm) and Vanduzi (16 cm) trees.

136

137 *Study sites and soil sampling*

138             In March 2017, in each area a geomorphological and soil survey was run in order to select the  
139 sampling sites. At each area we selected a rather flat area (plateau) with gentle slope (2-4%), with

140 mostly Oxisols developed from similar metamorphic parent rocks: granitoid rock (possibly gneissic-  
141 granite) at Vanduzi and Sussundenga (Cháuque *et al.*, 2019; Wijnhoud, 1997), and a migmatitic  
142 paragneiss at Macate (Cháuque *et al.*, 2019). In all cases, each soil was characterized by two master  
143 horizons: a brownish A horizon (umbric) and a reddish Bo (oxic) horizon (Table S1, see  
144 Supplementary Material for Table S1). In each area, for any land use (charcoal kiln, agricultural field,  
145 and forest) we selected two representative sites with similar micro-topography and, for the forest,  
146 vegetation. Since Oxisols are very weathered soils and the mean temperature of the area slightly differ  
147 among seasons, to evaluate eventual differences in terms of bacterial community along the year, we  
148 chose to run two sampling campaigns following the most different agricultural seasons: crop end in  
149 March 2017 (Autumn) and field preparation for seeding in November 2017 (Spring). In the charcoal  
150 kilns the profiles were opened in the middle of their extension, while those in the agricultural fields  
151 were opened at  $\approx 25$  m from the border with forest. In this latter, profiles were opened at  $\approx 1$  m from  
152 the trunk of one of the biggest trees of *Brachystegia spiciformis*. The maximum distance among  
153 sampling sites was about 30 m at Sussundenga and Macate, while at Vanduzi forest and field sites  
154 were about 700 m distant. For each sampling campaign, the position where to dig the soil profiles  
155 was selected after opening several manual mini-pits and auger holes. Once excavated, each profile  
156 was described according to Schoeneberger *et al.* (2012) and sampled by genetic horizons (A and Bo).  
157 A large amount of sample (about 4 kg) was collected from each horizon. The amount of profiles  
158 excavated was 9 (3 land uses x 3 locations) in March and 9 in November, for a total of 18 profiles  
159 and 36 horizon samples.

160 Samples were collected in sterilized polyethylene bags and stored at  $\approx 4$  °C inside a portable fridge  
161 during the field operations. Once in the laboratory, the samples were air-dried and then passed through  
162 a sieve (2 mm mesh) to remove the skeletal particles and coarse vegetal residues.

163



164 *Physicochemical and mineralogical analyses*

165 The pH was determined potentiometrically in H<sub>2</sub>O after one night of solid:liquid contact,  
166 using a combined glass-calomel electrode immersed into the suspension (1:2.5 solid:liquid ratio).  
167 Particle-size distribution was determined after dissolution of organic cements by NaClO at pH 9  
168 (Lavkulich and Wiens, 1970). Sand (2-0.05 mm) was recovered by wet sieving, while silt was  
169 separated from clay by sedimentation maintaining the columns at 19-20 °C. The amount of easily  
170 oxidizable organic carbon (EOOC) was estimated by the Walkley-Black method by K-dichromate  
171 digestion without heating (Nelson and Sommers, 1996). The total nitrogen (N) content was  
172 determined by the semi-micro Kjeldahl method and potentially plant-available phosphorous (P) was  
173 determined according to Olsen *et al.* (1954). The mineralogical assemblage was assessed by X-ray  
174 diffractometry on manually compressed powdered samples by using a Philips PW 1830  
175 diffractometer (Fe-filtered Co K $\alpha$ 1 radiation, 35 kV and 25 mA). Minerals were identified on the  
176 basis of their characteristic peaks (Brindley and Brown, 1980; Dixon and Schulze, 2002), while a  
177 semi-quantitative mineralogical composition was obtained by estimating the area of the diagnostic  
178 peaks by multiplying the peak height by its width at half-height.

179

180 *Microbial DNA extraction and sequencing*

181 Total microbial DNA was extracted from 100 mg of each soil sample using the E.Z.N.A.<sup>®</sup>  
182 Soil DNA Kit (Omega Bio-Tek, Inc., Georgia, USA) following the manufacturer's instruction. DNA-  
183 based analysis was preferred to mRNA analysis because in complex matrices like soil, RNA can be  
184 rapidly degraded by RNAases, with a consequent less reliability of the soil microbial composition  
185 (Nannipieri *et al.*, 2020). The extracted DNA was quantified by using the Qubit dsHS kit (Thermo  
186 Fisher, Milan, Italy) and standardized at 25 ng  $\mu$ L<sup>-1</sup>. One  $\mu$ l of each DNA suspension was used as  
187 template for PCR amplification by using primers 16SF (5'-  
188 TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGCCTACGGGNGGCWGCAG-3') and  
189 16SR (5'-

190 GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGGACTACHVGGGTATCTAATCC-3’)  
191 spanning the V3-V4 region of the 16S rRNA gene following the procedure described by Klindworth  
192 *et al.* (2013), and a negative control was included in the PCR reactions by replacing the DNA solution  
193 with water. The PCR amplicons were purified according to the Illumina metagenomic pipeline  
194 instructions. Briefly amplicons were cleaned using the Agencourt AMPure kit (Beckman coulter,  
195 Brea, USA) according to the manufacturer’s instructions; subsequently, DNA concentrations of the  
196 amplicons were determined using the Quant-iT PicoGreen dsDNA kit (Invitrogen Life Technology)  
197 following the manufacturer’s instructions. In order to ensure the absence of primer dimers and to  
198 assay the purity, the quality of generated amplicon libraries was evaluated by a Bioanalyzer 2100  
199 (Agilent, Palo Alto, CA, USA) using the High Sensitivity DNA Kit (Agilent). Following the  
200 quantitation, cleaned amplicons were mixed and combined in equimolar ratios. Paired-end  
201 sequencing (2x250 bp) using the Illumina MiSeq system (Illumina, San Diego, USA) were carried  
202 out at the Sequencing Platforms of the Fondazione Edmund Mach (FEM, San Michele a/Adige, Italy).

203

#### 204 *Bioinformatics analysis*

205 After sequencing, raw reads were merged using Flash software (Magoc and Salzberg, 2011)  
206 and analyzed with QIIME 1.9.0 software (Caporaso *et al.*, 2010); the detailed pipeline was described  
207 by Ferrocino *et al.* (2017). The USEARCH version 11 software (Edgar *et al.*, 2011) was adopted for  
208 chimera filtering, against the 16S reference databases. Centroid sequences of each operational  
209 taxonomic unit (OTU) cluster (at 97 % of similarity) by using UCLUST (Edgar, 2010) were mapped  
210 against Greengenes 16S rRNA gene database by means of the RDP Classifier, with a minimum  
211 confidence score of 0.80 (Wang *et al.*, 2007).

212 Centroids sequence were manually blasted to check the taxonomic identification. To avoid biases due  
213 to the different sequencing depths, OTU tables generated through QIIME were rarefied at the lowest  
214 number of reads and showed the highest taxonomy resolution that was reached. Alpha diversity index  
215 (Shannon and Chao1) were calculated by the QIIME alpha diversity script. The data generated by

216 sequencing were deposited in the NCBI Sequence Read Archive (SRA) and are available under the  
217 Bioprojects Accession Number-PRJNA550507 for replicate 1 and PRJNA631872 (biosamples  
218 accession number from SAMN14895357 to SAMN14895411) for replicate 2.

219

#### 220 *Statistical treatment of the data*

221 RStudio program (vv 1.3.1093) (RStudio Team, 2020) was used for statistical analysis. By  
222 ANOVA we assessed that the results obtained from the analyses of samples collected in the two  
223 sampling campaigns in terms of physicochemical properties (pH, particle-size distribution, EOOC,  
224 total N, and available P) did not differ (Table S2, see Supplementary Material for Table S2,  $P > 0.05$ ).  
225 Because of this, the samples collected in the two sampling campaigns were considered as replicates  
226 and ANOVA was run to test significant differences for sampling locations (Vanduzi, Sussundenga,  
227 and Macate), land uses (charcoal kiln, crop field, and forest), and horizons (A and Bo) (Table S3, see  
228 Supplementary Material for Table S3,  $P > 0.05$ ). To apply the ANOVA, we previously verified the  
229 normal distribution of the data and the equal variances. The improvement of the assumption to  
230 normality and homoscedasticity was verified on residuals by the Shapiro-Wilk statistical test (*stats* R  
231 package) (R Core team, 2013) and by Levene's test (*car* R package) (Fox and Weisberg, 2019), both  
232 at 5 % of significance level. When data were non-normally distributed, each numerical variable was  
233 transformed by the Box-Cox procedure (Meloun *et al.*, 2005). If the transformed data were normally  
234 distributed, a post-hoc Tukey's Honest Significant Difference (HSD) test with  $P \leq 0.05$  was used to  
235 compare the means. When normality was not respected, the Kruskal-Wallis test was applied to assess  
236 if the differences were significant. In case of heteroscedasticity, the Welch one-way ANOVA test  
237 was performed. ANOVA tests were deemed significant when  $P \leq 0.05$ . In case of heteroscedasticity  
238 and non-normality, we run the Friedman test (*rstatix* package) (Kassambara, 2020) combined with  
239 Kendall's W to measure the Friedman test effect size and pairwise Wilcoxon signed-rank tests. The  
240 arithmetic means and relative standard deviations for physicochemical properties (Tables I, II, and  
241 III) and OTUs were calculated for sampling locations (n=12), total land use (n=12), land use of each

242 area (n=4), total horizons (n=18), and horizon of each site (n=6). In doing this, technical replicates  
243 were treated as experimental replicates, as it often occurs in ecosystem scale experiments (Osburn *et*  
244 *al.*, 2019). Non-parametric pairwise Wilcoxon tests were used when appropriate to determine the  
245 significant differences of OTU abundance and alpha diversity. Spearman correlation analysis between  
246 OTUs and physicochemical properties was performed through the *psyc* package (Revelle, 2021) and  
247 plotted by using the function *corrplot* of RStudio program (vv 1.3.1093). The *P* values were adjusted  
248 for multiple testing using the Benjamini-Hochberg procedure, which assesses the false discovery rate  
249 (FDR).

250

## 251 RESULTS

### 252 *Soil morphology*

253 In all locations (Vanduzi, Sussundenga, and Macate), the soils were Oxisols due the presence  
254 of diagnostic Bo horizons (Soil Survey Staff, 2014) (Table S1, see Supplementary Material for Table  
255 S1). The A horizons under charcoal kiln showed a charcoal content always higher than 30 %, to  
256 become  $\approx 1$  % in the crop fields and to be absent under forests. The Bo horizons showed a reddish  
257 colour and, especially at Vanduzi, they displayed a relatively high content of Fe-Mn-oxides ( $\approx 5$  %).  
258 In general, both A and Bo horizons presented a good degree of aggregation, with the presence of sub-  
259 angular and angular blocks generally coarser in the A than in the Bo horizons (Table S1, see  
260 Supplementary Material for Table S1). The good state of aggregation, the coarse texture (from loamy  
261 sand to sandy loam), and the absence of any redoximorphic feature indicated these soils are well-  
262 drained and, consequently, with low water-holding capacity (Agrawal, 1991; Suzuki *et al.*, 2007).

263

### 264 *Microbiota diversity*

265 The relative abundances of bacterial taxa were examined at phylum rank to determine whether  
266 there were differences at the scale of location, land use, or horizon (Fig. 1). In total, 25 different phyla  
267 approximately totaled 96.5 % of the bacterial pool, with Actinobacteria (22 %), Proteobacteria (19

268   %), Chloroflexi (17 %), Firmicutes (15 %), Planctomycetes (10 %), Acidobacteria (5 %),  
269   Verrucomicrobia (3 %), Nitrospirae (2 %), and AD3 (1 %) as the most representative by considering  
270   the average relative abundance for all samples. Regarding the minor OTUs fraction, Bacteroidetes,  
271   Gemmatimonadetes, Armatimonadetes, Cyanobacteria, GAL15, Chlamydiae, TM7, OD1, and  
272   Crenarchaeota (relative abundance between 0.1 and 1 %) represented about 3.4 % of the total bacterial  
273   community. At Vanduzi and Macate, for the alpha diversity value we observed the highest number  
274   of OTUs and a higher richness (Chao1 and Shannon index) in the A horizons than in the Bo horizons  
275   (data not shown, FDR < 0.05), while no difference was observed among the land uses. Conversely,  
276   at Sussundenga, the alpha diversity value showed no difference between forest and crop field, which  
277   displayed a higher number of OTUs and a higher richness than the charcoal kiln (FDR < 0.05, data  
278   not shown); no difference was observed between the horizons.

279

#### 280   *Location effect*

281         The soils from Vanduzi showed the highest pH and the highest content of available P, while  
282   EOOC and total N were the greatest at Macate (Table I). Particle-size distribution was always  
283   dominated by the sand fraction and mineralogically by quartz, with minor contents of clay minerals  
284   and plagioclases. Regarding OTUs association, Vanduzi displayed the highest abundance of  
285   Actinobacteria, Firmicutes, Nitrospirae, and WS3; Sussundenga the highest for Firmicutes,  
286   Cyanobacteria, and WS4; Macate showed the highest presence of Chloroflexi, Planctomycetes,  
287   Verrucomicrobia, and WS3 (Fig. S2, see Supplementary Material for Figure S2, FDR < 0.05).  
288   Distributions at a low taxonomical rank were presented in Fig. S3 (see Supplementary Material for  
289   Figure S3) (FDR < 0.05), with the highest abundances summarized in Appendix 1. The results of  
290   bacteria diversity at phylum rank among locations were schematically synthesized in Fig. 2.

291

292 *Land use effect*

293 The soils under charcoal kiln had the highest pH, while both charcoal kilns and forests displayed  
294 the largest available P content (Table II). In all land uses, sand was the most represented separate and  
295 the mineralogy was dominated by quartz. In the different locations, the highest pH values were  
296 observed for the charcoal kilns of Vanduzi and Sussundenga (Table II). The largest available P  
297 content occurred at **Macate** for charcoal kilns, while at Sussundenga and **Vanduzi** there was a scarce  
298 dotation of this nutrient (Table II). Charcoal kilns showed the highest abundance of  
299 Gemmatimonadetes and OD1 and the lowest abundance of Armatimonadetes and Tenericutes. An  
300 opposite trend was observed for the same **taxa** in soils under crop field and forest (Fig. S4, see  
301 Supplementary Material for Figure S4, FDR < 0.05). Looking at each location, soils under charcoal  
302 kiln at Vanduzi abounded in Firmicutes, while at **Sussundenga showed** the lowest abundance of  
303 Planctomycetes (Fig. S5, see Supplementary Material for Figure S5, FDR < 0.05). At Macate, **the**  
304 **soils under charcoal kiln** showed the lowest abundance of **Armatimonadetes**, **while those** under forest  
305 showed the lowest abundance of Chlamydiae (Fig. S5, FDR < 0.05).  
306 At low taxonomical rank, differences of bacterial distribution were displayed in Fig. **S3** (FDR < 0.05),  
307 and briefly reported in Appendix 1, while differences at phylum level among land uses, and among  
308 land uses within location were synthetized in Figs. 2 and 3, respectively.

309

310 *Horizon effect*

311 As a whole, the pH and the contents of EEOC, total N, and available P were higher in the A  
312 compared with the Bo horizons, while, as expected for Oxisols, the clay content was much larger in  
313 the Bo than in the A horizons (Table III,  $P < 0.05$ ). In all the locations, EEOC and total N were the  
314 highest in the A horizons. At Sussundenga and Macate the sand content was the highest in the A  
315 horizon, while the clay abounded in the Bo horizon. Only at Sussundenga the available P abounded  
316 in the A horizons (Table III,  $P < 0.05$ ). Mineralogical assemblage was similar in all situations, with  
317 quartz as the most abundant mineral, always higher in the A than in the Bo horizons, while clay

318 minerals were always higher in the Bo than in the A horizons (Table III). With respect to soil use, in  
319 the charcoal kilns only the pH values were higher in the A than in the Bo horizons. In the crop fields,  
320 EEOC and available P showed the highest contents in the A horizons, while in the forests EEOC and  
321 total N were the largest in the A horizons.

322 By comparing the OTUs composition, the A horizons displayed the largest quantities of  
323 Actinobacteria, Planctomycetes, Verrucomicrobia, Bacteroidetes, Gemmatimonadetes, and TM7,  
324 whereas the Bo horizons displayed the highest abundance of AD3, GAL15, Thermi, and WPS-2 (Fig.  
325 S6, see Supplementary Material for Figure S6, FDR < 0.05). At Vanduzi, Verrucomicrobia and TM7  
326 were found to be the most abundant taxa in the A horizons, while Proteobacteria, Nitrospirae, AD3,  
327 and GAL15 were mainly associated with the Bo horizons (Fig. S7, see Supplementary Material for  
328 Figure S7, FDR < 0.05). At Sussundenga, Actinobacteria and Verrucomicrobia were predominant in  
329 the A horizons, while AD3 and GAL15 abounded in the Bo horizons (Fig. S8, see Supplementary  
330 Material for Figure S8, FDR < 0.05). At Macate, Actinobacteria and WS3 abounded in the A horizons,  
331 while AD3 and GAL15 predominated in the Bo horizons (Fig. S9, see Supplementary Material for  
332 Figure S9, FDR < 0.05). Considering the soil horizons under different land use, the Bo horizons under  
333 charcoal kiln was dominated by AD3 and GAL15 (Fig. S10, see Supplementary Material for Figure  
334 S10, FDR < 0.05). Under crop field, the A horizons were characterized by Actinobacteria,  
335 Bacteroidetes, Gemmatimonadetes, and TM7, while the Bo horizons were dominated by AD3 and  
336 GAL15 (Fig. S11, see Supplementary Material for Figure S11, FDR < 0.05). Under forest, the A  
337 horizons showed the highest abundance of Planctomycetes and Verrucomicrobia, with the Bo  
338 horizons dominated by AD3, Cyanobacteria, GAL15, and Thermi (Fig. S12, see Supplementary  
339 Material for Figure S12, FDR < 0.05).

340 At a low taxonomical rank, differences were displayed in Fig. S3 (FDR < 0.05), and details about the  
341 highest abundances between A and Bo horizons within locations were summarized in Appendix 2.  
342 The bacteria diversity at phylum rank between A and Bo horizons, and between horizons within  
343 location was synthesized in Figs. 2 and 3, respectively.

344

### 345 *Correlation between microbiota and physicochemical properties*

346 By plotting the correlation between OTUs of the most represented phyla and the soil  
347 physicochemical properties (Fig. S13, see Supplementary Material for Figure S13, FDR < 0.05), we  
348 observed that the presence of Actinobacteria was positively associated with available P, while  
349 Chloroflexi was directly associated with clay and inversely with sand, available P, and pH. Firmicutes  
350 were positively associated with pH and sand but inversely correlated with total N. Planctomycetes  
351 was negatively associated with pH and, together with Verrucomicrobia, they were positively  
352 correlated with EEOC, total N, and silt. Armatimonadetes and AD3 resulted negatively correlated  
353 with available P and sand, but positively correlated with clay. Bacteroidetes, Gemmatimonadetes,  
354 and TM7 were directly associated with pH and available P. GAL15 displayed the highest negative  
355 correlation with pH, EEOC, available P, and sand, and were positively correlated with clay, while  
356 OD1 displayed the opposite correlations (FDR < 0.05).

357

## 358 DISCUSSION

### 359 *Location effect*

360 The three locations differed in microbial community abundances for several taxa. In detail,  
361 Actinobacteria phylum (among which *Rubrobacteraceae*, *Streptomycetaceae*, and  
362 *Streptosporangiaceae* were the most abundant families and Micrococcales the most abundant order)  
363 was the dominant in the soils of Vanduzi. Actinobacteria phylum has been widely reported for soils  
364 under various environmental conditions, including Antarctica and Sahara (e.g., Saker *et al.*, 2015;  
365 Tytgat *et al.*, 2016); it is probably the wide adaptability of the species belonging to this phylum the  
366 reason of its abundance in the soils of Vanduzi. Araujo *et al.* (2020) found that some Actinobacteria  
367 taxa abounded in soils near to neutral pH, including *Rubrobacter* genus belonging to  
368 *Rubrobacteraceae* family. Instead, Koyama *et al.* (2014) reported a reduction of Actinobacteria in soils



369 enriched of N, while Prada Salcedo *et al.* (2014) found that some Actinobacteria strains can solubilize  
370 both calcium phosphate and Al-phosphate in acid soils, making P available in solution. Correlation  
371 plot of Fig. S13 showed that Actinobacteria was positively correlated with the available P but, as in  
372 the case of Vanduzi, also with the highest pH values and the lowest total N contents. At Vanduzi there  
373 was also the highest presence of Nitrospirae, specifically of the Nitrospirales order. Vipindas *et al.*  
374 (2020) described Nitrospirae as chemolithoautotrophic bacteria mainly involved in N mineralization,  
375 in particular in the oxidation of nitrite to nitrate. In fact, Wang *et al.* (2018) reported that the nitrate  
376 addition to soil resulted in the decline of Nitrospirae and of the nitrification activity. In addition, Zhou  
377 *et al.* (2015) associated a high presence of Nitrospirae to soils with neutral pH and not fertilized with  
378 N and P. It is therefore conceivable that bacteria of the Nitrospirae group abound in scarcely fertile  
379 soils where they play an important role producing nitrate by nitrite oxidation.

380 Sussundenga soils were characterized by the dominance of Cyanobacteria and WS4.  
381 Cyanobacteria abounded in the Sussundenga soils, where there was the largest quartz content, but  
382 they were scarce at Macate, where quartz was in the lowest quantity. The fact that the different  
383 distribution of quartz may influence Cyanobacteria abundances was ascribed to the adaptation of  
384 these bacteria to arid conditions (Lacap-Bugler *et al.*, 2017), which are well-expressed at the surface  
385 of grain quartz, one of the less hydrophilic silicates in soil because of its lack of isomorphic  
386 substitutions (Tarasevich *et al.*, 2002).

387 At Macate, soils showed the highest presence of Chloroflexi, Verrucomicrobia (among which the  
388 family *Chthoniobacteraceae*) and Planctomycetes (with the family *Gemmataceae*). Various studies  
389 have reported that Chloroflexi are involved in the organic matter decomposition and, consequently in  
390 the C and N cycling (e.g., Hug *et al.*, 2013; Ibrahim *et al.*, 2020). Chloroflexi abounded at Macate,  
391 where there were the highest amounts of EEOC and total N, even though this correlation was not  
392 statistically significant. Instead, at Macate, Verrucomicrobia were positively correlated with the  
393 contents of EEOC, total N, and silt, and the correlations were statistically significant. Similar results  
394 were reported by Buckley and Schmidt (2001), who found a positive correlation between

395 Verrucomicrobia and soil organic carbon, total N, and soil moisture. Also, Planctomycetes are  
396 directly correlated with EEOC, total N and silt, but inversely with pH. Zhao *et al.* (2018) also  
397 observed a significant correlation between soil organic carbon and Planctomycetes abundance.  
398 Firmicutes, represented in large amount by *Paenibacillaceae* and *Bacillaceae* families, abounded at  
399 Vanduzi and Sussundenga and showed a positive correlation with pH and sand content, but negative  
400 with total N. Vos *et al.* (2011) described *Paenibacillaceae* as mesophilic and termophilic, but also as  
401 neutrophilic and alkaliphilic bacteria. Since the soils at Vanduzi and Sussundenga displayed pH  
402 values closed to neutrality and the prevalence of sand particles that favour high temperatures  
403 transmission at depth in case of heat flow (Abu-Hamdeh and Reeder, 2000), we may suppose that  
404 Firmicutes proliferated in these soils because of these physicochemical properties.

405

#### 406 *Land use effect*

407 As expected, charcoal kilns represented a unique ecosystem, with peculiar microbial community  
408 if compared to crop field and forest like, for example, a higher abundance of OD1 and  
409 Gemmatimonadetes. Following the report of Coomes *et al.* (2017), who also found  
410 Gemmatimonadetes in soils under charcoal kiln, and the correlations reported in Fig. S13, we ascribed  
411 the presence of these bacteria in our charcoal kiln soils to the relatively large content of available P  
412 and relatively high pH values. A similar distribution is valid for OD1, which were largely abundant  
413 in charcoal kiln soils and resulted positively correlated with pH, available P, and sand, but negatively  
414 with clay (Fig. S13). Since pH showed the most significant variations between charcoal kiln soils and  
415 crop field/forest soils, we suggest OD1 bacteria are mainly influenced by soil reaction rather than the  
416 other correlated properties. On the contrary, Armatimonadetes were more abundant in crop field and  
417 forest soils than in charcoal kilns and showed a positive correlation with clay but a negative  
418 correlation with available P and sand. These results suggested a predilection of Armatimonadetes for  
419 soils scarce in available P. Moreover, Armatimonadetes have been found to be negatively correlated  
420 with pH but positively correlated with moisture (Tytgat *et al.*, 2016), indicating that soils under

421 charcoal kiln are less preferred by the species of this phylum because of the large content of charcoal,  
422 which commonly supplies soluble P to soil (Rafael *et al.*, 2020) and reduce soil moisture due to the  
423 overheating consequent to the dark colour. Tenericutes mainly abounded in forest soil, with no  
424 significant correlation to physicochemical properties. Lanc *et al.* (2013) reported that Tenericutes  
425 were particularly abundant in soils from Brazilian semi-arid forests during the rainy season. Although  
426 more investigation on this phylum is needed, we suppose Tenericutes proliferation is favoured by the  
427 presence of relatively high soil organic matter content and moisture, conditions that occurred in our  
428 forest soils (Scott and Kleb, 1996).

429 A few microbial differences among land uses were restricted to some locations. For example, at  
430 Vanduzi, Firmicutes abounded in the charcoal kiln area possibly because of i) the high pH values due  
431 to the alkalinising effect of ash and biochar (Fidel *et al.*, 2017) and ii) the sand content that favours  
432 the penetration of high temperatures in soil during charcoal production. As a support of this,  
433 Firmicutes belonging to the Bacillales order abound in soils after wildfire and burning treatments  
434 (Smith *et al.*, 2008; Sul *et al.*, 2013), while bacteria of the *Bacillaceae* family include spore-forming  
435 species able to resist the extremely high temperature (Battistuzzi and Hedges, 2009; Galperin, 2013).  
436 At Sussundenga, Planctomycetes showed the lowest abundance in the charcoal kiln soil. Yang *et al.*  
437 (2020) and Jenkins *et al.* (2017) observed a decrease of Planctomycetes when soil pH increased  
438 following fire or biochar addition. As a demonstration of this, Navarrete *et al.* (2015) reported a higher  
439 abundance of Planctomycetes in forest soils with low pH. Our results agreed with the above-  
440 mentioned studies, being the soil pH at Sussundenga the highest in the charcoal kiln soils and the  
441 relation between Planctomycetes and pH negative (Fig. S13). At Macate, differences were detected  
442 for Armatimonadetes, the least abundant phylum in charcoal kiln soils, and Chlamydiae, the least  
443 abundant in forest soils. We ascribed Chlamydiae distribution to the behaviour of some Chlamydiae  
444 bacteria as pathogens of arthropods (Horn *et al.*, 2004; Wagner and Horn, 2006), including soil  
445 isopods like woodlouse (Collingro *et al.*, 2020). Specifically, soil isopods are Chlamydiae's soil

446 dwelling that generally feed of decaying organic matter (Saska, 2008) including corn litter (Johnson  
447 *et al.*, 2012), which was the major remainders of cultivation in the Macate fields.

448

#### 449 *Horizon effect*

450 The horizon effect has marked a clear separation of the physicochemical properties and  
451 microbiota. The higher abundance of Actinobacteria in the A than in the Bo horizons appeared  
452 correlated with the highest contents of available P, EOO, and total N at Sussundenga and Macate  
453 and in the crop fields. Although Actinobacteria have been associated to soils with low organic carbon  
454 content (Sul *et al.*, 2013; Fu *et al.*, 2019), other studies demonstrated that their optimum growth  
455 substrate is represented by soils rich in organic matter and N, with neutral pH, good soil aeration, and  
456 moderate temperature (e.g., Tang *et al.*, 2016; Liu *et al.*, 2017; Dai *et al.*, 2018), conditions that  
457 mainly attained in the A horizons (Table III). In the soils at Vanduzi, Proteobacteria were the most  
458 abundant in the Bo horizons, probably because these horizons are particularly rich of Fe-Mn nodules  
459 ( $\approx 5\%$ ), and this property could have favoured bacteria of this phylum being Proteobacteria able to  
460 catalyse the Fe-oxidation reactions (Hedrich *et al.*, 2011). Planctomycetes (among which the  
461 *Phycisphaerae* family) abounded in the A horizons under forest, probably because species belonging  
462 to this phylum are involved in carbon and N turnover (Fuerst and Sagulenko, 2011). Like  
463 Planctomycetes, Verrucomicrobia (in detail *Chthoniobacteraceae* family and Pedosphaerales order)  
464 abounded in the A horizons especially at Vanduzi and Sussundenga, and under forest. In our case,  
465 Verrucomicrobia were largely present concomitant with the highest quantities of EOO, total N, and  
466 available P. At this regard, Sangwan *et al.* (2004) and O'Brien *et al.* (2016) recognized  
467 *Chthoniobacteraceae* as utilizers of saccharides derived from plant biomass or engaged in symbiosis  
468 with soil nematodes. Instead, Pedosphaerales were found by Bach *et al.* (2018) to abound in large  
469 macroaggregates rather than in microaggregates. Thus, the large abundance of Verrucomicrobia in  
470 the A horizons was ascribed to their relatively higher organic matter content, which fairly includes  
471 sugars, and the generalized coarser structure.

472 Bacteroidetes (among which the *Chitinophagaceae* family), Gemmatimonadetes, and TM7 abounded  
473 in the A horizons, particularly of the crop fields, and were positively correlated with pH values and  
474 available P (Fig. S13). As reported by Wolińska *et al.* (2017), Bacteroidetes are involved in the  
475 organic matter cycle and, joined with Gemmatimonadetes, they have been found associated with the  
476 degradation of complex organic polymers (Chaudhry *et al.*, 2012). In particular, *Chitinophagaceae*  
477 mainly colonize the rhizosphere rather than the bulk soil (Madhaiyan *et al.*, 2015) and have been  
478 found to be positively correlated with the C:N ratio (Dennis *et al.*, 2019). Furthermore, Zhou *et al.*  
479 (2015) reported of positive correlations between TM7 and the contents of total N, nitrates,  
480 ammonium, and soil organic matter. All this considering, the abundance of Bacteroidetes,  
481 Gemmatimonadetes, and TM7 in the A horizons was ascribed to a predilection for complex organic  
482 substrates with an incipient decaying of organic matter.

483 AD3 and GAL15 were more abundant in the Bo than in the A horizons. Looking at the correlation  
484 plot (Fig. S13), AD3 was directly correlated with clay and inversely correlated with available P and  
485 sand. This distribution was probably due to the general properties of Oxisols, which showed an  
486 increase of acidity and clay with increasing depth. As a support to this, Mesa *et al.* (2017) found  
487 abundant AD3 in biofilms and sediments of acid mine drainage. Also GAL15 resulted to be directly  
488 correlated with clay and inversely correlated with available P and sand, but also with pH and EEOC.  
489 Since the members of these taxa seemed to prefer oligotrophic habitats (e.g., Li *et al.*, 2020; Liu *et*  
490 *al.*, 2020), it is conceivable they diffused in the Bo rather than in the A horizons. Also the phyla  
491 Thermi and WPS-2 abounded in the Bo horizons. Since Thermi were found in hypolithic communities  
492 of Taklimakan Desert in China (Lacap-Bugler *et al.*, 2017) and WPS-2 were more abundant in  
493 unfertilized soils and in oil palm plantation than in primary and regenerated forests (Wood *et al.*,  
494 2017), we hypothesized that the members of these phyla prefer oligotrophic soil conditions, and  
495 consequently mainly inhabit the Bo horizons.

496 Only at Vanduzi, Proteobacteria and Nitrospirae showed a large abundance in the Bo horizons, with  
497 no significant correlation with the soil physicochemical properties (Fig. S13). Similar conditions were

498 found by Hedrich *et al.* (2011), who ascribed to Proteobacteria a high grade of adaptation and the  
499 peculiarity to survive with iron-oxidizing forms in presence of oxygen and preferably with neutral to  
500 acid pH. The diffusion of Nitrospirae in the Bo horizons fitted with their preference to colonize soil  
501 compartments with neutral pH and scarce N.

502

## 503 CONCLUSIONS

504 Oxisols submitted to slash and burn differed in terms of spatial and vertical changes for their  
505 bacterial diversity. Our study suggests that bacteria were affected by soil physicochemical properties  
506 reliant on both soil genesis and human activities. Actinobacteria, Nitrospirae, WS3, Chloroflexi,  
507 Verrucomicrobia, Planctomycetes, and Firmicutes varied among locations in conjunction with  
508 different pHs and nutrients availability, while Cyanobacteria abundance seemed to depend on quartz  
509 content. Also land use determined a strong selection of microbiota in particular under charcoal kilns,  
510 where soil physicochemical properties have been changed by temperature and addition of charcoal  
511 and ash. Gemmatimonadetes, OD1, Armatimonadetes, Firmicutes, and Planctomycetes were also  
512 affected by the presence of the charcoal kiln while, Tenericutes and Chlamydiae proliferated,  
513 respectively, in the soils under forest for the high organic matter content and moisture and in the soil  
514 under crop field at Macate because of mulching practices. Except for Tenericutes, no other significant  
515 difference in terms of taxa abundances and physicochemical properties were encountered between  
516 forests and crop fields, despite the forest fallow might let suppose a considerable soil fertility  
517 restoration – with consequent microbiota change – over time. Remarkable results were found along  
518 the soil profiles, confirming the importance of genetic horizons in determining microbiota  
519 composition. Actinobacteria, Planctomycetes, Verrucomicrobia, Bacteroidetes, Gemmatimonadetes,  
520 TM7, and WS3 were abundant in the A horizons, suggesting a predilection for eutrophic conditions,  
521 while AD3, GAL 15, Thermi, WPS-2, Proteobacteria, and Nitrospirae abounded in oligotrophic Bo  
522 horizons. These results allowed us recognizing two main groups of bacteria: those strongly affected

523 by spatial, temporal, and vertical variations, and those homogeneously distributed in soil  
524 independently from the physicochemical variations among horizons.

525 Our findings contribute to improving the knowledge on spatial, temporal, and vertical soil bacteria  
526 diversity, and dependence of this latter from physicochemical properties in Oxisols. More studies are  
527 needed to better disclose the relationships between microbiota and soil properties.

528

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540

## 541 SUPPLEMENTARY MATERIAL

542 Supplementary material for this article can be found in the online version.

543

## 544 **References**

545 Abu-Hamdeh N H, Reeder R C. 2000. Soil thermal conductivity: effects of density, moisture, salt  
546 concentration, and organic matter. *Soil Sci Soc Am J.* 64: 1285–1290.

547 Agrawal R P. 1991. Water and nutrient management in sandy soils by compaction. *Soil Till Res.* 19:  
548 121-130.

549 Araujo R, Gupta Vadakattu V S R, Reith F, Bissett A, Mele P, Franco C M M. 2020. Biogeography  
550 and emerging significance of Actinobacteria in Australia and Northern Antarctica soils. *Soil Biol*  
551 *Biochem.* 146: 107805.

552 Bach E M, Williams R J, Hargreaves S K, Yang F, Hofmockel K S. 2018. Greatest soil microbial  
553 diversity found in micro-habitats *Soil Biol Biochem.* 118: 217-226.

554 Battistuzzi F U, Hedges S B. 2009. A major clade of Prokaryotes with ancient adaptations to life on  
555 land. *Mol Biol Evol.* 26(2): 335-343.

556 Belda M, Holtanová E, Halenka T, Kalvová J. 2014. Climate classification revisited: from Köppen to  
557 Trewartha. *Climate Res.* 59: 1-13.

558 Brindley G W, Brown G. 1980. Crystal structures of clay minerals and their identification.  
559 Mineralogic Society Monograph No. 5. Mineralogical Society, London.

560 Buckley D H, Schmidt T M. 2001. Environmental factors influencing the distribution of rRNA from  
561 Verrucomicrobia in soil. *FEMS Microb Ecol.* 35: 105-112.

562 Caporaso J G, Kuczynski J, Stombaugh J, Bittinger K, Bushman F D, Costello E K, Knight R, Fierer  
563 N, Peña A G, Goodrich J K, Gordon J I, Huttley G A, Kelley S T, Knights D, Koenig J E, Ley  
564 R E, Lozupone C A, McDonald D, Muegge B D, Pirrung M, Reeder J, Sevinsky J R, Turnbaugh  
565 P J, Walters W A, Widmann J, Yatsunenko T, Zaneveld J, Knight R. 2010. QIIME allows analysis  
566 of high-throughput community sequencing data. *Nature Methods.* 7: 335–336.

567 Chaudhry V, Rehman A, Mishra A, Chauhan P S, Nautiyal C S. 2012. Changes in bacterial  
568 community structure of agricultural land due to long-term organic and chemical amendments.  
569 *Microb Ecol.* 64: 450-460.

570 Chaúque F R, Cordani U G, Jamal D L. 2019. Geochronological systematics for the Chimoio-  
571 Macossa frontal nappe in central Mozambique: Implications for the tectonic evolution of the  
572 southern part of the Mozambique belt. *J African Earth Sci.* 150: 47-67.



573 Chidumayo E N, Gambiza J, Grundy I. 1996. Managing miombo woodland. In Campbell B (ed.) The  
574 Miombo in Transition: Woodlands and Welfare in Africa, chapter 7, CIFOR, Bogor. pp 175-193.  
575 Climate-data. Available online at <https://en.climate-data.org/>.

576 Collingro A, Köstlbacher S, Horn M. 2020. Chlamydiae in the Environment. *Trends Microb.* 28: 877-  
577 888.

578 Coomes O T, Miltner B C. 2017. Indigenous charcoal and biochar production: potential for soil  
579 improvement under shifting cultivation systems. *Land Degr Devel.* 28: 811–821.

580 Dai Z, Su W, Chen H, Barberán A, Zhao H, Yu M, Yu L, Brookes P C, Schadt C W, Chang S X, Xu  
581 J. 2018. Long-term nitrogen fertilization decreases bacterial diversity and favors the growth of  
582 Actinobacteria and Proteobacteria in agro-ecosystems across the globe. *Glob Change Biol.* 24:  
583 3452–3461.

584 Dennis P G, Newsham K K, Rushton S P, O’Donnell A G, Hopkins D W. 2019. Soil bacterial  
585 diversity is positively associated with air temperature in the maritime Antarctic. *Scientific*  
586 *Reports.* 9: 2686.

587 Dewees P, Campbell B, Katerere Y, Siteo A, Cunningham A B, Angelsen A, Wunder S. 2011.  
588 Managing the miombo woodlands of southern Africa: Policies, incentives, and options for the  
589 rural poor. Washington DC, Program on Forests (PROFOR) pp 77.

590 Dixon J B, Schulze S G (Eds.). 2002. Soil Mineralogy with Environmental Applications. Number 7  
591 in the Soil Science Society of America Book Series. *Soil Sci Soc Am*, Inc., Madison, Wisconsin,  
592 USA

593 Drexler, K.A., 2020. Government extension, agroecology, and sustainable food systems in Belize  
594 milpa communities: a socio-ecological systems approach. *J Agric, Food Syst, Commun Devel.*  
595 ISSN: 2152-0801. Available online at <https://www.foodsystemsjournal.org>.

596 Edgar R C. 2010. Search and clustering orders of magnitude faster than BLAST. *Bioinformatics*  
597 26:2460 –2461. <https://doi.org/10.1093/bioinformatics/btq461>.

598 Edgar R C, Haas B J, Clemente J C, Quince C, Knight R. 2011. Applied and Environmental  
599 Microbiology improves sensitivity and speed of chimera detection. *Bioinformatics* 27:2194–  
600 2200. <https://doi.org/10.1093/bioinformatics/btr381>.

601 FAO. 2015. Shifting Cultivation, Livelihood and Food Security. New and Old Challenges for  
602 Indigenous Peoples in Asia. Available online at <http://www.fao.org/3/a-i4580e.pdf>

603 Ferrocino I, Bellio A, Romano A, Macori G, Rantsiou K, Decastelli L, Cocolin L. 2017. RNA-based  
604 amplicon sequencing reveals the microbiota development during ripening of artisanal vs.  
605 industrial Lard d'Arnad. *Appl Environ Microbiol.* 83: 983-17.

606 Fidel R B, Laird D A, Thompson M L, Lawrinenko M. 2017. Chemosphere characterization and  
607 quantification of biochar alkalinity. *Chemosphere.* 167: 367-373.

608 Fox J, Weisberg S. 2019. An R Companion to Applied Regression, Third edition. Sage, Thousand  
609 Oaks CA. <https://socialsciences.mcmaster.ca/jfox/Books/Companion/>.

610 Fu X, Wang J, Sainju U M, Zhao F, Liu W. 2019. Soil microbial community and carbon and nitrogen  
611 fractions responses to mulching under winter wheat. *Appl Soil Ecol.* 139: 64-68.

612 Fuerst J A, Sagulenko E. 2011. Beyond the bacterium: planctomycetes challenge our concepts of  
613 microbial structure and function. *Nature Rev Microbiol.* 9: 13-18.

614 Gay-des-Combes J M, Robroek B J M, Hervé D, Guillaume T, Pistocchi C, Mills R T E, Buttler A.  
615 2017a. Slash-and-burn agriculture and tropical cyclone activity in Madagascar: Implication for  
616 soil fertility dynamics and corn performance. *Agric, Ecosyst Environ.* 239: 207-218.

617 Gay-des-Combes J M, Sanz Carrillo C, Jozef B, Robroek M, Jassey V E J, Mills R T E, Arif M S,  
618 Falquet L, Frossard E, Buttler A. 2017b. Tropical soils degraded by slash-and-burn cultivation  
619 can be recultivated when amended with ashes and compost. *Ecol Evolut.* 7: 5378-5388.

620 Hedrich S, Schlömann M, Johnson D B. 2011. The iron-oxidizing proteobacteria. *Microbiology.* 157:  
621 1551-1564.

622 Horn M, Collingro A, Schmitz-Esser S, Beier C L, Purkhold U, Fartmann B, Brandt P, Nyakatura G  
623 J, Droege M, Frishman D, Rattei T, Mewes H-W, Wagner M. 2004. Illuminating the evolutionary  
624 history of Chlamydiae. *Reports Science*. 304, 728-731.

625 Hug L A, Castelle C J, Wrighton K C, Thomas B C, Sharon I, Frischkorn K R, Williams K H, Tringe  
626 S G, Banfield J F. 2013. Community genomic analyses constrain the distribution of metabolic  
627 traits across the *Chloroflexi* phylum and indicate roles in sediment carbon cycling. *Microbiome*.  
628 1: 1–17.

629 Ibrahim M M, Tong C, Hu K, Zhou B, Xing S, Mao Y. 2020. Biochar-fertilizer interaction modifies  
630 N-sorption, enzyme activities and microbial functional abundance regulating nitrogen retention  
631 in rhizosphere soil. *Sci Total Environ*. 739: 140065.

632 Jakovac C C, Peña-Claros M, Mesquita R C G, Bongers F, Kuyper T W. 2016. Swiddens under  
633 transition: Consequences of agricultural intensification in the Amazon. *Agric, Ecosyst Environ*.  
634 218: 116-125.

635 Jenkins J R, Viger M, Arnold E C, Harris Z M, Ventura M, Miglietta F, Girardin C, Edwards R J,  
636 Rumpel C, Fornasier F, Zavalloni C, Tonon G, Alberti G, Taylor G. 2017. Biochar alters the soil  
637 microbiome and soil function: results of next-generation amplicon sequencing across Europe.  
638 *GCB Bioen*. 9: 591-612.

639 Johnson W A, Alfaress S, Whitworth R J, McCornack B P. 2012. Crop residue and residue  
640 management effects on *Armadillidium vulgare* (Isopoda: Armadillidiidae) populations and  
641 soybean. *J Econ Entom*. 12: 1629-1639.

642 Juo A S R, Manu A. 1996. Chemical dynamics in slash-and-burn agriculture. *Agric, Ecosyst Environ*.  
643 58: 49-60.

644 Kabisa M, Ncheengamwa H. 2020. Innovations in sustainable charcoal production: Are charcoal  
645 associations the key to greening the value chain in Zambia? Technical report, November 2019.

646 Kleinman P J A, Pimentel D, Bryant R B. 1995. The ecological sustainability of slash and burn  
647 agriculture. *Agric, Ecosyst Environ*. 52: 235-249.

648 Kassambara A. 2020. rstatix: pipe-friendly framework for basic statistical tests. R package version  
649 0.4.0. <https://CRAN.R-project.org/package=rstatix>.

650 Klindworth A, Pruesse E, Schweer T, Peplies J, Quast C, Horn M, Glöckner F O. 2013. Evaluation  
651 of general 16S ribosomal RNA gene PCR primers for classical and next-generation sequencing-  
652 based diversity studies. *Nucleic Acids Res.* 41(1):e1.

653 Kottek M, Grieser J, Beck C, Rudolf B, Rubel F. 2006. World Map of the Köppen-Geiger climate  
654 classification updated. *Meteorologische Zeitschrift.* 15: 259-263.

655 Koyama A, Wallenstein M D, Simpson R T, Moore J C. 2014. Soil bacterial community composition  
656 altered by increased nutrient availability in Arctic tundra soils. *Front Microbiol.* 5: 1-16.

657 Kukla J, Whitfeld T, Cajthaml T, Baldrian P, Veselá-Šimáčková H, Novotný V, Frouz J. 2018. The  
658 effect of traditional slash-and-burn agriculture on soil organic matter, nutrient content, and  
659 microbiota in tropical ecosystems of Papua New Guinea. *Land Degr Devel.* 30: 166-177.

660 Lacap-Bugler D C, Lee K K, Archer S, Gillman L N, Lau M C Y, Leuzinger S, Lee C K, Maki T,  
661 McKay C P, Perrott J K, de los Rios-Murillo A, Warren-Rhodes K A, Hopkins D W, Pointing S  
662 B. 2017. Global diversity of desert hypolithic Cyanobacteria. *Front Microbiol.* 8: 1-17.

663 Lanc M D, Kavamura V N, Gouve R, Andreote D, Mendes R, de Melo I S. 2013. Water regime  
664 influences bulk soil and rhizosphere of *Cereus jamacaru* bacterial communities in the brazilian  
665 caatinga biome. *PLOS one.* 8: e73606.

666 Lauber C L, Strickland M S, Bradford M A, Fierer N. 2008. The influence of soil properties on the  
667 structure of bacterial and fungal communities across land-use types. *Soil Biol Biochem.* 40: 2407-  
668 2415.

669 Lavkulich L M, Wiens J H. 1970. Comparison of organic matter destruction by hydrogen peroxide  
670 and sodium hypochlorite and its effect on selected mineral constituents. *Soil Sci Soc Am Proceed.*  
671 34: 755-758.

672 Li W, Tian X, Sheng H, Ekawati D, Zhou Y, Zhang R. 2020. Response of bacterial compositions to  
673 soil biochemical properties under mulching-intensive management in a *Phyllostachys edulis*  
674 forest. *Appl Soil Ecol.* 150: 103436.

675 Liu M, Zhang W, Wang X, Wang F, Dong W, Hu C, Liu B, Sun R. 2020. Nitrogen leaching greatly  
676 impacts bacterial community and denitrifiers abundance in subsoil under long-term fertilization.  
677 *Agric, Ecosyst Environ.* 294: 106885.

678 Liu X, Cong J, Lu H, Xue Y, Wang X, Li D, Zhang Y. 2017. Community structure and elevational  
679 distribution pattern of soil Actinobacteria in alpine grasslands. *Acta Ecolog Sinica.* 37: 213-218.

680 Madhaiyan M, Poonguzhali S, Senthilkumar M, Pragatheswari D, Lee J S, Lee K C. 2015.  
681 *Arachidicoccus rhizosphaerae* gen. nov., sp. nov., a plant-growth-promoting bacterium in the  
682 family *Chitinophagaceae* isolated from rhizosphere soil. *Inter J Syst Evol Microb.* 65: 578-586.

683 Magoc T, Salzberg S L. 2011. FLASH: Fast length adjustment of short reads to improve genome  
684 assemblies. *Bioinfo.* 27: 2957-2963.

685 Maria R M, Yost R. 2006. A survey of soil fertility status of four agroecological zones of  
686 Mozambique. *Soil Sci.* 171: 902–914.

687 Meloun M, Sánka M, Němec P, Křítková S, Kupka K. 2005. The analysis of soil cores polluted with  
688 certain metals using the Box-Cox transformation. *Environ Pollut.* 137: 273–280.

689 Mendes L W, Tsai S M, Navarrete A A, de Hollander M, van Veen J A, Kuramae E E. 2015. Soil-  
690 Borne Microbiome: Linking Diversity to Function. *Microb Ecol.* 70: 255–265.

691 Mertz O, Leisz S J, Heinemann A, Rerkasem K, Dressler T, Dressler W, Pham V C, Vu K C, Schmidt-  
692 Vogt D, Colfer C J P, Epprecht M, Padoch C, Potter L. 2009a. Who Counts? Demography of  
693 Swidden Cultivators in Southeast Asia. *Human Ecol.* 37: 281-289.

694 Mesa V, Gallego J L R, González-Gil R, Lauga B, Sánchez J, Méndez-García C, Peláez A I. 2017.  
695 Bacterial, archaeal, and eukaryotic diversity across distinct microhabitats in an acid mine  
696 drainage. *Front Microb.* 8: 1-17.

697 Nannipieri P, Ascher J, Ceccherini M T, Landi L, Pietramellara G, Renella G. 2017. Microbial  
698 diversity and soil functions. *Europ J Soil Sci.* 68: 12-26.

699 Nannipieri P, Ascher-Jenull J, Ceccherini M T, Pietramellara G, Renella G, Schloter M 2020. Beyond  
700 microbial diversity for predicting soil functions: A mini review. *Pedosphere* 30: 5–17.

701 Nath T K, Jashimuddin M, Hasan Md K, Shahjahan Md, Pretty J. 2016. The sustainable  
702 intensification of agroforestry in shifting cultivation areas of Bangladesh. *Agrofor Syst.* 90: 405-  
703 416.

704 Navarrete A A, Soares T, Rossetto R, van Veen J A, Tsai S M, Kuramae E E. 2015. Verrucomicrobial  
705 community structure and abundance as indicators for changes in chemical factors linked to soil  
706 fertility. *Antonie van Leeuwenhoek* 108: 741-752.

707 Nelson D W, Sommers L E. 1996. Total carbon, organic carbon and organic matter. Sparks D L, Page  
708 A L, Helmke P A, Loeppert R H, Soltanpour P N, Tabatabai M A, Johnston C T, Sumner M E  
709 (Eds.), Part 3: Chemical Methods. SSSA, Wisconsin USA.

710 Nourou Sall S, Masse D, Badiane Ndour N Y, Chotte J L. 2006. Does cropping modify the  
711 decomposition function and the diversity of the soil microbial community of tropical fallow soil?.  
712 *Appl Soil Ecol.* 31: 211-219.

713 Olsen S R, Cole C V, Watanabe F S, Dean L A. 1954. Estimation of available phosphorus in soils by  
714 extraction with sodium bicarbonate. US Department of Agriculture, Washington (Circ. 939).

715 Osburn E D, McBride S G, Aylward F O, Badgley B D, Strahm B D, Knoepp J D, Barrett J E. 2019.  
716 Soil bacterial and fungal communities exhibit distinct long-term responses to disturbance in  
717 temperate forests. *Front Microbiol.* 10:2872. doi: 10.3389/fmicb.2019.02872

718 O'Brien S L, Gibbons S M, Owens S M, Hampton-Marcell J, Johnston E R, Jastrow J D, Gilbert J A,  
719 Meyer F, Antonopoulos D A. 2016. Spatial scale drives patterns in soil bacterial diversity. *Environ*  
720 *Microb.* 18: 2039-2051.

721 Prada Salcedo L D, Prieto C, Correa Franco M. 2014. Screening phosphate solubilizing actinobacteria  
722 isolated from the rhizosphere of wild plants from the Eastern Cordillera of the Colombian Andes.  
723 *African J Microb Res.* 8: 734-742.

724 Qu J H, Yuan H L, Yang J S, Li H F, Chen N. 2009. *Lacibacter cauensis* gen. nov., sp. nov., a novel  
725 member of the phylum *Bacteroidetes* isolated from sediment of a eutrophic lake. *Inter J Syst Evol*  
726 *Microb.* 59: 1153-1157.

727 RStudio Team 2020. RStudio: Integrated Development for R. RStudio, PBC, Boston, MA.  
728 <http://www.rstudio.com/>.

729 R Core Team 2013. R: A language and environment for statistical computing. R Foundation for  
730 Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0. <http://www.R-project.org/>.

731 Rafael R B A, Fernández-Marcos M L, Cocco S, Ruello M L, Fornasier F, Corti G. 2020. Increased  
732 phosphorus availability to corn resulting from the simultaneous applications of phosphate rock,  
733 calcareous rock, and biochar to an acid sandy soil. *Pedosphere.* 30: 719-733.

734 Revelle, W. 2021. psych: procedures for psychological, psychometric, and personality research.  
735 northwestern university, evanston, illinois. R package version 2.1.6, [https://CRAN.R-](https://CRAN.R-project.org/package=psych)  
736 [project.org/package=psych](https://CRAN.R-project.org/package=psych).

737 Saker R, Meklat A, Bouras N, Zitouni A, Mathieu F, Spröer C, Klenk H P, Sabaou N. 2015. Diversity  
738 and antagonistic properties of culturable halophilic actinobacteria in soils of two arid regions of  
739 septentrional Sahara: M'zab and Zibans. *Annual Microb.* 65: 2241-2253.

740 Saliou Sarr P, Sugiyama A, Boyogueno Begoude A D, Yazaki K, Araki S, Nawata E. 2019. Diversity  
741 and distribution of Arbuscular Mycorrhizal Fungi in cassava (*Manihot esculenta* Crantz)  
742 croplands in Cameroon as revealed by Illumina MiSeq. *Rhizosphere.* 10: 100-147.

743 Sangwan P, Chen X, Hugenholtz P, Janssen P H. 2004. *Chthoniobacter flavus* gen. nov., sp. nov., the  
744 first pure-culture representative of Subdivision Two, *Spartobacteria* classis nov., of the phylum  
745 Verrucomicrobia. *Appl Environ Microb.* 70: 5875-5881.

746 Saska P. 2008. Granivory in terrestrial isopods. *Ecol Entom.* 33: 742-747.

747 Schoeneberger P J, Wysocki D A, Benham E C, and Soil Survey Staff. 2012. Field book for describing  
748 and sampling soils Version 3.0. Natural Resources Conservation Service, National Soil Survey  
749 Center, Lincoln, NE.

750 Scott W D, Kleb, H.R., 1996. The Influence of prairie and forest vegetation on soil moisture and  
751 available nitrogen. *The Am Midland Natural*. 136: 222-231.

752 Siteo A. 2004. Miombo woodlands and HIV/AIDS interactions Mozambique country report forestry  
753 working paper. FAO, Rome.

754 Smith N R, Kishchuk B E, Mohn W W. 2008. Effects of wildfire and harvest disturbances on forest  
755 soil bacterial communities. *Appl Environ Microb*. 74: 216-224.

756 Sofo A, Ricciuti P, Fausto C, Mininni A N, Crecchio C, Scagliola M, Malerba A D, Xiloyannis C,  
757 Dichio B. 2019. The metabolic and genetic diversity of soil bacterial communities depends on  
758 the soil management system and C/N dynamics: The case of sustainable and conventional olive  
759 groves. *Appl Soil Ecol*. 137: 21–28.

760 Soil Survey Staff, 2014. Keys to Soil Taxonomy, 12<sup>th</sup> edition. United States Department in  
761 Agriculture Natural Resources Conservation Service, Washington, DC.

762 Soil Survey Staff, 2015. Illustrated guide to soil taxonomy, version 2. U.S. Department of Agriculture,  
763 Natural Resources Conservation Service, National Soil Survey Center, Lincoln, Nebraska.

764 Styger E, Rakotondramasy H M, Pfeffer M J, Fernandes E C M, Bates D M. 2007. Influence of slash-  
765 and-burn farming practices on fallow succession and land degradation in the rainforest region of  
766 Madagascar. *Agric Ecosyst Environ*. 119, 257-269.

767 Sul W J, Asuming-Brempong S, Wang Q, Turlousse D M, Penton C R, Deng Y, Rodrigues J L M,  
768 Adiku S G K, Jones J W, Zhou J, Cole J R, Tiedje J M. 2013. Tropical agricultural land  
769 management influences on soil microbial communities through its effect on soil organic carbon.  
770 *Soil Biol Biochem*. 65: 33-38.

771 Suzuki S, Noble A D, Ruaysoongnern S, Chinabut N. 2007. Improvement in water-holding capacity  
772 and structural stability of a sandy soil in Northeast Thailand. *Arid Land Res Manag*. 21: 37-49.



773 Tang H, Shi X, Wang X, Hao H, Zhang X-M, Zhang L-P. 2016. Environmental controls over  
774 Actinobacteria communities in ecological sensitive Yanshan Mountains zone. *Frontiers in*  
775 *Microb.* 7: 1-13.

776 Tarasevich Y I, Polyakova I G, Polyakov V E. 2002. Hydrophilicity-hydrophobicity characteristics  
777 of solid surfaces and the state of water near surfaces of a various nature. *Adsorp Sci Techn.* 20:  
778 927-935.

779 Thomaz E L. 2018. Dynamics of aggregate stability in slash-and-burn system: Relaxation time, decay,  
780 and resilience. *Soil Till Res.* 178: 50-54.

781 Thomaz E L, Antoneli V, Doerr S H. 2014. Effects of fire on the physicochemical properties of soil  
782 in a slash-and-burn agriculture. *Catena.* 122: 209-215.

783 Tytgat B, Verleyen E, Sweetlove M, D'hondt S, Clercx P, Van Ranst E, Peeters K, Roberts S,  
784 Namsaraev Z, Wilmotte A, Vyverman W, Willems A. 2016. Bacterial community composition  
785 in relation to bedrock type and macrobiota in soils from the Sør Rondane Mountains, East  
786 Antarctica. *FEMS Microb Ecol.* 92: 1-13.

787 Ulrich A, Becker R. 2006. Soil parent material is a key determinant of the bacterial community  
788 structure in arable soils. *FEMS Microb Ecol.* 56: 430-443.

789 Vipindas P V, Mujeeb R K M, Jabir T, Thasneem T R, Mohamed Hatha A A. 2020. Diversity of  
790 sediment bacterial communities in the South Eastern Arabian Sea. *Reg Studies Marine Sci.* 35:  
791 101153.

792 Vos P, Garrity G, Jones D, Krieg N R, Ludwig W, Rainey F A, Schleifer K-H, Whitman W B. 2011.  
793 Bergey's Manual of Systematic Bacteriology. Vol 3: The Firmicutes. Springer Science &  
794 Business Media.

795 Wagner M, Horn M. 2006. The Planctomycetes, Verrucomicrobia, Chlamydiae and sister phyla  
796 comprise a superphylum with biotechnological and medical relevance. *Current Opinion in*  
797 *Biotechn.* 14: 241-249. Wang C, Liu D, Bai E. 2018. Decreasing soil microbial diversity is

798 associated with decreasing microbial biomass under nitrogen addition. *Soil Biol Biochem.* 120:  
799 126-133.

800 Wang C, Liu D, Bai E. 2018. Decreasing soil microbial diversity is associated with decreasing  
801 microbial biomass under nitrogen addition. *Soil Biol Biochem.* 120:126–133.  
802 <https://doi.org/10.1016/j.soilbio.2018.02.003>.

803 Wang Q, Garrity G M, Tiedje J M, Cole J R. 2007. Naive Bayesian classifier for rapid assignment of  
804 rRNA sequences into the new bacterial taxonomy. *Appl Environ Microbiol.* 73:5261–5267.  
805 <https://doi.org/10.1128/AEM.00062-07>.

806 Wijnhoud J D. 1997. Solos e outros recursos naturais da Estação Agrária de Sussundenga, vol. 1:  
807 Relatório. Série da Terra e Água do Instituto Nacional De Investigação Agrónomica,  
808 comunicação n°93.

809 Wolińska A, Kuźniar A, Zielenkiewicz U, Izak D, Szafranek-Nakonieczna A, Banach A, Błaszczuk  
810 M. 2017. Bacteroidetes as a sensitive biological indicator of agricultural soil usage revealed by a  
811 culture-independent approach. *Appl Soil Ecol.* 119: 128-137.

812 Wood S A, Gilbert J A, Leff J W, Fierer N, D'Angelo H, Bateman C, Gedallovich S M, Gillikin C M,  
813 Gradoville M R, Mansor P, Massmann A, Yang N, Turner B L, Brearley F Q, McGuire K L. 2017.  
814 Consequences of tropical forest conversion to oil palm on soil bacterial community and network  
815 structure. *Soil Biol Biochem.* 112: 258-268.

816 Xu C, Yang Z, Qian W, Chen S, Liu X, Lin W, Xiong D, Jiang M, Chang C-T, Huang Jr C, Yang Y.  
817 2019. Runoff and soil erosion responses to rainfall and vegetation cover under various  
818 afforestation management regimes in subtropical montane forest. *Land Degrad Develop.* 30:  
819 1711-1724.

820 Yang S, Zheng Q, Yang Y, Yuan M, Ma X, Chiariello N R, Docherty K M, Field C B, Gutknecht, J  
821 L M, Hungate B A, Niboyet A, Le Roux X, Zhou J. 2020. Fire affects the taxonomic and  
822 functional composition of soil microbial communities, with cascading effects on grassland  
823 ecosystem functioning. *Global Change Biol.* 26: 431-442.

824 Yin C, Jones K L, Peterson D E, Garrett K A, Hulbert S H, Paulitz T C. 2010. Members of soil  
825 bacterial communities sensitive to tillage and crop rotation. *Soil Biol Biochem.* 42: 2111-2118.

826 Zhao F Z, Fan X D, Ren C J, Zhang L, Han X H, Yang G H, Wang J, Doughty R. 2018. Changes of  
827 the organic carbon content and stability of soil aggregates affected by soil bacterial community  
828 after afforestation. *Catena.* 171: 622-631.

829 Zhou J, Guan D, Zhou B, Zhao B, Ma M, Qin J, Jiang X, Chen S, Cao F, Shen D, Li J. 2015. Influence  
830 of 34-years of fertilization on bacterial communities in an intensively cultivated black soil in  
831 northeast China. *Soil Biol Biochem.* 90: 42-51.

832

833