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Detection of phenicol-oxazolidinone resistance gene optrA in *Aerococcus viridans* from bovine faeces, Italy

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**First detection of phenicol-oxazolidinone resistance gene  
*optrA* in *Aerococcus viridans* from bovine faeces, Italy**

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Manuscripts

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Letter

2   **First detection of phenicol-oxazolidinone resistance gene *optrA* in**  
3   ***Aerococcus viridans* from bovine faeces, Italy**

4

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26 Sir,

27

28 antibiotic resistance genes (ARGs) are now ubiquitous also in non-clinical environments.  
29 Specifically, livestock farms are considered major reservoirs of ARGs and pose a risk for  
30 human health since ARGs can be spread to human bacterial pathogens. Although many  
31 antimicrobials administered to animals are used exclusively in veterinary applications,  
32 most of them can drive co-selection of genes for the resistance to drugs critical in human  
33 medicine.

34 Oxazolidinones, including linezolid and tedizolid, are effective antimicrobial agents for  
35 the treatment of severe human infections due to MDR Gram-positive bacteria and they  
36 have never been approved for animal use. Nevertheless, acquired linezolid-resistance has  
37 arisen in bacteria of animal origin, as well as in human isolates, due to extensive use of  
38 phenicols in veterinary medicine that provide the selective pressure for a rapid  
39 dissemination of the *cfr*,<sup>1</sup> *optrA*<sup>2</sup> and *poxtA*<sup>3</sup> linezolid resistance genes on animal farms.<sup>4</sup>  
40 Analysis of the prevalence and distribution of linezolid-resistance genes in animal  
41 settings is important for evaluating the risk to public health and for development of  
42 control measures.

43 During a routine surveillance aimed at the detection of linezolid resistance genes in  
44 enterococci of animal origin, pooled faecal samples were collected from cattle farms  
45 where florfenicol was used previously. The samples were inoculated in buffered peptone  
46 water supplemented with florfenicol (10 mg/L) and incubated at 44°C for 48 hours.

47 Then, 0.1 mL was inoculated in Slanetz Bartley agar plates supplemented with  
48 florfenicol (10 mg/L) and the enterococci grown on this medium were tested by PCR for  
49 the presence of *cfr*, *optrA* and *poxtA* genes.<sup>5</sup> Isolates carrying at least one linezolid  
50 resistance gene were identified by MALDI-TOF (Vitek-MS, bioMérieux). Interestingly,  
51 one *optrA*-positive isolate (called 1417-4A) belonged to the *Aerococcus viridans* species.

52 Susceptibility testing, carried out by reference broth microdilution ([www.clsi.org](http://www.clsi.org)) and  
53 interpreted according to the EUCAST clinical breakpoints (version 10.0,

54 [www.eucast.org](http://www.eucast.org)) revealed that MICs of linezolid, tedizolid, and florfenicol for *A.*  
55 *viridans* 1417-4A were 1 mg/L, 1 mg/L, and 16 mg/L, respectively. In filter mating  
56 experiments, using florfenicol (10 mg/L) for selection,<sup>6</sup> *optrA* gene was successfully  
57 transferred from *A. viridans* 1417-4A to *E. faecium* 64/3 recipient (transfer frequency,  
58  $1.7 \times 10^{-5}$  per recipient). Two randomly selected transconjugants showed MICs of  
59 tedizolid and florfenicol identical to those for the donor, instead a linezolid MIC of 4  
60 mg/L was recorded for both transconjugants. It is notable that a 4-fold increase in the  
61 linezolid MICs was detected in transconjugants compared to that of *E. faecium* 64/3

62 recipient (1 mg/L) suggesting that *optrA* of *A. viridans* could confer reduced linezolid  
63 susceptibility in enterococcal background.

64 In S1-PFGE and hybridization assays,<sup>6</sup> an *optrA*-specific probe hybridized with a ca. 30-  
65 kb plasmid both in donor and transconjugants.

66 In order to characterize the *optrA* genetic element, whole-genome sequence (WGS) of *A.*  
67 *viridans* 1417-4A was carried out by Illumina MiSeq platform (MicrobesNG,  
68 Birmingham, UK) using a 2 x 250 paired end approach. De novo assembly of WGS data  
69 was performed using the SPAdes software (<http://bioinf.spbau.ru/spades>).

70 Bioinformatics analysis revealed that *optrA* gene was located on a new 37,845-bp  
71 plasmid, named pAv-optrA (G+C content, 32.0%) (accession no. MW364930). The map  
72 of the plasmid is shown in Figure 1 and the major characteristics of the ORFs are  
73 detailed in Table S1. Four relevant areas were detectable in pAv-*optrA*: (i) an antibiotic  
74 resistance region containing *optrA* (responsible for phenicols and oxazolidinone  
75 resistance) and *erm(B)* (responsible for macrolide, lincosamide and streptogramin B  
76 resistance) genes. The *optrA* gene was 98.0% identical to the DNA reference sequence  
77 (GenBank accession no. KP399637.1). Moreover, twenty-two amino acid changes were  
78 detected in the protein sequence compared to that of OptrA<sub>E349</sub> (96% identity, 98%  
79 similarity)(Figure S1);<sup>2</sup> (ii) a conjugation region spanning from *orf10* to *orf23*; (iii) a  
80 segment harbouring *repA* (*orf28*) and *parA* (*orf30*) genes responsible for plasmid  
81 replication and partitioning, respectively. The *repA* gene was not typeable by plasmid  
82 finder (<https://cge.cbs.dtu.dk/services/PlasmidFinder/>); (iv) a complete type 1  
83 restriction-modification cassette (*orf35*, *orf1* and *orf2*) showing 96.0% DNA sequence  
84 identity with the corresponding region of *poxtA*-carrying pC25-1 and pC27-2 plasmids.<sup>7</sup>

85 A post-segregational killing mechanism might be responsible for maintaining of pAV-  
86 optrA in *A. viridans* 1417-4A host.<sup>8</sup>

87 WGS analysis also ruled out the presence of *cfr*, *cfr*-like and *poxtA* genes. No mutations  
88 were detected in the genes encoding the 23S rRNA and ribosomal proteins.

89 Aerococci are widespread in the environment and frequently isolated from foods and  
90 animals and now are increasingly recognized as human pathogens.<sup>9</sup> To the best of our  
91 knowledge, this is the first report of the *optrA* gene in *Aerococcus* species, carried by a  
92 conjugative plasmid. *A. viridans* caused different human infection such as urinary tract  
93 infections, osteomyelitis, septic arthritis, septicaemia and endocarditis, therefore the  
94 presence of linezolid resistance *optrA* gene in this species and its ability to transfer to  
95 human pathogen *E. faecium*, are cause for concern.

96

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99    and by Progetto Strategico di Ateneo 2017 – Polytechnic University of Marche,  
100   “In the hunt of new antibiotics: active compounds from both chemical synthesis  
101   and natural sources”.

102

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104    **Transparency declarations**

105    None to declare

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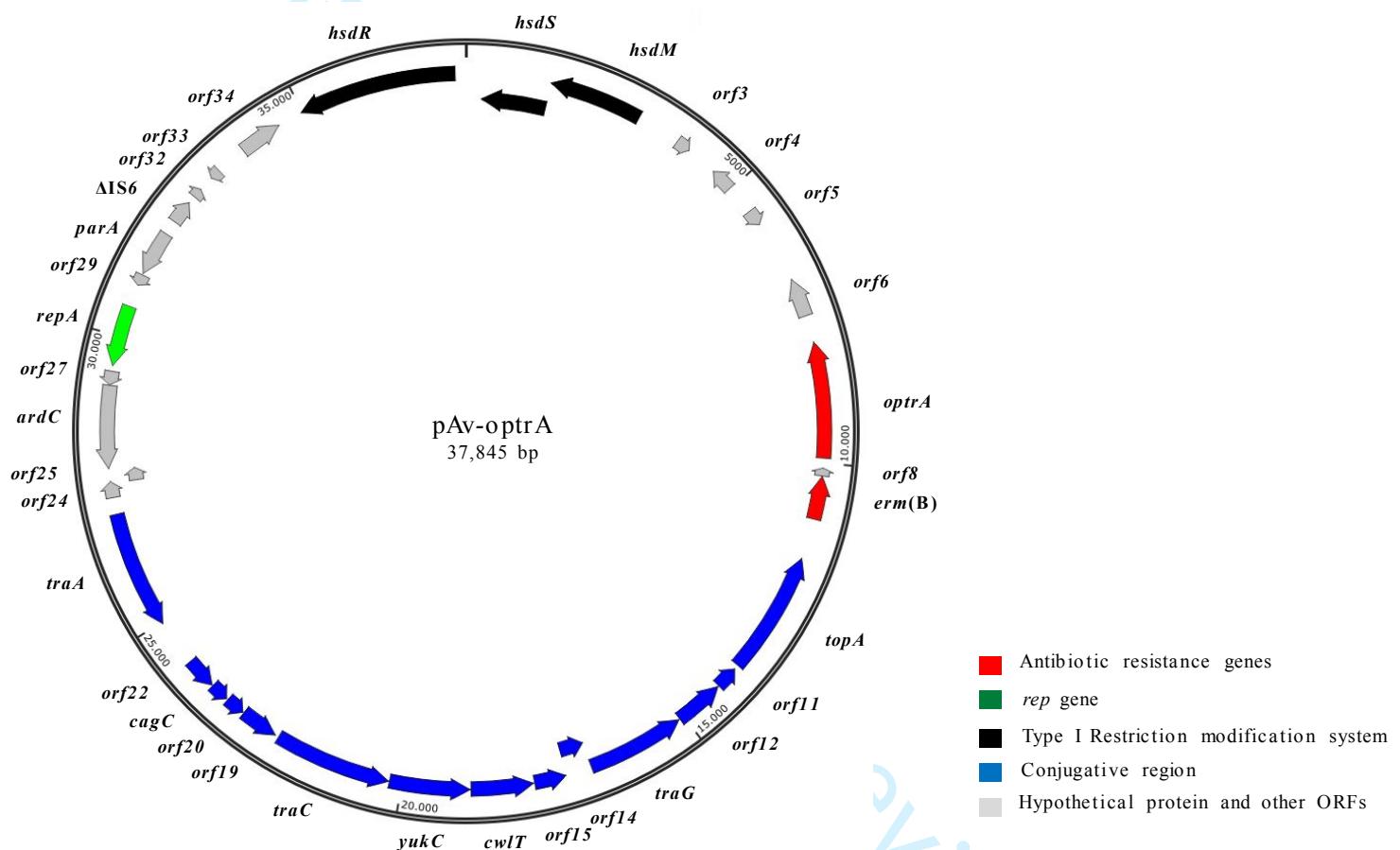
137 **Figure**

138

139 **Figure 1.** Schematic representation of the *optrA*-carrying pAv-*optrA* plasmid (16,500 bp)  
140 from *A. viridans* 1417-4A (accession no. MW364930).

141 Arrows indicate the position and direction of transcription of different genes.

142



143 **Supplementary materials**

144

145 **Table S1.** Amino acid sequence identities/similarities of putative proteins encoded by the pAv-optrA (GenBank accession no. MW364930).

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	ORF	Start (bp)	Stop (bp)	Size (amino acids)	Predicted function
153	<i>orf1</i>	1,449	283	388	Restriction endonuclease S subunit
154	<i>orf2</i>	3,038	1,449	528	Type I restriction-modification system DNA methylase subunit
155	<i>orf3</i>	3,797	4,039	80	DNA-invertase hin
156	<i>orf4</i>	4,979	4,581	132	
157	<i>orf5</i>	5,489	5,761	90	
158	<i>orf6</i>	7,486	6,833	217	
159	<i>orf7</i>	9,915	7,948	655	ABC-F type ribosomal protection protein Optra
160	<i>orf8</i>	10,213	10,082	43	
161	<i>orf9</i>	10,955	10,218	245	Ribosomal RNA adenine methylase
162	<i>orf10</i>	13,753	11,648	701	DNA topoisomerase IA
163	<i>orf11</i>	14,223	13,831	130	
164	<i>orf12</i>	15,082	14,237	281	
165	<i>orf13</i>	16,770	15,097	557	Type IV secretory system Conjugative DNA transfer
166	<i>orf14</i>	17,207	16,770	145	
167	<i>orf15</i>	17,764	17,222	180	
168	<i>orf16</i>	18,835	17,783	350	CwlT-like N-terminal lysozyme domain
169	<i>orf17</i>	20,214	18,850	454	Protein secretion system
170	<i>orf18</i>	22,223	20,229	664	Type-IV secretion system protein TraC
171	<i>orf19</i>	22,941	22,291	216	
172	<i>orf20</i>	23,289	22,957	110	

BLASTP analysis<sup>a</sup>

Most significant database match

Accession no.	% Amino acid identity (% amino acid similarity)
WP_111856479.1	65 (72)
WP_111856478.1	98 (99)
SPT61043.1	100 (100)
WP_160516845.1	93 (95)
HCT97980.1	43 (61)
HAP3453445.1	100 (100)
HAP3438646.1	99 (99)
WP_000085855.1	100 (100)
WP_001038790.1	100 (100)
WP_104846964.1	75 (85)
WP_068710574.1	43 (64)
WP_192982111.1	51 (72)
WP_138472019.1	86 (92)
WP_091636415.1	54 (73)
WP_157850759.1	46 (63)
WP_053083829.1	62 (77)
EGP5098947.1	43 (68)
WP_053083830.1	77 (87)
WP_138472028.1	64 (81)
WP_138472031.1	54 (77)

176	<i>orf21</i>	23,634	23,314	106	Cag pathogenicity island, type IV secretory system	Hypothetical protein [ <i>Carnobacterium</i> sp. 1290_CSPC]	WP_048729905.1	63 (78)
177						Hypothetical protein [ <i>Carnobacterium</i> sp. 1290_CSPC]	WP_048729908.1	40 (66)
178	<i>orf22</i>	24,188	23,646	180		MobA/MobL family protein	WP_053083831.1	52 (68)
179	<i>orf23</i>	26,975	24,972	667		Hypothetical protein [ <i>Carnobacterium</i> sp. 1290_CSPC]	WP_048729913.1	42 (71)
180	<i>orf24</i>	27,266	27,532	88		Hypothetical protein [ <i>Carnobacterium</i> sp. 1290_CSPC]	WP_048729916.1	62 (80)
181	<i>orf25</i>	27,507	27,728	73		ImmA/IrrE family metallo-endopeptidase	WP_048729918.1	76 (83)
182	<i>orf26</i>	29,159	27,756	467	Antirestriction protein ArdC	[ <i>Carnobacterium</i> sp. 1290_CSPC]		
183						Hypothetical protein [ <i>Carnobacterium</i> sp. 1290_CSPC]	WP_048729921.1	97 (97)
184	<i>orf27</i>	29,398	29,165	77		Replication initiator protein A (RepA)	WP_048729642.1	76 (85)
185	<i>orf28</i>	30,526	29,492	344		Hypothetical protein [ <i>Carnobacterium</i> sp. 1290_CSPC]	WP_048729641.1	92 (96)
186	<i>orf29</i>	31,097	30,912	61		ParA family protein [ <i>Carnobacterium</i> sp. 1290_CSPC]	WP_048729640.1	89 (93)
187	<i>orf30</i>	31,901	31,128	257	Plasmid partitioning protein	IS6 family transposase, partial [ <i>E. faecium</i> ]	WP_147239629.1	94 (95)
188	<i>orf31</i>	32,141	32,548	135	DDE transposase	Hypothetical protein [Bacilli]	WP_164507104.1	100 (100)
189	<i>orf32</i>	32,677	32,847	56		Hypothetical protein FOB80_07155 [ <i>A. viridans</i> ]	QGS37267.1	100 (100)
190	<i>orf33</i>	33,279	33,106	57		Class I SAM-dependent methyltransferase	WP_149358818.1	99 (100)
191	<i>orf34</i>	33,808	34,545	245	Methyltransferase domain	[ <i>Carnobacterium</i> sp. PL12RED10]		
192						Type-1 restriction enzyme R protein [ <i>A. viridans</i> ]	SPT61052.1	99 (99)
193	<i>orf35</i>	37,663	34,976	895	DEAD-like helicases superfamily			
194								

195 <sup>a</sup>For each ORF, only the most significant identity detected is listed.

196      **Figure S1.** Amino acid sequence alignment of the OptrA proteins of *A. viridans* 1417-4A (**Av**) and *E.*  
 197      *faecalis* E349 (**Ef**). Mismatches are highlighted in yellow.  
 198  
 199

200 <b>Av</b>	1	LSEATFAIASTYVKEDMKMQYKIINGAVYYDGNMVLENIGIEINDNEKIAIVGRNGCGKT	60
201		+S+ATFAIAST KEDMKMQYKIINGAVYYDGNMVLENIGIEINDNEKIAIVGRNGCGKT	
202 <b>Ef</b>	1	MSKATFAIASTNAKEDMKMQYKIINGAVYYDGNMVLENIGIEINDNEKIAIVGRNGCGKT	60
203			
204      Query	61	TLLKAIIGEIELEEGTGESEFQVIKTGNPYISYLRQMPFEDESIISMVDEVRTVFKTLIDM	120
205		TLLKAIIGEIELEEGTGESEFQVIKTGNPYISYLRQMPFEDESIISMVDEVRTVFKTLIDM	
206      Sbjct	61	TLLKAIIGEIELEEGTGESEFQVIKTGNPYISYLRQMPFEDESIISMVDEVRTVFKTLIDM	120
207			
208      Query	121	EKKMKQLIDKMENQCDDKIINEYSDIERYMALGLTYQKEYETMIRSMGFTEADDKKPI	180
209		E KMKQLIDKMENQ DDKIINEYSDI ERYMALGLTYQKEYETMIRSMGFTEAD KKPI	
210      Sbjct	121	ENKMKQLIDKMENQYDDKIINEYSDISERYMALGLTYQKEYETMIRSMGFTEADYKKPI	180
211			
212      Query	181	SEFSGGQRTKIAFIKILLTKPDILLDEPTNHLDIETIQWLESYLSYKSTLVIISHDRM	240
213		SEFSGGQRTKIAFIKILLTKPDILLDEPTNHLDIETIQWLESYLSYKSTLVIISHDRM	
214      Sbjct	181	SEFSGGQRTKIAFIKILLTKPDILLDEPTNHLDIETIQWLESYLSYKSTLVIISHDRM	240
215			
216      Query	241	FLNRIVDKVYEIEWGETKCYGNYSafeEQKRENHIKQQKDYLQQIEIERITRLIERFR	300
217		FLNRIVDKVYEIEWGETKCYGNYSafeEQKRENHIKQQKDYLQQIEIERITRLIERFR	
218      Sbjct	241	FLNRIVDKVYEIEWGETKCYGNYSafeEQKRENHIKQQKDYLQQIEIERITRLIERFR	300
219			
220      Query	301	YKPTKAKMVQSKIKLLQRMQILNAPDQYDTKTYMSKFQPRISSSRQVLSSVSELVIGYDTP	360
221		YKPTKAKMVQSKIKLLQRMQILNAPDQYDTKTYMSKFQPRISSSRQVLSSVSELVIGYDTP	
222      Sbjct	301	YKPTKAKMVQSKIKLLQRMQILNAPDQYDTKTYMSKFQPRISSSRQVLSSASELVIGYDTP	360
223			
224      Query	361	LAKVNFNLERGQKLGIVGSNGIGKSTLLKTLMDGASALSGDFKFGYNVEISYFDQQLAQI	420
225		LAKVNFNLERGQKLGIVGSNGIGKSTLLKTLG +ALSGDFKFGYNVEISYFDQQLAQI	
226      Sbjct	361	LAKVNFNLERGQKLGIVGSNGIGKSTLLKTLMGVAALSGDFKFGYNVEISYFDQQLAQI	420
227			
228      Query	421	SGDDTLFEIFQSEYPELNDTEVRTALGSFQFSGDDVFRPVSSLGGKEVRLTLCKLLYKR	480
229		SGDDTLFEIFQSEYPELNDTEVRTALGSFQFSGDDVFRPVSSLGGKEVRLTLCKLLYKR	
230      Sbjct	421	SGDDTLFEIFQSEYPELNDTEVRTALGSFQFSGDDVFRPVSSLGGKEVRLTLCKLLYKR	480
231			
232      Query	481	TNVLILDEPTNHMDIIGKENLENILCSYKGTIIIFVSHDRFTNKIADRLLVFDKGVEFV	540
233		TNVLILDEPTNHMDIIGKENLENILCSY+GTIIIFVSHDRFTNKIADRLLVFDKGVEFV	
234      Sbjct	481	TNVLILDEPTNHMDIIGKENLENILCSYQGTIIIFVSHDRFTNKIADRLLVFDKGVEFV	540
235			
236      Query	541	EESTYGEYEKKRINSEKPFNYINVEKKVEKNNTVKGDRNSIEKEVKKEKRIEKLEVLI	600
237		+STYGEYEKKR+NSEKPFN I VE+KVEKNNTVKGDRNSIEKEVKKEKRIEKLEVLI	
238      Sbjct	541	QESTYGEYEKKRMNSEKPFNNIKVEQKVEKNNTVKGDRNSIEKEVKKEKRIEKLEVLI	600
239			
240      Query	601	YDEELERLNKIISQPNNSSDYIVLTELQKSIDEVKRCQGIYFNEWEQLMGELEV	655
241		YDEELERLNKIIS+PNNSSDYIVLTE+QKSID+VKRCQGYFNEWEQLM ELEV	
242      Sbjct	601	YDEELERLNKIISEPNNSSDYIVLTIQKSIDDVKRCQGNYFNEWEQLMRELEV	655