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## Coffee By-products Derived Resources. A Review

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### **ABSTRACT**

Coffee is the most common beverage and there are two main different methods to process coffee cherries. Coffee husk is the main by-product of coffee processing by dry method and is available in large quantities throughout the years, but its main application has been limited to animal feeding or energy production. Most of the coffee husk is disposed in landfills or arable land, usually with no care of its fate and changes to the source of pollution, especially in developing countries. ~~Food industry by-product is one of the challenges~~ Coffee husk can have several re-uses, ~~and-but~~ it is important to have environment-friendly methods to change ~~the coffee husk~~ it into usable material or material to be recycled in nature because of its ~~relatively high~~ important content of organic matter, chemical nutrients, and secondary compounds. The aim of this review is to recollect the amounts and uses of the coffee industry by-products, giving emphasis to ~~the-its~~ transformation into compost because of their large content of nutrients and the need to introduce high valuable organics into the soil.

**Key Words:** Coffee-Husk, Component, Detoxification, Application, Waste-Management.

### **1. INTRODUCTION**

24 Demand for coffee in the last 150 years was more than in the past not only because of the  
25 increased population and urban development but also because coffee has become one of the most  
26 consumed beverages in the world [1]. Nowadays coffee trade is economically at the second place  
27 of the world rank after petroleum [2]. According to the International Coffee Organization (ICO),  
28 in the 2016/2017 crop year the world coffee production was  $\approx$ 152 million of 60-kilograms bags  
29 (for a total of  $\approx$ 9 million tons), for an economic value of  $\approx$ 90 billion dollars [3]. The world  
30 population of  $\approx$ 7.6 billion in 2017 is expected to reach 8.6 billion in 2030, and 9.8 billion in  
31 2050, with one third of the population concentrate in urban areas [4]. Because of this, also  
32 production and consumption of coffee are expected to increase concerning the actual levels.

33 Even though the first plantation of coffee was done in Yemen by Arab people in 13<sup>th</sup> century  
34 with seeds transferred from Ethiopia [5], nowadays Brazil, Vietnam, Indonesia, Colombia,  
35 Ethiopia, India, and Mexico are the major producers of coffee, with Brazil producing half of the  
36 world production [6]. The coffee plant belongs to the *Rubiaceae* family and, among the  
37 numerous species present in nature, currently only *Coffea arabica* L. (known as Arabica) and  
38 *Coffea canephora* L. (known as Robusta) have an important economic value [3]. Coffee  
39 processing industry produces huge amount of by-products since from 30 to 50% of coffee fruit  
40 weight is waste [6]. Due to the high amount of coffee seeds production, several re-using  
41 solutions have been proposed, but a win-win solution to manage the considerable amount of  
42 coffee husk is needed [7].

43 Because of this, the aims of this review were to report of 1) different processing methods of  
44 coffee cherries, introduce the main by-products of coffee processing, and emphasize on the  
45 needed detoxification of coffee husk obtained by dry method; and 2) the several usages of coffee

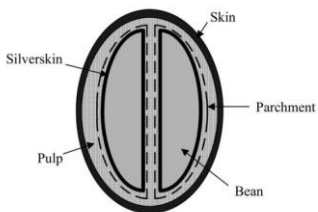
46 husk in industry and agriculture. All the themes referring to processes and uses were discussed  
47 also by synthesizing advantages and disadvantages reported in the literature.

48

## 49 **2. COFFEE SEED ANATOMY**

50 The economic lifespan of coffee tree is maximum of 30 years. The shrub is perennial and can  
51 reach a height of 10 meters. The first flowers are produced when plants are 3-4 years old, and are  
52 creamy white and sweetly scented, appearing in clusters in the axis of the leaves. Two fertilized  
53 ovules of coffee flower ovary start to grow up two months after fertilization. Adequate water  
54 supply is important to break the dormancy in the third month. The ovary size increases, and the  
55 embryotic sac grows and fills with endosperm. Till the end of the fifth month after fertilization,  
56 weight and volume of fruits increase significantly. Between sixth and eighth months after  
57 fertilization, the fruit reach to maturity, represented by an oval drupe of 18 mm in length and 10-  
58 15 mm in diameter. The ripe fruit has bright red or yellow color, and it is also called “cherry”  
59 [8][9][10].

60 The coffee fruit has four different layers protecting the seed that must be removed to collect the  
61 two beans forming the seed. The outer layer is the skin (epicarp or exocarp), with waxy  
62 substance and red color. The second layer is the pulp (mesocarp), that is a slim layer of  
63 pectinaceous materials. The third layer is the parchment (endocarp), with polysaccharide  
64 covering. The last layer, sticking to the seed, is named silverskin or chaff [11].



65

66 Figure1: Schematic picture of coffee fruit from [11].

67

### 68 **3. COFFEE PROCESSING METHODS AND BY-PRODUCTS**

69 To keep the quality of the seeds and preserve them from pathogens, they must be extracted from  
70 the four layers forming the other part of the fruit. The industrial process of coffee seeds  
71 preparation can be made following two main methods: dry and wet methods. The dry method is  
72 the traditional one, but it is also the simplest and environment-friendly since it produces less  
73 amount of solid and liquid by-product. Following this method, after having selected and cleaned  
74 the cherries, these are sun-dried with frequent turning to obtain a relatively homogenous drying.  
75 Thus, the outer layers of the cherries are removed by a hulling machine, and the coffee beans are  
76 roasted and bagged. With the wet method (or washed method) more equipment and water are  
77 needed in comparison to the dry method. With this method, the quality of the coffee beans is  
78 higher than with the dry method because the bean components are better preserved, and the  
79 number of defective seeds is less. Following this method, after the cherries are sorted and  
80 cleaned, the pulp is separated by a squeezing machine and the seeds are roasted [3]. Every ton of  
81 fresh coffee cherry produces 0.12-0.18 tons of coffee husk with the dry method and 0.5 tons of  
82 coffee pulp with the wet method [12]. Even though the quality of the obtained coffee seeds  
83 depends on the processing, the seeds produced with both methods have their own market since  
84 the beverage obtained with seeds submitted to the dry process is less acidic than those obtained  
85 with the wet method.

86 Each step of the coffee processing from coffee fruit to a cup of coffee, including separation of  
87 coffee seeds, roasting, packing, and making a drink, produces by-products.

88

89 **3.1. Spent coffee ground.** This by-product is the result of coffee brewing from coffee making  
90 such as homemade coffee and coffee machines or indirect way like instant coffee and beverage  
91 factories. It has a dark brown color, coarse texture, and high moisture [11]. The content of lipids  
92 in the fresh spent coffee ground is around 2% on a weight basis, with palmitic and linoleic acids  
93 covering 35% of the total extractable oil [13]. This by-product is also rich of vitamin E since, in  
94 classic espresso coffee and in coffee machines coffee, only 1 and 5%, respectively, of this  
95 vitamin is extracted. Therefore, coffee ground cakes can be used as a source of liposoluble  
96 antioxidant vitamins [14].

97

98 **3.2. Defective and premature coffee beans.** Both coffee harvesting and roasting process  
99 produce two types of by-products, respectively: immature and defective beans. These beans must  
100 be removed from the mass of the valuable beans since they might decrease the quality of the  
101 final products [15]. In fact, beans from defective cherries have higher amounts of free amino-  
102 acids and phenols and contain fewer sugars than normal beans because they did not reach proper  
103 maturity [16]. As an alternative use for these low-grade coffee beans, Alves *et al.* [11] suggested  
104 to use them for the extraction of chlorogenic acid or caffeine, for their potential applications in  
105 the food and pharmaceutical sectors.

106

107 **3.3. Silverskin.** Coffee silverskin is a so thin layer sticking to the coffee seeds that detaches only  
108 during coffee roasting [17]. Coffee silverskin has antioxidant activity, because of the presence of  
109 melanoidins [18], prebiotic activity [19], and contains dietary fiber [20]. These valuable  
110 components have encouraged studies on the production of body weight control beverages, diet  
111 bread, biscuits [17][21], and cosmetic products [22].

112  
113 **3.4. Coffee pulp.** Coffee pulp represents  $\approx 35\%$  of the coffee fruit [11] and is a by-product of the  
114 wet method in coffee hulling process. In the coffee pulp, the content of phenolic acids is slightly  
115 higher than in coffee husk, 1.5% vs. 1.2%. Among the phenolic acids comprising the coffee pulp,  
116 flavan-3-ols, hydroxycinnamic acids, flavonols, and anthocyanidins are the most represented  
117 [23].

118  
119 **3.5. Coffee husk.** Coffee husk is the main by-product of the dry method and is formed by all the  
120 layers at once, including dried skin, pulp, mucilage, and the parchment [24]. When the coffee  
121 cherry is dried,  $\approx 12\text{-}18\%$  of the dried fruit weight is coffee husk [5]. In general, amount of  
122 components and indexes of coffee husk vary according to the coffee species, the geographical  
123 origin of the cherries, and the chosen method of processing [25], which explains the differences  
124 in the composition reported by many authors [26][2][27][11][6][15]. For example, in Table 1  
125 we reviewed, compared and synthesized composition and physicochemical properties of coffee  
126 husk. However, taking into consideration the values reported in Table 1, Alves *et al.* [11]  
127 reported completely different amounts of lignocellulosic polymers, with 24.5% cellulose, 29.7%  
128 hemicellulose, and 23.7% lignin. It is desirable that in future studies coffee husks be classified  
129 according to their properties.

130

Table 1: Main composition and physicochemical properties of coffee husk.

	Value	References
<i>Organic component (g kg<sup>-1</sup>)</i>		
Carbohydrates	580-850	[27][6][11]
Cellulose	430	[2][27]

Hemicellulose	70	[2][27]
Lipids	5-30	[27][6][11]
Total fiber	240	[2]
Ash	25-62	[28][11]
Protein	80-110	[27][6][11]
Caffeine	10	[2][6][11]
Tannins	50	[2][6][11]
Chlorogenic acid	25	[2]
Pectic substance	16	[2]
Lignins	90	[2]
<i>Sugar content (g kg<sup>-1</sup>)</i>		
Reducing sugar	120	[29]
Total sugar	140	[29]
Sucrose	20	[29]
<i>Physicochemical parameters</i>		
pH (1:10)	5.35-6.63	[30][31]
EC (dS m <sup>-1</sup> )	2.24-3.1	[30][31]
Organic carbon (g kg <sup>-1</sup> )	545	[30]
Organic matter (g kg <sup>-1</sup> )	815	[31]
C/N ratio	29.8-40	[30][31]

131

132

133 **3.5.1 Macro- and micro-nutrients.** Coffee husk is rich of macro and micro-nutrients, with  
 134 considerable amount of N (1720-1830 mg kg<sup>-1</sup>), P (80 mg kg<sup>-1</sup>), K (20 600 mg kg<sup>-1</sup>), and others  
 135 (Table 2). Positively enough, it contains small amounts of Na.

136

Table 2: Elemental content of coffee husk. Values are expressed on a dry matter basis.

Element (mg kg <sup>-1</sup> )	Coffee Husk	References
Total content of inorganic elements	5000-30 000	[32]
N	1720-1830	[32][30]
P	80	[32]
K	20 600	[32]
Ca	2210	[32]
Mg	790	[32]



Fe	260	[32]
Cu	20	[32]
Mn	60	[32]
Zn	10	[32]
B	91.4	[33]
S	1100	[33]
Se	0.19	[32]
Na	40	[32]

137

138 **3.5.2 Amino acids.** The coffee husk contains a protein content ranging from 8 to 11 % on a dry  
 139 matter basis [34], with a relatively high content of amino acids such as glutamic acid (7.7% of  
 140 the total protein content) and aspartic acid (7.1%) [34]. Glutamic acid is responsible for the  
 141 transport of glutamine and other amino acids through the blood, and its presence decreases the  
 142 need to consume sugar and alcoholic beverages. The aspartic acid is involved in the metabolism  
 143 of DNA and RNA, but also in protecting the liver and boosting the immune system. So, coffee  
 144 industry by-products are a source of amino acids that could be evaluated as dietary  
 145 phytochemicals useful for human beings. Dietary supplements and/or food fortification based on  
 146 coffee by-product production may be feasible too [35][36]. Table 3 shows a comprehensive view  
 147 of the main amino acids present in the coffee husk.

148

Table 3: Content of [protein and of](#) the main amino acids in coffee husk and pulp. From [27]  
 [34].

<b>Protein content</b>	<b>8-11%, on a dry matter content</b>
<b>Amino acid</b>	<b>% with respect to the total protein content</b>
Glutamic acid	7.7
Aspartic acid	7.1
Leucine	4.7
Glycine	4.2
Proline	3.7
Valine	3.7

Alanine	3.5
Lysine	3.4
Serine	3.3
Isoleucine	3.3
Threonine	3.1
Phenylalanine	3.0
Arginine	2.8
Histidine	2.5
Tyrosine	1.9
Methionine	0.3
Cystine	0.3

149

150 **3.5.3 Volatile oils.** Al-Yousef and Amina [37], working on coffee husk from *Coffea arabica* L.,  
151 reported of the content of a volatile oil made of at least 55 molecules. As reported in Table 4, the  
152 main chemical compositions of volatile oil in the essential oil of coffee husk, as determined by  
153 gas chromatography-mass spectroscopy (GC-MS), is mainly represented by butylatedhydroxy  
154 (65.83%), with much smaller content of 1,2-benzenedicarboxylic acid (7.28%), phenylethyl  
155 alcohol and octanoic acid (1.69% each), and 2,3-isopropylidene-6-decoxyhexo (1.63%).  
156 According to the mass spectra observation, 30% and 40% of the compounds present in the oil are  
157 hydrocarbon and oxygenated constituents, respectively, while aromatic compounds dominates.  
158 Volatile components showed antibacterial, antifungal, and antioxidant potentiality that are  
159 helpful in the treatment of infection diseases. In Table 4, the total time required to analyse a  
160 single sample was 58 min; and the components were identified on the basic of GC-MS retention  
161 time. M+ represents molecular ions, which are important for determining the molecular weight  
162 by GC-MS.

163

Table 4: Content of the main volatile oils in the essential oil of coffee husks. From [37].

	Required time (min)	Area (%)	M <sup>+</sup> (g)
--	---------------------	----------	--------------------

Butylated hydroxytoluene	24.2	65.83	220.18
1,2-benzenedicarboxylic acid	31.92	7.28	278.34
Phenylethyl alcohol	13.66	1.69	122.09
Octanoic acid	15.76	1.69	144.21
2,3-isopropylidene-6-deoxyhexo	26.12	1.63	220
Decane, 1,1'-oxybis-	47.9	1.59	298.54
Nonanoic acid	18.4	1.58	158.16
1,2-benzenedicarboxylic acid	33.78	1.37	278.35
Beta-d-arabino-2-hexulopyran	24.66	1.17	234.00
Oxalic acid, 2-ethylhexyl tetr	44.78	1.11	398.61
Hexatriacontane	49.36	1.00	506.97

M+ is the molecular ion, expressed as the ratio between mass and charge number of ions (M/Z); since Z is almost always 1 in GC-MS, M+ is mainly generally the mass (g) of the ionic molecule

Al-Yousef and Amina [37], evaluated the volatile oil and total alcohol extract of coffee husk for their antimicrobial activity with respect to three well-known antibiotics like ~~Ampicillin and Doxycycline~~ ~~doxycycline~~, were used as positive control against bacteria, while ~~and Nystatin~~ ~~nystatin~~, was used as the control antifungal drug. In the experiment, minimum inhibitory concentration of ethanol extracts as well as volatile oil of coffee husk against drug resistant clinical strains was determined. The results are shown in Table 5. Both volatile oil and total alcohol extract of coffee husk reduced the growth of *Staphylococcus aureus*, *Escherichia coli*, *Pseudomonas aeruginosa*, and *Candida albicans* colonies by 50 to 104% with respect to three tested antibiotics, with the alcohol extract being more efficient than volatile oil for *Staphylococcus aureus* and *Pseudomonas aeruginosa*.

Table 5. Antimicrobial activity and minimum inhibitory concentration of essential oil and total alcohol extract of coffee husk from *Coffea arabica* L. on the growth of four infective microbes [37].

Samples	<i>Staphylococcus aureus</i>	<i>Escherichia coli</i>	<i>Pseudomonas aeruginosa</i>	<i>Candida albicans</i>
	Zone of inhibition (mm)			
Coffee husk essential oil	14.0 ± 1.3 (-66.7%)	17.0 ± 1.9 (-68.0%)	13.0 ± 2.0 (-50.0%)	15.0 ± 1.3 (-5.2%)

Total alcohol extract	22.0 ± 0.5 (-104.8%)	17.0 ± 2.5 (-68.0%)	20.0 ± 1.1 (-83.3%)	14.0 ± 1.2 (-0.9%)
Ampicillin	21	-	-	-
Doxycycline	-	25	24	-
Nystatin	-	-	-	23

Samples	Minimum inhibitory concentration (mg mL <sup>-1</sup> )			
	<i>Staphylococcus aureus</i>	<i>Escherichia coli</i>	<i>Pseudomonas aeruginosa</i>	<i>Candida albicans</i>
Coffee husk essential oil	0.8	0.8	0.8	0.8
Total alcohol of coffee husk	3.2	>3.2	3.2	>3.2

\*Values of Minimum inhibitory concentration is given in % v/v for dry oils

178

179 **3.5.4 Phenolic acids.** The main phenolic acids of fresh coffee husk are caffeine, tannins, and  
 180 chlorogenic acids (for quantities see Table 1), whose presence prevents various uses of fresh  
 181 coffee husk because of their ecotoxicological concerns [38]. For example, fresh coffee husk is  
 182 not suitable for animal feeding because of its anti-nutritional properties due to the excess of  
 183 phenolic acids [39]. Thus, chlorogenic acid has phytotoxic effects able to decrease seed  
 184 germination and plant growth and, because of this, it cannot be distributed in soil as soil fertilizer  
 185 [40][38]. In addition, caffeine and tannins negatively affect aquatic organisms like algae, sea  
 186 urchin, and fishes, which develop morphological and behavioral abnormalities [41].

187 *Caffeine.* The alkaloid caffeine (1,3,7-trimethylxanthine) has been found in more than 60 plant  
 188 species, with the highest levels in coffee beans, tea, and cocoa. Other two alkaloids of the  
 189 xanthine derivative group are theobromine (3,7-dimethylxanthine) and theophylline (1,3-  
 190 dimethylxanthine) [42]. Generally, caffeine has positive effects on humans as it has chemical  
 191 structure like that of adenosine, so it is well-known as adenosine receptor. Because of this,  
 192 caffeine may help to be relaxed and sleep [43]. Caffeine can be also able to contrast obesity and  
 193 diabetes [44], as well as Parkinson's [45] and Alzheimer's symptoms [46]. However, caffeine  
 194 has negative effects on the environment as it is toxic to aquatic organisms and mammals, and  
 195 has negative effects on animals, plants, fungal and bacterial growth [47].

196 *Tannins*. Tannins are commonly found in the bark of vascular plants and, to a lesser extent, into  
 197 leaves, fruit, flowers, and seeds [48]. Tannins are considered as anti-nutritional compound, and  
 198 this aspect limits the use of coffee husk in animal feed [49]. Benefits of tannins for human health  
 199 include antibacterial and antifungal activity, ~~which destroys and prevents the development of~~  
 200 ~~bacteria and fungi~~ [50], antimicrobial activity, ~~which is being~~ effective against ~~bacteria, fungi,~~  
 201 ~~parasites, and some viruses~~ [51], ~~and~~ anti-inflammatory [52], and anti-allergy [53] activities.  
 202 Tannins are also known for their low biodegradability; because of this reason they tend to remain  
 203 for long time in the environment and accumulates in the food chain [54].

204 *Chlorogenic acid*. The esterification of caffeic acid with quinic acid produces chlorogenic acid,  
 205 which is a soluble polyphenol [55] that plays many human health benefits, including neuronal  
 206 cell death protection [56] and anticancer activity [57][58]. Chlorogenic acid plays positive roles  
 207 also in plant functions including cell wall synthesis, wound healing, and root hair formation [59].  
 208 However, depending on its concentration, it may play a negative role especially in roots [60].  
 209 Villarino *et al.* [61], reported the inhibitory effect of chlorogenic acid on fungal growth, due to  
 210 its role on plant defence. The content of other phenolic acids extracted from coffee husk is  
 211 reported in Table 6.

212

Table 6. Phenolic components of coffee husk (modified from [62]).

Method	Condition/Solvent	Epicatechin	Gallic acid	Tannic acid	Protocatechuic acid	Vanillic acid
µg of gallic acid equivalent per kg of coffee husk (dry matter)						
Ultrasonication	Ethanol	-	-	-	-	2346.7
Soxhlet	Ethyl acetate	47.6	3869.2	-	-	-
	Ethanol	-	-	3859.2	-	-
SFE* CO <sub>2</sub>	200bar/40°C	-	14.85	-	-	-
	300bar/40°C	32.55	-	-	12.4	-

213 \* SFE: supercritical fluid extraction.

214

215 Secondary metabolites in coffee husk such as caffeine and other phenolic compounds are good  
 216 source of antioxidants. Table 7 shows the antioxidant capacity of aqueous extract of coffee husk  
 217 evaluated following DPPH (2,2-diphenyl-1-picryl-hydrazyl-hydrate) radical sequestration  
 218 method and inhibition of co-oxidation of  $\beta$ -carotene and linoleic acid percentage [63]. For this  
 219 experiment, coffee fruits were randomly collected at four different farm locations, from two  
 220 plants in the northern (husk 1 and grains 1), southern (husk 2 and grains 2), eastern (husk 3 and  
 221 grains 3), western (husk 4 and grains 4) and central region (husk 5 and grains 5) of the plantation  
 222 [63]. Results indicated that micro-environmental conditions present in the plantation affect the  
 223 antioxidant capacity of aqueous extract of coffee husk.

224

Table 7. Antioxidant capacity of aqueous extracts of coffee husk evaluated by the free DPPH radical sequestration method and inhibition of co-oxidation of  $\beta$ -carotene and linoleic acid [63].

Sample	DPPH EC50* (mg mL <sup>-1</sup> )	Inhibition of co-oxidation of $\beta$ -carotene and linoleic acid (%)
Coffee Husk 1	4.71 <sup>f</sup>	40.78 <sup>bc</sup>
Coffee Husk 2	3.57 <sup>h</sup>	34.88 <sup>c</sup>
Coffee Husk 3	4.44 <sup>g</sup>	43.74 <sup>abc</sup>
Coffee Husk 4	2.73 <sup>j</sup>	44.55 <sup>abc</sup>
Coffee Husk 5	3.44 <sup>i</sup>	40.80 <sup>bc</sup>
Coffee Grain 1	15.09 <sup>a</sup>	68.58 <sup>a</sup>
Coffee Grain 2	11.48 <sup>b</sup>	66.43 <sup>ab</sup>
Coffee Grain 3	10.44 <sup>c</sup>	58.22 <sup>abc</sup>
Coffee Grain 4	10.10 <sup>d</sup>	64.65 <sup>ab</sup>
Coffee Grain 5	7.53 <sup>e</sup>	68.22 <sup>a</sup>

225  
 226  
 227

\* EC50 = half maximal effective concentration.  
 For each column, means followed by different letters differed for  $P < 0.05$ , by the Tukey test.

228 **3.5.5 Lignocellulosic materials.** Cellulose, hemicellulose and lignin are the principal  
 229 lignocellulosic components forming plant cell walls. Lignocellulosic compounds like phenolic

230 acids often prevent coffee husk usage and degradations, so it is necessary to find techniques able  
231 to break down these substances ~~prevention and let these materials to recycle quickly~~ [64][65].

232 According to Oliveira *et al.* [66], the lignin of coffee husk represents a significant resource for  
233 enabling use of coffee husk as raw material for ~~emerging~~ biorefineries where lignin can be  
234 separated from other coffee husk components with a pre-treatment ~~with~~ by diluted acid followed  
235 by soda extraction. ~~Also, the~~ The extracted lignin can be then ~~wet~~-oxidized under aqueous and  
236 alkaline conditions, in order to produce valuable products such as low molecular weight  
237 biochemicals.

238 Many factors, like lignin content, crystallinity of cellulose, and particle size, limit the  
239 digestibility of hemicellulose and cellulose. Pre-treatments improve the digestibility of the  
240 lignocellulosic material. Each pre-treatment has its own effect(s) on ~~the~~ cellulose, hemicellulose,  
241 and lignin. Many thermal, acid, alkaline, and oxidative pre-treatments- have been evaluated for  
242 improving biodegradability of lignocellulose substrates [64]. For example, Baêta *et al.* [67] pre-  
243 treated coffee husk by a steam explosion technique that increased the bioavailability and  
244 biodegradability of cellulose,— broke down the lignocellulose structural components, and  
245 produced soluble organic compounds. This method is effective for increasing the anaerobic  
246 biodegradability too.

247 **3.5.6 Flavorings.** The most important characters of coffee as a beverage are acidity, aroma, and  
248 taste. Without acidity, the coffee is approximately tasteless [68]. Sampaio *et al.* [69] reported that  
249 coffee husk is a valuable by-product due to its aroma and presence of sugars that can be  
250 converted to ethanol. Table 8 shows flavor and aroma inside beverages made by different  
251 concentration of coffee husk and pineapple juice.

252

Table 8. The sensorial analysis of flavor, aroma and overall appearance of the beverage developed with the rinds of coffee with different concentrations and pineapple juice [63].

Sample	Flavor	Aroma	Overall impression
	Score		
F1*	2.96 ± 1.88 <sup>b</sup>	4.25 ± 2.06 <sup>b</sup>	3.75 ± 1.90 <sup>b</sup>
F2**	4.86 ± 2.02 <sup>a</sup>	5.44 ± 1.91 <sup>a</sup>	5.05 ± 2.01 <sup>a</sup>
F3***	5.48 ± 2.11 <sup>a</sup>	5.44 ± 1.95 <sup>a</sup>	5.48 ± 2.05 <sup>a</sup>

For each column, means followed by different letters differ for  $P < 0.05$ , by the Tukey test.

\*100% coffee husk extract; \*\*90% coffee husk extract+10% concentrated pineapple juice; \*\*\*80% coffee husk extract+20% concentrated pineapple juice. The evaluation was assessed by non-trained 52 judges who used a structured 9-point hedonic scale (1 = I greatly dislike, 9 = I enjoyed it very much).

According to Tables 7 and 8, aqueous extracts of coffee husk represent a promising natural source of bioactive phytochemicals, also because of their low levels of antinutrients [63]. Neves *et al.* [63] noticed that the beverage incorporated with concentrated pineapple juice presented the greatest acceptability, besides increasing the antioxidant capacity of the product. Thus, the formulated beverages constitute a promising alternative for the beverage market, given the meaningful content of phenolic constituents derived from coffee husk.

**3.5.7 Detoxification.** Phytotoxic compounds like caffeine, chlorogenic acid, and tannins (Table 1), if released into the environment from coffee waste, can have severe ecotoxicological effects. Although there are many benefits of caffeine consumption to human health, studies indicate that environmental leaching of caffeine has detrimental effects on several organisms [70]. Therefore, detoxification of coffee husks from phytotoxic compounds and antinutritional factors, or at least degrading them to a plausibly safe level for reusing or recycling, is necessary. Detoxification of coffee husk with physical, chemical, or biological methods were studied by [71][72][42], while a general review of enzymatic and microbial methods to remove caffeine is reported by [73]. Some physical (percolation), chemical (alcohol extraction), or microbial (fermentation with fungi) treatment(s) can reduce the phenolic content in coffee husk. Among all the systems to reduce the toxic effect of coffee husk, treatments with bacteria and/or fungi, and composting



275 are the most used [treatments](#) for coffee husk, and for other coffee by-products like coffee pulp  
276 and silverskin because they are more efficient and economical for controlling huge amount of  
277 waste. High concentrations of bacteria are required for caffeine detoxification since caffeine has  
278 a toxic effect for bacteria, and 0.1% concentration of caffeine inhibits protein synthesis in  
279 bacteria and yeast [71][74]. However, some microorganisms can grow in presence of caffeine  
280 and survival is due to their capacity to degrade it [75][76]. Several studies were carried out to  
281 investigate the use of purines, including caffeine, as a source of energy for microorganism  
282 growth [77]. Although fungi growing on caffeine have been isolated, most of the studies were  
283 done with bacteria isolated from soil, mainly belonging to the *Pseudomonas* group, with  
284 emphasis on *Pseudomonas putida* [78]. Yamaoka-Yano and Mazzafera [79] used *P. putida* strain  
285 and, after a short incubation periods of 9 days, observed 40% reduction of caffeine. Brand et al.  
286 [71] tested biological detoxification of coffee husk by filamentous fungi (*Rhizopus*,  
287 *Phanerochaete*, and *Aspergillus* spp.) using a solid-state fermentation system in which coffee  
288 husk was used as the sole source of C and N. *Rhizopus arrizus* LPB-79 strain showed great  
289 results on caffeine and tannins degradation (87% and 65%, respectively), which were obtained in  
290 6 days at pH 6.0 and at 60% moisture.

291 The toxicity of coffee leachate were studied in laboratory by standardised toxicity tests on  
292 aquatic organisms [80][7], and results showed that the half maximal effective concentration  
293 (EC50) of coffee leachate was 6.02% v/v on the bacterium *Vibrio fischeri*, lower for the  
294 bacterium *Daphnia similis* (EC50 of 1.5%), and even less for the microcrustacean *Ceriodaphnia*  
295 *dubia* (EC50 of 0.12%). The reduced EC50 values from bacteria to water fleas was explain as the  
296 result of increased exposure to ingestion. There are good studies on caffeine toxicity, but no  
297 toxicity test has been performed on leachate from coffee by-products. Furthermore, there are

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298 several paths through which coffee can enter the environment such as processing/roasting or the  
299 retail consumption, suggesting that there is a major gap in toxicity data for coffee industry by-  
300 products that requires urgent attention [7].

301

#### 302 **4. USES OF COFFEE HUSK**

303 Coffee husk is the main coffee by-product that has been the topic of several studies in order to  
304 use it in industrial activities, to produce biofuel, as contaminants sorbent, dietary fiber, and  
305 bioactive compounds, for the extraction of enzymes, or in agriculture as animal feeding or for  
306 making compost, silage, biochar, or mushroom substrata.

307

#### 308 **4.1. INDUSTRIAL USES**

309 **4.1.1. Coffee husk in ceramic industry.** Generally, coffee producing countries use coffee husk  
310 as solid fuel and this method produces huge amount of ashes that has environmentally side  
311 effects. Ashes obtained by coffee husk combustion (that collected from ash dumps) are rich in  
312 alkaline and alkaline-earth metals that are a candidate ~~of~~ for replacing the scarce and expensive  
313 feldspars traditionally used as fluxing component in clay based ceramic formulation. Results  
314 shows that adding 25-40 wt.% of ashes in common clay-based ceramic formulation had the best  
315 result in ceramic quality [81].

316 **4.1.2. Coffee husk in particleboards production.** Bekalo and Reinhardt [26] and Nuamsrinuan  
317 *et al.* [82] studied the use of coffee husk for partial replacement of wood (up to 50%) in the  
318 production of particleboards. ~~They tested for the mechanical properties, swelling and water~~  
319 ~~absorption standards.~~ The results of particle sheet from milling process passed the standard tests  
320 ~~of mechanical properties.~~ but while swelling and water absorption did not indicated that all the

321 ~~coffee husk sheet were not passed it. The effect of coffee husk particle size and isocyanate~~  
322 ~~adhesive on mechanical properties was clarified.~~ The coffee husk-wood board showed great  
323 promise for its use in structural and nonstructural panel products based on superior flexural and  
324 internal bond properties.

325 **4.1.3. Flavor production.** Food flavoring compounds can be produced by chemical synthesis or  
326 extracted from natural materials. Nowadays the second way is highly demanding since the  
327 obtained products are considered safer and healthier than those obtained via synthesis. Plants are  
328 acceptable source of essential oils and flavors, but their value ~~also~~ depends on factors like  
329 weather conditions and plant diseases. Due to the presence of anti-nutritional factors such as  
330 caffeine and tannins, coffee husk cannot be used directly as a flavour source [83]. Instead, when  
331 coffee husk is treated by steam to remove caffeine and chlorogenic acids, it can be used for  
332 aroma production by using fungi of the genus *Ceratocystis* [83]. Soares *et al.* [83] tested coffee  
333 husk as raw material for fruits flavor production **by solid state fermentation** and found that  
334 different dosages of glucose can determine the production of different flavors such as banana and  
335 pineapple.

336

## 337 4.2. FUEL PRODUCTION

338 **4.2.1. Coffee husk as a solid fuel.** The use of coffee husk as solid fuel is the simplest way to  
339 manage problems due to disposal or accumulation in nature, even though the production of ash is  
340 also raising concerns. In fact, the ash derived from coffee husk combustion is often the object of  
341 illegal covert disposal and the source of environmental impacts [81]. About 70% of the coffee  
342 husks produced in Kenya is used as solid fuel [84]. The coffee husk is carbonized in a kiln and  
343 then ground, coagulated, and molded in form of briquettes prior to being packed into bags. The

344 obtained coffee charcoal briquettes have better quality than wood charcoal [29][68] but, as for  
345 other agricultural residues, carbonization is not the best choice to recover energy from coffee  
346 husk, as its combustion efficiency is minimal because of not exactly suitable physicochemical  
347 properties such as low bulk density, low ash melting point, and high volatile matter content [85].

348 **4.2.2. Gasification of coffee husk.** In order to find solutions to improve energy recovery from  
349 coffee husk, gasification is a possibility to increase energy recovering by producing ignitable gas  
350 through a partial incineration at elevated temperatures and moderate heating rates. The obtained  
351 gas is a mixture of CO, H<sub>2</sub>, CH<sub>4</sub>, CO<sub>2</sub>, and N<sub>2</sub>, and temperature level is key to improve gas  
352 quality [86]. In coffee producing countries, biomass energy has potential to be the most abundant  
353 sustainable renewable energy but, to reach this goal, there is a ~~the~~ necessity to develop and  
354 sustain contemporary technologies that increase the biomass-to-energy conversion. One way can  
355 be the high temperature air/steam gasification of biomass [87]. Wilson *et al.* [87] studied coffee  
356 husk experimental gasification under high temperature conditions by batch facility and found  
357 positive influence of high temperature on increasing the gasification process. Experiments  
358 carried out at 4% O<sub>2</sub> concentration obtained the highest gasification rate (96% of the coffee  
359 husk), while 82.80% and 71.29% of the husk was gasified with gasification conditions at 2 and  
360 3%, respectively. Miito and Banadda [86] found that the 46.6 million tons per year of coffee  
361 husk produced in Uganda, with a heating value of 18.34 MJ kg<sup>-1</sup>, will address a 0.7% of the total  
362 energy consumed in the country, while protecting the environment too. The same use could be  
363 feasible in the countries where coffee is produced and processed, namely where coffee husk  
364 abounds.

365 **4.2.3. Ethanol production from coffee husk.** Coffee husk has also good potential to be used  
366 for bio-ethanol production and, as for gasification, temperature and yeast concentration are key

367 to control the quality of the production with batch fermentation method [88][27]. The availability  
368 of cellulose, hemicellulose, and lignin in coffee husk is similar to that of other agricultural  
369 residues such as sugarcane bagasse, barley and wheat straws, rice husk, and others. ~~But~~ However,  
370 because of the high amount of coffee husk generated, the toxic nature of coffee husk, the high  
371 percentage of fermentable sugar, and the presence of high concentration of carbohydrates, it  
372 could be a good source of raw material for bio-alcohol production [89].

373

#### 374 4.3. ADSORPTION OF CONTAMINANTS

375 Cu, Cr, Cd, Ni, Hg, Pb, and Zn are the most abundant pollutants in industrial wastewater.  
376 Common methods for the removal of heavy metals from wastewaters include ion-exchange,  
377 filtration, electrochemical treatment, chemical precipitation, and adsorption. Since the activated  
378 charcoal used to remove organic and inorganic pollutants from aqueous effluents is expensive,  
379 and the activated charcoal produced from coffee husk showed high specific surface and porosity  
380 [90], it is a valid solution to reduce costs for wastewater treatment [91][92]. In addition, the  
381 adsorbed metals can easily desorb and the biomass be ready for final disposal [93]. Adsorption of  
382 Pb [94], Ni [95], cyanide [96], dye contaminants [97], and antibiotic norfloxacin [98] by coffee  
383 husk in batch mode is used to decontaminate aqueous solution. Berhe *et al.* [93] studied the  
384 efficiency of coffee husk to adsorb Pb(II) from industrial effluents using batch experiment and  
385 found that, at optimum adsorption conditions ~~adsorption~~ (pH 5 and- 90 min of contact time at  
386 200 rpm), there was the maximum adsorption efficiency of 95.14%.

387

#### 388 4.4. PRODUCTS OBTAINED BY FERMENTATION

389 **4.4.1. Organic acids.** Coffee husk is a cheap and available substrate to produce organic acids  
390 like gibberellin and citric acid by fermentation techniques. Shankaranand and Lonsane [99]  
391 produced citric acid from coffee husk by using *Aspergillus niger* under solid state fermentation  
392 method, and by every 10 g of coffee husk they produced 1.5 g of citric acids. Machado *et al.*  
393 [100] evaluated the feasibility of employing coffee husk as a substrate to produce gibberellic acid  
394 in both solid-state fermentation and submerged fermentation tests.

395 **4.4.2. Enzymes.** Coffee by-products can be also used to produce enzymes like pectinase,  
396 tannase, and caffeinase by two main industrial enzyme production methods: solid-state  
397 fermentation and submerged fermentation [101]. However, Battestin and Macedo [102] produced  
398 tannase from coffee husk by using *Paecilomyces variotti*, while Murthy and Naidu [101] studied  
399 the production of amylase, protease, and xylanase by fungal organisms.

400

#### 401 **4.5. BIOACTIVE COMPOUNDS**

402 Bioactive compounds are an extra nutritional factor typically present in small quantities in foods  
403 that have been intensively studied to evaluate their effects on health. Some of those are phenolic  
404 compounds or antibiotic molecules, while other have an anti-inflammatory, hepatoprotective,  
405 and antioxidant activity, or the ability to improve cognitive capabilities [7]. Agro-industrial by-  
406 products are good sources of bioactive compounds and have been explored as sources of natural  
407 antioxidants [103].

408 **4.5.1. Dietary fiber.** Agro-wastes are great sources of dietary fiber, which include cellulose,  
409 hemicelluloses, lignin, pectin, gums, and other polysaccharides. The soluble and insoluble  
410 dietary fibers have a wide range of health benefits, such as reduction of the risks of  
411 gastrointestinal diseases, cardiovascular diseases, and obesity [104]. The kind of coffee and the

412 degree of roasting and extraction method influence the dietary fiber content and structural  
413 characterization of coffee husk and other coffee by-products, as determined by the Association of  
414 Official Agricultural Chemistry – (AOAC) methods for the determination of dietary fiber is well  
415 known as a global standard method used for labelling food products [105].

416 **4.5.2. Anthocyanins.** Anthocyanins are flavonoid compounds responsible for the red/blue color  
417 of many fruits and flowers. By using concentrated methanol as extractant, Prata and Oliveira  
418 [106] reported cyanidin 3-rutinoside as the dominant anthocyanin in coffee husk, so ~~it~~ this latter  
419 could be used as a source for anthocyanin pigments as natural food colorant. Anthocyanins is  
420 extracted by concentrated methanolic from coffee husk.

421

## 422 4.6. AGRICULTURE

423 Uses of coffee husk in agriculture can be many, but the high content of phenolic acids and the  
424 mutagenic effect of caffeine suggest that recycling of coffee husk in agriculture should be  
425 preceded by detoxification process(es) able to decrease the concentration of these components.

426 Information about detoxification of coffee husk is given at point 3.5.7.

427 **4.6.1. Animal food.** Agricultural industry by-products like coffee husk as livestock food is  
428 important to reduce the food competition and help to environment sustainability. Yearly coffee  
429 roasting industry produces million tons of coffee husks that contain valuable nutrients like  
430 proteins, carbohydrates, and minerals. Huge amount of production and good nutrients content  
431 make coffee husk a good material for animal food [107]. However, the idea of using coffee husk  
432 for ruminants, pigs, chickens, fishes, and rabbits was released several decades ago, but the result  
433 was not so bright and acceptable. In fact, National Dairy Board [108] reported that coffee husk is  
434 not a delicious food for cattle, which can tolerate only small portion of it because of the content

435 of phenolic components. Fishes and poultries are even more sensitive than cattle and pigs, so the  
436 quantity of coffee husk in their diet must be small [34], unless to submit the husk to a  
437 detoxification process.

438 **4.6.2. Mushroom bed.** Coffee husk is an appropriate bed for mushroom growth because of its  
439 availability and cheap price and, due to its fragmented nature, no grinding is needed before  
440 application, but it needs disinfection. Coffee husk is a good substrate for mushroom bed,  
441 especially for *Lentinula edodes* (shiitake) and *Flammulina velutipes* species [109]. Fermentation  
442 of coffee husk by the fungus *Pleurotus ostreatus* increased protein and cellulose contents and  
443 decreased the proportion of lignin, tannins, and caffeine. Further, when fermentation of coffee  
444 husk increased, the volatile fatty acid and digestible dry matter decreased [109].

445 **4.6.3. Biochar.** Biochar is charcoal produced by pyrolysis of organic materials and can be used  
446 as soil amendment and fertilizer [110]. The biochar quality depends on the nature of the raw  
447 material and temperature. Acid soils, which abound in tropical areas, have deficiencies in plant  
448 nutrients like N, P, K, Ca, and Mg, and consequently have low crop production rates. Adding  
449 biochar reduces the soil acidity due to the alkalinity of biochar and increases the availability of  
450 nutrients and water [111]. Studies on coffee husk biochar showed that it improved soil chemical  
451 properties by increasing pH, electrical conductivity, cation exchange capacity, organic matter,  
452 total N, and available P [112]. Dume *et al.* [113] reported that the application of 15 tons ha<sup>-1</sup>  
453 coffee husk biochar that had been produced at 500°C temperature had positive result on soil  
454 fertility and yield. Deal *et al.* [114] compared the performance of biochar in five different  
455 feedstocks (coffee husk, maize cobs, eucalyptus wood, groundnut shells, and rice husks) in the  
456 humid tropics. Results showed that biochar from coffee husk were the most productive in the



457 maize field. The soil pH in tropical area are so acidic (pH=4.7) and pH increasing because of  
458 soluble coffee husk biochar improve soil quality and efficiency for crop production.

459 **4.6.4. Silage and composting.** Silage is the direct usage of organic residues on soil surface  
460 without any treatment, while composting is the biological decomposition of organic waste  
461 promoted by bacteria, fungi, worms, and other organisms under controlled aerobic conditions to  
462 obtained a partially decayed organic matter [115]. The chemical composition of coffee husk in  
463 terms on nutritive elements like N, K, P, and others (Table 2) makes it suitable to be used as  
464 amendment in agricultural soils.

465 **4.6.4.1. Silage.** Coffee husk silage can be a good option for K depleted soils, but there is the risk  
466 of phytotoxic production [116]. Application of raw coffee husk in the field inhibits the plants,  
467 specially the roots, while anaerobic decomposition increases the emission of greenhouse gases  
468 [30]. It was observed that addition of coffee husk on soil provided an increase in dry matter  
469 content, but also decreased the buffering capacity responsible for maintaining soil pH [15].  
470 However, coffee husk spread at the soil surface may decrease the soil erosion, temperature, and  
471 evapotranspiration. So, notwithstanding the problems due to its phytotoxic activity, coffee husk  
472 can help in land reclamation [116].

473 **4.6.4.2. Composting.** One of the most important problem in coffee industry is by-products  
474 accumulation and, subsequently, economic and environmental costs for their management due to  
475 their potential contamination effect caused by the leaching of phenolic compounds. In fact,  
476 notwithstanding the many different uses to which coffee husk can be addressed, in coffee  
477 producer countries every year huge amounts of coffee husk are produced and, especially in  
478 developing countries, much of this husk is released in the land without any pre-treatment.  
479 Instead, phenolic acids content and mutagenic effects of caffeine require to treat coffee husk

480 before land distribution to reduce its environmental concern. As reported at point 3.5.7, there are  
481 several ways to remove inhibitors from coffee husk, but composting is the most affordable,  
482 environmentally friendly, and efficient system. Because of this, different investigations aimed to  
483 improve waste management and ecosystem sustainability have been done on coffee husk so to  
484 transform a disposal problem into a valuable product for agriculture. Composting of coffee husk  
485 with other organic materials or alone is one of the best ways to profitably manage coffee husk  
486 since the process has capacity to solve management problems like mass accumulation and  
487 detoxification. Composting by oxygen-driven biological methods allows easily recycling great  
488 amounts of agricultural by-products and producing high-quality fertilizers [117][30]. Coffee  
489 husk has characteristics that make it suitable to be composted; for instance, it has a C/N ration  
490 around 30 [30] and is rich in lignocelluloses materials, which makes it an ideal substrate for  
491 microbial processes [2]. Inoculation of lignocellulosic waste materials with lignin-degrading  
492 microorganisms accelerates the composting process and improves compost quality and the  
493 humification process [118].

494 Dzung *et al.* [119] studied coffee husk supplemented with cow manure and lime. The mixture  
495 was composted for 3 months and then was supplemented with 0.1% (w/w) effective  
496 microorganisms like N<sub>2</sub>-fixing *Azotobacter* sp. and *Bacillus megaterium*; the authors found that  
497 the quality of the obtained compost was better than some bio-organic fertilizers present on the  
498 agriculture market. This compost was applied on coffee field and the results showed that soil  
499 fertility, nutrient content in the coffee leaves, and the growth of the coffee plants were improved  
500 in comparison with the control. Sekhar *et al.* [120] applied different dosages of coffee husk  
501 compost with NPK fertilizers in various amounts in the paddy field and found that applications  
502 of 4 ton ha<sup>-1</sup> of coffee husk compost plus 80 kg ha<sup>-1</sup> N, 60 kg ha<sup>-1</sup> P, and 50 kg ha<sup>-1</sup> K gave the

503 highest grain and straw yield production. Kassa and Workayehu [31] evaluated the quality of  
504 composts comparing the quality of only coffee husk compost with mixtures made of coffee  
505 husk+cow dung, coffee husk+*Millettia ferruginea*, coffee husk+cow dung+*Millettia ferruginea*,  
506 and coffee husk+effective microorganism, and concluded that the mixtures coffee husk+*Millettia*  
507 *ferruginea* and coffee husk+cow dung+*Millettia ferruginea* gave the highest quality composts. In  
508 the coffee husk composting experiments run by Bidappa [121][108] and Tuan [122], as we may  
509 deduce from the fact that the use of these composts improved soil fertility and crop yield, a  
510 strong reduction of phenolic compounds was obtained. Shemekite *et al.* [30] used cow dung and  
511 green wastes as co-substrates in the composting of coffee husk and monitored the  
512 physicochemical changes and the microbial community dynamics during the composting  
513 process. While at the beginning of the process the microbial communities of all the compost piles  
514 differed, they were similar at the end, as shown by DGGE fingerprints and microarray analysis.  
515 Improving soil fertility and plant growth is one of the benefits coming from compost application  
516 in agriculture. Other helpful impacts are the decrease of soil erosion and evapotranspiration,  
517 which may contribute to land reclamation. Thus, since composting process disinfects organic  
518 wastes from pathogens and weed seeds and stabilized C, N, and other nutrients in the organic  
519 fraction, applying compost to the field can help to maintain or increase the soil organic matter  
520 content, biological activity, and porosity, so helping water, air, and plant roots to penetrate easily  
521 the soil [123][117].

522

#### 523 **4.7. RESUMING OF COFFEE HUSK APPLICATIONS APPROACH**

524 Table 9 shows a comprehensive view of the possible uses of coffee husk ~~as it comes~~  
525 ~~from~~obtained by processing coffee ~~processing~~cherries by dry method.

Table 9. Possible uses of coffee husk in industrial, fuel, agri-food, and agriculture activities.

Application	Reference
<i>Industrial use</i>	
Ceramic	[81]
Particleboard	[26][82]
Flavor extraction	[83]
<i>Fuel</i>	
Solid fuel	[84][85]
Gasification	[86][87]
Ethanol production	[88][89][27]
<i>Contaminants adsorption</i>	
Lead (Pb)	[94][93]
Nickel (Ni)	[95]
cyanide	[96]
dye contaminants	[97]
norfloxacin	[98]
<i>Fermented products</i>	
Organic acid	[99][100]
Enzymes	[102]
<i>Bioactive compounds</i>	
Dietary fiber	[104]
Anthocyanin	[106]
<i>Agriculture</i>	
Animal food	[34]
Mushroom bed	[109]
Biochar	[112]
Silage	[116]
Compost	[30][120][121][122]

527

## 528 **5. CONCLUSIONS**

529 Coffee consumption in the world increases every year and the same happens for its by-products.

530 Coffee husk is the main by-product of coffee roasting process by dry method and is one of the

531 most abundant by-products that are spread in the land, giving rise to some environmental  
532 concerns. Nonetheless, coffee husk components make this material suitable to be used in several  
533 ways in many industrial, fuel, agri-food, and agriculture activities. Because of its high content of  
534 phenolic compounds, the use of coffee husk may require detoxification, and many systems have  
535 been identified to reduce the toxic effect of coffee husk; all these systems are reported in this  
536 review. The use of coffee husk as direct or indirect fuel is one of the most practiced way to  
537 recycle it but, because of its content in nutritive elements, the use in agriculture should be  
538 promoted, especially in acid soils, possibly after composting instead to be directly used as soil  
539 silage. However, the lack of local application and performing of the scientific results obtained at  
540 a global scale is a challenge that should be the topic of future studies in order to improve  
541 recycling of these valuable materials and increase soil fertility.

#### 542 **References**

- 543 [1] M. Daglia, A. Papetti, C. Gregotti, F. Bertè, G. Gazzani, In vitro antioxidant and ex vivo  
544 protective activities of green and roasted coffee, *J. Agric. Food Chem.* 48 (2000) 1449–  
545 1454. <https://doi.org/10.1021/jf990510g>.
- 546 [2] P.S. Murthy, M. Madhava Naidu, Sustainable management of coffee industry by-products  
547 and value addition - A review, Elsevier B.V., 2012.  
548 <https://doi.org/10.1016/j.resconrec.2012.06.005>.
- 549 [3] L.R. Batista, S.M. Chalfoun de Souza, C.F. Silva e Batista, R.F. Schwan, *Coffee: Types*  
550 *and Production*, 1st ed., Elsevier Ltd., 2015. [https://doi.org/10.1016/B978-0-12-384947-](https://doi.org/10.1016/B978-0-12-384947-2.00184-7)  
551 [2.00184-7](https://doi.org/10.1016/B978-0-12-384947-2.00184-7).

- 552 [4] 2014 revision of the World Urbanization Prospects | Latest Major Publications - United  
553 Nations Department of Economic and Social Affairs, (n.d.).  
554 [https://www.un.org/en/development/desa/publications/2014-revision-world-urbanization-](https://www.un.org/en/development/desa/publications/2014-revision-world-urbanization-prospects.html)  
555 [prospects.html](https://www.un.org/en/development/desa/publications/2014-revision-world-urbanization-prospects.html) (accessed October 30, 2019).
- 556 [5] L.C. Monaco, M.R. Sondahl, A. Carvalho, O.J. Crocomo, W.R. Sharp, Applications of  
557 tissue culture in the improvement of coffee, *Appl. Fundam. Asp. Plant Cell, Tissue, Organ*  
558 *Cult.* (1977).
- 559 [6] L.S. Oliveira, A.S. Franca, An Overview of the Potential Uses for Coffee Husks, Elsevier  
560 Inc., 2014. <https://doi.org/10.1016/B978-0-12-409517-5.00031-0>.
- 561 [7] B. Janissen, T. Huynh, Chemical composition and value-adding applications of coffee  
562 industry by-products: A review, *Resour. Conserv. Recycl.* 128 (2018) 110–117.  
563 <https://doi.org/10.1016/j.resconrec.2017.10.001>.
- 564 [8] Clarke R. J., Macrae R., *Coffee: Volume 1: Chemistry*, springer, 1985.  
565 [https://books.google.it/books?hl=en&lr=&id=PaByBgAAQBAJ&oi=fnd&pg=PT9&dq=C](https://books.google.it/books?hl=en&lr=&id=PaByBgAAQBAJ&oi=fnd&pg=PT9&dq=Coffee++Chemistry++Volume+1++Chemistry&ots=SURyr0R7V0&sig=QTX530dIfzTCE7ZLKBRWEHoBhGs&redir_esc=y#v=onepage&q=Coffee+Chemistry+Volume+1+Chemistry&f=false)  
566 [offee++Chemistry++Volume+1++Chemistry&ots=SURyr0R7V0&sig=QTX530dIfzTCE7](https://books.google.it/books?hl=en&lr=&id=PaByBgAAQBAJ&oi=fnd&pg=PT9&dq=Coffee++Chemistry++Volume+1++Chemistry&ots=SURyr0R7V0&sig=QTX530dIfzTCE7ZLKBRWEHoBhGs&redir_esc=y#v=onepage&q=Coffee+Chemistry+Volume+1+Chemistry&f=false)  
567 [ZLKBRWEHoBhGs&redir\\_esc=y#v=onepage&q=Coffee Chemistry Volume 1](https://books.google.it/books?hl=en&lr=&id=PaByBgAAQBAJ&oi=fnd&pg=PT9&dq=Coffee++Chemistry++Volume+1++Chemistry&ots=SURyr0R7V0&sig=QTX530dIfzTCE7ZLKBRWEHoBhGs&redir_esc=y#v=onepage&q=Coffee+Chemistry+Volume+1+Chemistry&f=false)  
568 [Chemistry&f=false](https://books.google.it/books?hl=en&lr=&id=PaByBgAAQBAJ&oi=fnd&pg=PT9&dq=Coffee++Chemistry++Volume+1++Chemistry&ots=SURyr0R7V0&sig=QTX530dIfzTCE7ZLKBRWEHoBhGs&redir_esc=y#v=onepage&q=Coffee+Chemistry+Volume+1+Chemistry&f=false) (accessed June 15, 2020).
- 569 [9] H. Steinhart, *Coffee: growing, processing, sustainable production—a guidebook for*  
570 *growers, processors, traders, and researchers*. Jean Nicolas Wintgens (Ed). Wiley-VCH  
571 Verlag, Weinheim, 2004. 976 pp, ISBN 3-527-30731-1, *J. Sci. Food Agric.* 85 (2005)  
572 1966–1966. <https://doi.org/10.1002/jsfa.2234>.

- 573 [10] C. Rodrigues, R. Maia, M. Ribeirinho, P. Hildebrandt, L. Gautz, T. Prohaska, C. Máguas,  
574 Coffee, in: *Compr. Anal. Chem.*, 2013: pp. 573–598. [https://doi.org/10.1016/B978-0-444-](https://doi.org/10.1016/B978-0-444-59562-1.00022-0)  
575 [59562-1.00022-0](https://doi.org/10.1016/B978-0-444-59562-1.00022-0).
- 576 [11] R.C. Alves, F. Rodrigues, M. Antónia Nunes, A.F. Vinha, M.B.P.P. Oliveira, State of the  
577 art in coffee processing by-products, Elsevier Inc., 2017. [https://doi.org/10.1016/B978-0-](https://doi.org/10.1016/B978-0-12-811290-8.00001-3)  
578 [12-811290-8.00001-3](https://doi.org/10.1016/B978-0-12-811290-8.00001-3).
- 579 [12] S. Roussos, M. de los Angeles Aquihuatl, M. del Refugio Trejo-Hernández, I. Gaime  
580 Perraud, E. Favela, M. Ramakrishna, M. Rimbault, G. Viniegra-González,  
581 Biotechnological management of coffee pulp - isolation, screening, characterization,  
582 selection of caffeine-degrading fungi and natural microflora present in coffee pulp and  
583 husk, *Appl. Microbiol. Biotechnol.* 42 (1995) 756–762.  
584 <https://doi.org/10.1007/BF00171958>.
- 585 [13] R.M. Couto, J. Fernandes, M.D.R.G. da Silva, P.C. Simões, Supercritical fluid extraction  
586 of lipids from spent coffee grounds, *J. Supercrit. Fluids.* 51 (2009) 159–166.  
587 <https://doi.org/10.1016/j.supflu.2009.09.009>.
- 588 [14] R.C. Alves, S. Casal, M.B.P.P. Oliveira, Tocopherols in coffee brews: Influence of coffee  
589 species, roast degree and brewing procedure, *J. Food Compos. Anal.* 23 (2010) 802–808.  
590 <https://doi.org/10.1016/j.jfca.2010.02.009>.
- 591 [15] A.S. Franca, L.S. Oliveira, Coffee processing solid wastes: Current uses and future  
592 perspectives, 2009.
- 593 [16] P. Mazzafera, Chemical composition of defective coffee beans, *Food Chem.* 64 (1999)

- 594 547–554. [https://doi.org/10.1016/S0308-8146\(98\)00167-8](https://doi.org/10.1016/S0308-8146(98)00167-8).
- 595 [17] S.I. Mussatto, E.M.S. Machado, S. Martins, J.A. Teixeira, Production, Composition, and  
596 Application of Coffee and Its Industrial Residues, *Food Bioprocess Technol.* 4 (2011)  
597 661–672. <https://doi.org/10.1007/s11947-011-0565-z>.
- 598 [18] L.F. Ballesteros, J.A. Teixeira, S.I. Mussatto, Chemical, Functional, and Structural  
599 Properties of Spent Coffee Grounds and Coffee Silverskin, *Food Bioprocess Technol.* 7  
600 (2014) 3493–3503. <https://doi.org/10.1007/s11947-014-1349-z>.
- 601 [19] R.C. Borrelli, F. Esposito, A. Napolitano, A. Ritieni, V. Fogliano, Characterization of a  
602 New Potential Functional Ingredient: Coffee Silverskin, *J. Agric. Food Chem.* 52 (2004)  
603 1338–1343. <https://doi.org/10.1021/jf034974x>.
- 604 [20] A. Pourfarzad, H. Mahdavian-Mehr, N. Sedaghat, Coffee silverskin as a source of dietary  
605 fiber in bread-making: Optimization of chemical treatment using response surface  
606 methodology, *LWT - Food Sci. Technol.* 50 (2013) 599–606.  
607 <https://doi.org/10.1016/j.lwt.2012.08.001>.
- 608 [21] N. Martinez-Saez, M. Ullate, M.A. Martin-Cabrejas, P. Martorell, S. Genovés, D. Ramon,  
609 M.D. Del Castillo, A novel antioxidant beverage for body weight control based on coffee  
610 silverskin, *Food Chem.* 150 (2014) 227–234.  
611 <https://doi.org/10.1016/j.foodchem.2013.10.100>.
- 612 [22] F. Rodrigues, C. Pereira, F.B. Pimentel, R.C. Alves, M. Ferreira, B. Sarmiento, M.H.  
613 Amaral, M.B.P.P. Oliveira, Are coffee silverskin extracts safe for topical use? An in vitro  
614 and in vivo approach, *Ind. Crops Prod.* 63 (2015) 167–174.



- 615 <https://doi.org/10.1016/j.indcrop.2014.10.014>.
- 616 [23] M.A. Ramirez-Coronel, N. Marnet, V.S.K. Kolli, S. Roussos, S. Guyot, C. Augur,  
617 Characterization and Estimation of Proanthocyanidins and Other Phenolics in Coffee Pulp  
618 (*Coffea arabica*) by Thiolysis-High-Performance Liquid Chromatography, J. Agric. Food  
619 Chem. 52 (2004) 1344–1349. <https://doi.org/10.1021/jf035208t>.
- 620 [24] P. Esquivel, V.M. Jiménez, Functional properties of coffee and coffee by-products, Food  
621 Res. Int. 46 (2012) 488–495. <https://doi.org/10.1016/j.foodres.2011.05.028>.
- 622 [25] P.N. Navya, S.M. Pushpa, Production, statistical optimization and application of  
623 endoglucanase from *Rhizopus stolonifer* utilizing coffee husk, Bioprocess Biosyst. Eng.  
624 36 (2013) 1115–1123. <https://doi.org/10.1007/s00449-012-0865-3>.
- 625 [26] S.A. Bekalo, H.-W. Reinhardt, Fibers of coffee husk and hulls for the production of  
626 particleboard, Mater. Struct. 43 (2010) 1049–1060. [https://doi.org/10.1617/s11527-009-](https://doi.org/10.1617/s11527-009-9565-0)  
627 9565-0.
- 628 [27] B.M. Gouvea, C. Torres, A.S. Franca, L.S. Oliveira, E.S. Oliveira, Feasibility of ethanol  
629 production from coffee husks, Biotechnol. Lett. 31 (2009) 1315–1319.  
630 <https://doi.org/10.1007/s10529-009-0023-4>.
- 631 [28] C.F. Mhilu, Analysis of Energy Characteristics of Rice and Coffee Husks Blends, ISRN  
632 Chem. Eng. 2014 (2014) 1–6. <https://doi.org/10.1155/2014/196103>.
- 633 [29] M.R. Adams, J. Dougan, Waste Products, Coffee. (1987) 257–291.  
634 [https://doi.org/10.1007/978-94-009-3417-7\\_9](https://doi.org/10.1007/978-94-009-3417-7_9).

- 635 [30] F. Shemekite, M. Gómez-Brandón, I.H. Franke-Whittle, B. Praehauser, H. Insam, F.  
636 Assefa, Coffee husk composting: An investigation of the process using molecular and  
637 non-molecular tools, *Waste Manag.* 34 (2014) 642–652.  
638 <https://doi.org/10.1016/j.wasman.2013.11.010>.
- 639 [31] H. Kassa, T. Workayehu, Evaluation of some additives on coffee residue (coffee husk and  
640 pulp) quality as compost, southern Ethiopia, *Int. Invent. J. Agric. Soil Sci.* 2 (2014) 2408–  
641 7254. <http://internationalinventjournals.org/journals/IIJAS>.
- 642 [32] M.C.S. Da Silva, J. Naozuka, J.M.R. Da Luz, L.S. De Assunção, P. V. Oliveira, M.C.D.  
643 Vanetti, D.M.S. Bazzolli, M.C.M. Kasuya, Enrichment of *Pleurotus ostreatus* mushrooms  
644 with selenium in coffee husks, *Food Chem.* 131 (2012) 558–563.  
645 <https://doi.org/10.1016/j.foodchem.2011.09.023>.
- 646 [33] A. Ronix, O. Pezoti, L.S. Souza, I.P.A.F. Souza, K.C. Bedin, P.S.C. Souza, T.L. Silva,  
647 S.A.R. Melo, A.L. Cazetta, V.C. Almeida, Hydrothermal carbonization of coffee husk:  
648 Optimization of experimental parameters and adsorption of methylene blue dye, *J.*  
649 *Environ. Chem. Eng.* 5 (2017) 4841–4849. <https://doi.org/10.1016/j.jece.2017.08.035>.
- 650 [34] H.L. Didanna, A critical review on feed value of coffee waste for livestock feeding, *World*  
651 *J. Biol. Biol. Sci.* 2 (2014) 72–86. <http://wsrjournals.org/journal/wjbbs>.
- 652 [35] M. Akram, H.M. Asif, M. Uzair, N. Akhtar, A. Madni, S.M. Ali Shah, Z.U. Hasan, A.  
653 Ullah, Amino acids: A review article, *J. Med. Plants Res.* 5 (2011) 3997–4000.
- 654 [36] No Title The 20 Amino Acids and Their Functionse, (2017).  
655 <https://www.lifepersona.com/the-20-amino-acids-and-their-functions>.

- 656 [37] H.M. Al-Yousef, M. Amina, Essential oil of Coffee arabica L. Husks: A brilliant source of  
657 antimicrobial and antioxidant agents., *Biomed. Res.* 29 (2018) 174–180.  
658 <https://doi.org/10.4066/biomedicalresearch.29-17-867>.
- 659 [38] A.S. Fernandes, F.V.C. Mello, S. Thode Filho, R.M. Carpes, J.G. Honório, M.R.C.  
660 Marques, I. Felzenszwalb, E.R.A. Ferraz, Impacts of discarded coffee waste on human  
661 and environmental health, *Ecotoxicol. Environ. Saf.* 141 (2017) 30–36.  
662 <https://doi.org/10.1016/j.ecoenv.2017.03.011>.
- 663 [39] J.H. Low, W.A.W.A. Rahman, J. Jamaluddin, Structural elucidation of tannins of spent  
664 coffee grounds by CP-MAS <sup>13</sup>C NMR and MALDI-TOF MS, *Ind. Crops Prod.* 69 (2015)  
665 456–461. <https://doi.org/10.1016/j.indcrop.2015.03.001>.
- 666 [40] F. Al-Charchafchi, F. Al-Quadani, Effect of Chlorogenic Acid on Germination and  
667 Seedling Growth, and on the Enzymes Activity Extracted from *Artemisia herba alba*  
668 ASSO. Part I: Germination and Seedling Growth, 2010.  
669 <https://doi.org/10.1016/j.chemosphere.2012.03.053>.
- 670 [41] A. Zarrelli, M. DellaGreca, M.R. Iesce, M. Lavorgna, F. Temussi, L. Schiavone, E.  
671 Criscuolo, A. Parrella, L. Previtiera, M. Isidori, Ecotoxicological evaluation of caffeine  
672 and its derivatives from a simulated chlorination step, *Sci. Total Environ.* 470–471 (2014)  
673 453–458. <https://doi.org/10.1016/j.scitotenv.2013.10.005>.
- 674 [42] P. Mazzafera, Degradação de cafeína por microrganismos e o emprego da palha e polpa  
675 de café descafeinados na alimentação animal, *Sci. Agric.* 59 (2002) 815–821.  
676 <https://doi.org/10.1590/S0103-90162002000400030>.

- 677 [43] R. Basheer, R.E. Strecker, M.M. Thakkar, R.W. McCarley, Adenosine and sleep–wake  
678 regulation, *Neurochem. Sleep Wakefulness*. 73 (2008) 337–362.  
679 <https://doi.org/10.1017/CBO9780511541674.013>.
- 680 [44] A. Hino, H. Adachi, M. Enomoto, K. Furuki, Y. Shigetoh, M. Ohtsuka, S.I. Kumagae, Y.  
681 Hirai, A. Jalaldin, A. Satoh, T. Imaizumi, Habitual coffee but not green tea consumption is  
682 inversely associated with metabolic syndrome. An epidemiological study in a general  
683 Japanese population, *Diabetes Res. Clin. Pract.* 76 (2007) 383–389.  
684 <https://doi.org/10.1016/j.diabres.2006.09.033>.
- 685 [45] J. Trevitt, K. Kawa, A. Jalali, C. Larsen, Differential effects of adenosine antagonists in  
686 two models of parkinsonian tremor, *Pharmacol. Biochem. Behav.* 94 (2009) 24–29.  
687 <https://doi.org/10.1016/j.pbb.2009.07.001>.
- 688 [46] A.J. Carman, P.A. Dacks, R.F. Lane, D.W. Shineman, H.M. Fillit, Current evidence for  
689 the use of coffee and caffeine to prevent age-related cognitive decline and Alzheimer’s  
690 disease, *J. Nutr. Heal. Aging*. 18 (2014) 383–392. [https://doi.org/10.1007/s12603-014-](https://doi.org/10.1007/s12603-014-0021-7)  
691 [0021-7](https://doi.org/10.1007/s12603-014-0021-7).
- 692 [47] P. Mohanpuria, S.K. Yadav, Retardation in seedling growth and induction of early  
693 senescence in plants upon caffeine exposure is related to its negative effect on Rubisco,  
694 *Photosynthetica*. 47 (2009) 293–297. <https://doi.org/10.1007/s11099-009-0045-0>.
- 695 [48] Z. Osman, Thermomechanical analysis of the tannins of *Acacia Nilotica* spp. *Nilotica* as a  
696 rapid tool for the evaluation of wood–based adhesives, *J. Therm. Anal. Calorim.* 107  
697 (2011) 709–716. <https://doi.org/10.1007/s10973-011-1721-4>.

- 698 [49] A. Pandey, C.R. Soccol, P. Nigam, D. Brand, R. Mohan, S. Roussos, Biotechnological  
699 potential of coffee pulp and coffee husk for bioprocesses, *Biochem. Eng. J.* 6 (2000) 153–  
700 162. [https://doi.org/10.1016/S1369-703X\(00\)00084-X](https://doi.org/10.1016/S1369-703X(00)00084-X).
- 701 [50] A.C.C. Sanches, G.C. Lopes, C.V. Nakamura, B.P. Dias Filho, J.C.P. De Mello,  
702 Antioxidant and antifungal activities of extracts and condensed tannins from  
703 *Stryphnodendron obovatum* Benth., *Rev. Bras. Ciencias Farm. J. Pharm. Sci.* 41 (2005)  
704 101–107. <https://doi.org/10.1590/S1516-93322005000100012>.
- 705 [51] M.M. Cowan, Plant products as antimicrobial agents, *Clin. Microbiol. Rev.* 12 (1999)  
706 564–582. <https://doi.org/10.1128/cmr.12.4.564>.
- 707 [52] J. Prothmann, M. Sun, P. Spégel, M. Sandahl, C. Turner, J. Scheuba, V.K. Wronski, J.M.  
708 Rollinger, U. Grienke, C. Santos-Buelga, A. Scalbert, G. Wu, S.K. Johnson, J.F.  
709 Bornman, S.J. Bennett, V. Singh, A. Simic, Z. Fang, J.K. Hellström, A.R. Törrönen, P.H.  
710 Mattila, M.Y.J. Kim, J.N. Hyun, J.G.J.A. Kim, J.C. Park, M.Y.J. Kim, J.G.J.A. Kim, S.J.  
711 Lee, S.C. Chun, I.M. Chung, W.Z. Kaluza, R.M. McGrath, T.T.C. Roberts, H.H.  
712 Schroder, R. Jambunathan, E.T. Mertz, A. Papadopoulou, R.J. Green, R.A. Frazier, V. De  
713 Freitas, N. Mateus, L.G. Butler, A.E. Hagerman, L.G. Butler, A. Khoddami, M.  
714 Mohammadrezaei, T.T.C. Roberts, D. Hahn, J. Faubion, L. Rooney, Relationship between  
715 phenolic compounds, anthocyanins content and antioxidant activity in colored barley  
716 germplasm, *J. Agric. Food Chem.* 53 (2017) 1713. <https://doi.org/10.1021/jf901434d>.
- 717 [53] D. Bagchi, M. Bagchi, S.J. Stohs, D.K. Das, S.D. Ray, C.A. Kuszynski, S.S. Joshi, H.G.  
718 Pruess, Free radicals and grape seed proanthocyanidin extract: Importance in human  
719 health and disease prevention, *Toxicology.* 148 (2000) 187–197.

- 720 [https://doi.org/10.1016/S0300-483X\(00\)00210-9](https://doi.org/10.1016/S0300-483X(00)00210-9).
- 721 [54] S. KOYUNLUOGLU, I. ARSLAN-ALATON, G. EREMEKTAR, F. GERMIRLI-  
722 BABUNA, Pre-Ozonation of Commercial Textile Tannins: Effects on Biodegradability  
723 and Toxicity, *J. Environ. Sci. Heal. Part A*. 41 (2006) 1873–1886.  
724 <https://doi.org/10.1080/10934520600779083>.
- 725 [55] L. Gauthier, M.N. Bonnin-Verdal, G. Marchegay, L. Pinson-Gadais, C. Ducos, F.  
726 Richard-Forget, V. Atanasova-Penichon, Fungal biotransformation of chlorogenic and  
727 caffeic acids by *Fusarium graminearum*: New insights in the contribution of phenolic  
728 acids to resistance to deoxynivalenol accumulation in cereals, *Int. J. Food Microbiol.* 221  
729 (2016) 61–68. <https://doi.org/10.1016/j.ijfoodmicro.2016.01.005>.
- 730 [56] Y. Mikami, T. Yamazawa, Chlorogenic acid, a polyphenol in coffee, protects neurons  
731 against glutamate neurotoxicity, *Life Sci.* 139 (2015) 69–74.  
732 <https://doi.org/10.1016/j.lfs.2015.08.005>.
- 733 [57] F. Barahuie, D. Dorniani, B. Saifullah, S. Gothai, M.Z. Hussein, A.K. Pandurangan, P.  
734 Arulselvan, M.E. Norhaizan, Sustained release of anticancer agent phytic acid from its  
735 chitosan-coated magnetic nanoparticles for drug-delivery system, *Int. J. Nanomedicine.* 12  
736 (2017) 2361–2372. <https://doi.org/10.2147/IJN.S126245>.
- 737 [58] F. Barahuie, B. Saifullah, D. Dorniani, S. Fakurazi, G. Karthivashan, M.Z. Hussein, F.M.  
738 Elfghi, Graphene oxide as a nanocarrier for controlled release and targeted delivery of an  
739 anticancer active agent, chlorogenic acid, *Mater. Sci. Eng. C*. 74 (2017) 177–185.  
740 <https://doi.org/10.1016/j.msec.2016.11.114>.

- 741 [59] M. Narukawa, K. Kanbara, Y. Tominaga, Y. Aitani, K. Fukuda, T. Kodama, N.  
742 Murayama, Y. Nara, T. Arai, M. Konno, S. Kamisuki, F. Sugawara, M. Iwai, Y. Inoue,  
743 Chlorogenic acid facilitates root hair formation in lettuce seedlings, *Plant Cell Physiol.* 50  
744 (2009) 504–514. <https://doi.org/10.1093/pcp/pcp010>.
- 745 [60] M.J. Reigosa, E. Pazos-Malvido, Phytotoxic effects of 21 plant secondary metabolites on  
746 *Arabidopsis thaliana* germination and root growth, *J. Chem. Ecol.* 33 (2007) 1456–1466.  
747 <https://doi.org/10.1007/s10886-007-9318-x>.
- 748 [61] M. Villarino, P. Sandín-España, P. Melgarejo, A. De Cal, High chlorogenic and  
749 neochlorogenic acid levels in immature peaches reduce monilinia laxa infection by  
750 interfering with fungal melanin biosynthesis, *J. Agric. Food Chem.* 59 (2011) 3205–3213.  
751 <https://doi.org/10.1021/jf104251z>.
- 752 [62] K.S. Andrade, R.T. Gonálvez, M. Maraschin, R.M. Ribeiro-Do-Valle, J. Martínez, S.R.S.  
753 Ferreira, Supercritical fluid extraction from spent coffee grounds and coffee husks:  
754 Antioxidant activity and effect of operational variables on extract composition, *Talanta.*  
755 88 (2012) 544–552. <https://doi.org/10.1016/j.talanta.2011.11.031>.
- 756 [63] J.V.G. das NEVES, M.V. BORGES, D. de M. SILVA, C.X. dos S. LEITE, M.R.C.  
757 SANTOS, N.G.B. de LIMA, S.C. da S. LANNES, M.V. da SILVA, Total phenolic  
758 content and primary antioxidant capacity of aqueous extracts of coffee husk: chemical  
759 evaluation and beverage development, *Food Sci. Technol.* 39 (2019) 348–353.  
760 <https://doi.org/10.1590/fst.36018>.
- 761 [64] A.T.W.M. Hendriks, G. Zeeman, Pretreatments to enhance the digestibility of

- 762 lignocellulosic biomass, *Bioresour. Technol.* 100 (2009) 10–18.  
763 <https://doi.org/10.1016/j.biortech.2008.05.027>.
- 764 [65] L. Laureano-Perez, F. Teymouri, H. Alizadeh, B.E. Dale, Understanding factors that limit  
765 enzymatic hydrolysis of biomass: Characterization of pretreated corn stover, *Appl.*  
766 *Biochem. Biotechnol. - Part A Enzym. Eng. Biotechnol.* 124 (2005) 1081–1099.  
767 <https://doi.org/10.1385/ABAB:124:1-3:1081>.
- 768 [66] F. de Carvalho Oliveira, K. Srinivas, G.L. Helms, N.G. Isern, J.R. Cort, A.R. Gonçalves,  
769 B.K. Ahring, Characterization of coffee (*Coffea arabica*) husk lignin and degradation  
770 products obtained after oxygen and alkali addition, *Bioresour. Technol.* 257 (2018) 172–  
771 180. <https://doi.org/10.1016/j.biortech.2018.01.041>.
- 772 [67] B.E.L. Baêta, P.H. de M. Cordeiro, F. Passos, L.V.A. Gurgel, S.F. de Aquino, F. Fdz-  
773 Polanco, Steam explosion pretreatment improved the biomethanization of coffee husks,  
774 *Bioresour. Technol.* 245 (2017) 66–72. <https://doi.org/10.1016/j.biortech.2017.08.110>.
- 775 [68] R. Padmapriya, J.A. Tharian, T. Thirunalasundari, Coffee waste management-An  
776 overview, *Int J Curr Sci.* 9 (2013) 83–91.  
777 [http://www.currentsciencejournal.info/issuespdf/Coffee waste12.pdf](http://www.currentsciencejournal.info/issuespdf/Coffee%20waste12.pdf).
- 778 [69] A. Sampaio, G. Dragone, M. Vilanova, J.M. Oliveira, J.A. Teixeira, S.I. Mussatto,  
779 Production, chemical characterization, and sensory profile of a novel spirit elaborated  
780 from spent coffee ground, *LWT - Food Sci. Technol.* 54 (2013) 557–563.  
781 <https://doi.org/10.1016/j.lwt.2013.05.042>.
- 782 [70] A.S. Fernandes, F.V.C. Mello, S. Thode Filho, R.M. Carpes, J.G. Honório, M.R.C.



- 783 Marques, I. Felzenszwalb, E.R.A. Ferraz, Impacts of discarded coffee waste on human  
784 and environmental health, *Ecotoxicol. Environ. Saf.* 141 (2017) 30–36.  
785 <https://doi.org/10.1016/j.ecoenv.2017.03.011>.
- 786 [71] D. Brand, A. Pandey, S. Roussos, C.R. Soccol, Biological detoxification of coffee husk by  
787 filamentous fungi using a solid state fermentation system, *Enzyme Microb. Technol.* 27  
788 (2000) 127–133. [https://doi.org/10.1016/S0141-0229\(00\)00186-1](https://doi.org/10.1016/S0141-0229(00)00186-1).
- 789 [72] A. Pandey, C.R. Soccol, P. Nigam, V.T. Soccol, Biotechnological potential of agro-  
790 industrial residues. I: Sugarcane bagasse, *Bioresour. Technol.* 74 (2000) 69–80.  
791 [https://doi.org/10.1016/S0960-8524\(99\)00142-X](https://doi.org/10.1016/S0960-8524(99)00142-X).
- 792 [73] S. Gokulakrishnan, K. Chandraraj, S.N. Gummadi, Microbial and enzymatic methods for  
793 the removal of caffeine, *Enzyme Microb. Technol.* 37 (2005) 225–232.  
794 <https://doi.org/10.1016/j.enzmictec.2005.03.004>.
- 795 [74] B.A. Kihlman, Effects of caffeine on the genetic material, *Mutat. Res. Mol. Mech.*  
796 *Mutagen.* 26 (1974) 53–71.
- 797 [75] C. V. RAJ, S. DHALA, Effect of Naturally Occurring Xanthines on Bacteria. I.  
798 Antimicrobial, *Appl. Microbiol.* 13 (1965) 432–436.
- 799 [76] C.A. Woolfolk, Metabolism of N methylpurines by a *Pseudomonas putida* strain isolated  
800 by enrichment on caffeine as the sole source of carbon and nitrogen, *J. Bacteriol.* 123  
801 (1975) 1088–1106.
- 802 [77] G.D. Vogels, C. Van Der Drift, Degradation of purines and pyrimidines by

- 803 microorganisms, *Bacteriol. Rev.* 40 (1976) 403–468.
- 804 [78] T.J. Burr, A. Caesar, M.N. Schrollh, Beneficial plant bacteria, *CRC. Crit. Rev. Plant Sci.* 2  
805 (1984) 1–20. <https://doi.org/10.1080/07352688409382186>.
- 806 [79] D.M. Yamaoka-Yano, P. Mazzafera, Descafeinação da palha de café por bactérias, in:  
807 *Congr. Bras. Pesqui. Cafe.*, 1996: pp. 36–39.
- 808 [80] M.T. Moore, S.L. Greenway, J.L. Farris, B. Guerra, Assessing caffeine as an emerging  
809 environmental concern using conventional approaches, *Arch. Environ. Contam. Toxicol.*  
810 54 (2008) 31–35. <https://doi.org/10.1007/s00244-007-9059-4>.
- 811 [81] W. Acchar, E.J.V. Dultra, A.M. Segadães, Untreated coffee husk ashes used as flux in  
812 ceramic tiles, *Appl. Clay Sci.* 75–76 (2013) 141–147.  
813 <https://doi.org/10.1016/j.clay.2013.03.009>.
- 814 [82] K.N. Nisakorn Nuamsrinuan, Patcharin Naemchanthara, Pichet Limsuwan, Fabrication  
815 and Characterization of Particle Board from Coffee Husk Waste., *Appl. Mech. Mater.* 891  
816 (2019) 111–116.  
817 <https://doi.org/https://doi.org/10.4028/www.scientific.net/AMM.891.111>.
- 818 [83] M. Soares, P. Christen, A. Pandey, C.R. Soccol, Fruity flavour production by *Ceratocystis*  
819 *fimbriata* grown on coffee husk in solid-state fermentation, *Process Biochem.* 35 (2000)  
820 857–861. [https://doi.org/10.1016/S0032-9592\(99\)00144-2](https://doi.org/10.1016/S0032-9592(99)00144-2).
- 821 [84] H. Nyangito, Delivery of services to smallholder coffee farmers and impacts on  
822 production under liberalization in Kenya, Kenya Institute for Public Policy Research and

- 823 Analysis, 2000.
- 824 [85] M. Saenger, E.U. Hartge, J. Werther, T. Ogada, Z. Siagi, Combustion of coffee husks,  
825 Renew. Energy. 23 (2001) 103–121. [https://doi.org/10.1016/S0960-1481\(00\)00106-3](https://doi.org/10.1016/S0960-1481(00)00106-3).
- 826 [86] G.J. Miito, N. Banadda, A short review on the potential of coffee husk gasification for  
827 sustainable energy in Uganda, F1000Research. 6 (2017) 1–11.  
828 <https://doi.org/10.12688/f1000research.10969.1>.
- 829 [87] L. Wilson, G.R. John, C.F. Mhilu, W. Yang, W. Blasiak, Coffee husks gasification using  
830 high temperature air/steam agent, Fuel Process. Technol. 91 (2010) 1330–1337.  
831 <https://doi.org/10.1016/j.fuproc.2010.05.003>.
- 832 [88] B.C. Saha, M.A. Cotta, others, Fuel ethanol production from agricultural residues: current  
833 status and future prospects, J Biotechnol. 136 (2008) S285--S286.
- 834 [89] K.L. Somashekar, K.A.A. Appaiah, Coffee cherry husk - A potential feed stock for  
835 alcohol production, Int. J. Environ. Waste Manag. 11 (2013) 410–419.  
836 <https://doi.org/10.1504/IJEW.2013.054242>.
- 837 [90] M. Gonçalves, M.C. Guerreiro, L.C.A. de Oliveira, C.S. De Castro, A friendly  
838 environmental material: Iron oxide dispersed over activated carbon from coffee husk for  
839 organic pollutants removal, J. Environ. Manage. 127 (2013) 206–211.  
840 <https://doi.org/10.1016/j.jenvman.2013.05.017>.
- 841 [91] W.E. Oliveira, A.S. Franca, L.S. Oliveira, S.D. Rocha, Untreated coffee husks as  
842 biosorbents for the removal of heavy metals from aqueous solutions, J. Hazard. Mater.

- 843 152 (2008) 1073–1081. <https://doi.org/10.1016/j.jhazmat.2007.07.085>.
- 844 [92] I. Anastopoulos, M. Karamesouti, A.C. Mitropoulos, G.Z. Kyzas, A review for coffee  
845 adsorbents, *J. Mol. Liq.* 229 (2017) 555–565.  
846 <https://doi.org/10.1016/j.molliq.2016.12.096>.
- 847 [93] S. Berhe, D. Ayele, A. Tadesse, A. Mulu, Adsorption Efficiency of Coffee Husk for  
848 Removal of Lead ( II ) from Industrial Effluents : Equilibrium and kinetic study, *Int. J.*  
849 *Sci. Res. Publ.* 5 (2015) 1–8.
- 850 [94] B.G. Alhogbi, Potential of coffee husk biomass waste for the adsorption of Pb(II) ion from  
851 aqueous solutions, *Sustain. Chem. Pharm.* 6 (2017) 21–25.  
852 <https://doi.org/10.1016/j.scp.2017.06.004>.
- 853 [95] M. Hernández Rodriguez, J. Yperman, R. Carleer, J. Maggen, D. Daddi, G. Gryglewicz, B.  
854 Van der Bruggen, J. Falcón Hernández, A. Otero Calvis, Adsorption of Ni(II) on spent  
855 coffee and coffee husk based activated carbon, *J. Environ. Chem. Eng.* 6 (2018) 1161–  
856 1170. <https://doi.org/10.1016/j.jece.2017.12.045>.
- 857 [96] Mebrahtom Gebresemati, Nigus Gabbiye, O. Sahu, Sorption of cyanide from aqueous  
858 medium by coffee husk: Response surface methodology, *J. Appl. Res. Technol.* 15 (2017)  
859 27–35. <https://doi.org/10.1016/j.jart.2016.11.002>.
- 860 [97] L.S. Oliveira, A.S. Franca, T.M. Alves, S.D.F. Rocha, Evaluation of untreated coffee  
861 husks as potential biosorbents for treatment of dye contaminated waters, *J. Hazard. Mater.*  
862 155 (2008) 507–512. <https://doi.org/10.1016/j.jhazmat.2007.11.093>.

- 863 [98] M. Paredes-Laverde, J. Silva-Agreto, R.A. Torres-Palma, Removal of norfloxacin in  
864 deionized, municipal water and urine using rice (*Oryza sativa*) and coffee (*Coffea*  
865 *arabica*) husk wastes as natural adsorbents, J. Environ. Manage. 213 (2018) 98–108.  
866 <https://doi.org/10.1016/j.jenvman.2018.02.047>.
- 867 [99] V.S. Shankaranand, B.K. Lonsane, Coffee husk: an inexpensive substrate for production  
868 of citric acid by *Aspergillus niger* in a solid-state fermentation system, World J.  
869 Microbiol. Biotechnol. 10 (1994) 165–168. <https://doi.org/10.1007/BF00360879>.
- 870 [100] C.M.M. Machado, B.H. Oliveira, A. Pandey, C.R. Soccol, Coffee Husk as Substrate for  
871 the Production of Gibberellic Acid by Fermentation, in: T. Sera, C.R. Soccol, A. Pandey,  
872 S. Roussos (Eds.), Coffee Biotechnol. Qual. Proc. 3rd Int. Semin. Biotechnol. Coffee  
873 Agro-Industry, Londrina, Brazil, Springer Netherlands, Dordrecht, 2000: pp. 401–408.  
874 [https://doi.org/10.1007/978-94-017-1068-8\\_37](https://doi.org/10.1007/978-94-017-1068-8_37).
- 875 [101] P. Murthy, M. Naidu, Protease production by *Aspergillus oryzae* in solid-state  
876 fermentation utilizing coffee by-products, World Appl Sci J. 8 (2010) 199–205.
- 877 [102] V. Ballestin, G.A. Macedo, Effects of temperature, pH and additives on the activity of  
878 tannase produced by *Paecilomyces variotii*, Electron. J. Biotechnol. 10 (2007) 191–199.  
879 <https://doi.org/10.2225/vol10-issue2-fulltext-9>.
- 880 [103] I. Fki, N. Allouche, S. Sayadi, The use of polyphenolic extract, purified hydroxytyrosol  
881 and 3,4-dihydroxyphenyl acetic acid from olive mill wastewater for the stabilization of  
882 refined oils: A potential alternative to synthetic antioxidants, Food Chem. (2005).  
883 <https://doi.org/10.1016/j.foodchem.2004.09.014>.

- 884 [104] F. Figuerola, M.L. Hurtado, A.M. Estévez, I. Chiffelle, F. Asenjo, Fibre concentrates from  
885 apple pomace and citrus peel as potential fibre sources for food enrichment, *Food Chem.*  
886 (2005). <https://doi.org/10.1016/j.foodchem.2004.04.036>.
- 887 [105] L. Prosky, N.-G. Asp, I. Furda, J.W. Devries, T.F. Schweizer, B.F. Harland,  
888 Determination of Total Dietary Fiber in Foods, Food Products, and Total Diets:  
889 Interlaboratory Study, *J. Assoc. Off. Anal. Chem.* 67 (1984) 1044–1052.  
890 <https://doi.org/10.1093/jaoac/67.6.1044>.
- 891 [106] E.R.B.A. Prata, L.S. Oliveira, Fresh coffee husks as potential sources of anthocyanins,  
892 *LWT - Food Sci. Technol.* 40 (2007) 1555–1560.  
893 <https://doi.org/10.1016/j.lwt.2006.10.003>.
- 894 [107] M.R. Adams, J. Dougan, Biological management of coffee processing wastes, *Trop. Sci.*  
895 (1981). [http://agris.fao.org/agris-](http://agris.fao.org/agris-search/search.do?recordID=US201302154806#.Xbg0HYuLCho.mendeley)  
896 [search/search.do?recordID=US201302154806#.Xbg0HYuLCho.mendeley](http://agris.fao.org/agris-search/search.do?recordID=US201302154806#.Xbg0HYuLCho.mendeley) (accessed  
897 October 29, 2019).
- 898 [108] National Dairy Board, *Nutritive Value of Commonly Available Feeds and Fodders in*  
899 *India, 2012.*
- 900 [109] M. Catarina, M. Kasuya, J. Maria, C. Moura, C. Braga, P. Bento, Production of Selenium-  
901 Enriched Mushrooms in Coffee Husks and Use of This Colonized Residue, *Coffee Heal.*  
902 *Dis. Prev.* (2015) 301–309. <https://doi.org/10.1016/B978-0-12-409517-5.00033-4>.
- 903 [110] R.B.A. Rafael, M.L. Fernandez-Marcos, S. Cocco, M.L. Ruello, F. Fornasier, G. Corti,  
904 Benefits of Biochars and NPK Fertilizers for Soil Quality and Growth of Cowpea (*Vigna*

- 905        *unguiculata* L. Walp.) in an Acid Arenosol, Pedosphere. 29 (2019) 311–333.  
906        [https://doi.org/10.1016/S1002-0160\(19\)60805-2](https://doi.org/10.1016/S1002-0160(19)60805-2).
- 907 [111] R.B.A. Rafael, M.L. Fernandez-Marcos, S. Cocco, M.L. Ruello, D.C. Weindorf, V.  
908        Cardelli, G. Corti, Assessment of Potential Nutrient Release from Phosphate Rock and  
909        Dolostone for Application in Acid Soils, Pedosphere. 28 (2018) 44–58.  
910        [https://doi.org/10.1016/S1002-0160\(17\)60437-5](https://doi.org/10.1016/S1002-0160(17)60437-5).
- 911 [112] H. Pühringer, Effects of different biochar application rates on soil fertility and soil water  
912        retention in on-farm experiments on smallholder farms in Kenya, Lennart Hjelms Väg 9,  
913        SE-75007 Uppsala. (2016) 72. <http://stud.epsilon.slu.se>.
- 914 [113] B. Dume, G. Berecha, S. Tulu, Characterization of biochar produced at different  
915        temperatures and its effect on acidic nitosol of jimma, southwest ethiopia, Int. J. Soil Sci.  
916        10 (2015) 63–73. <https://doi.org/10.3923/ijss.2015.63.73>.
- 917 [114] C. Deal, C.E. Brewer, R.C. Brown, M.A.E. Okure, A. Amoding, Comparison of kiln-  
918        derived and gasifier-derived biochars as soil amendments in the humid tropics, Biomass  
919        and Bioenergy. 37 (2012) 161–168. <https://doi.org/10.1016/j.biombioe.2011.12.017>.
- 920 [115] M.N.S. Htet, N.N. Than, R.N. Soomro, X. Ya-Dong, H. Jiang-Bo, Comparison of  
921        nutrients composition , forage and silage yields of maize ( *Zea mays* L ), Sch. J. Agric.  
922        Vet. Sci. 3 (2016) 474–479. <https://doi.org/10.21276/sjavs.2016.3.7.5>.
- 923 [116] A.T. MATOS, v (Residues disposal in coffee post-processing), Borém, FM, Pós-Colheita  
924        Do Café (Coffee Post Process. Lavras Ed. UFLA. (2008) 161–201.

- 925 [117] E. Epstein, *The science of Composting*, Taylor & Francis, 1997.  
926 <https://doi.org/https://doi.org/10.1201/9780203736005>.
- 927 [118] M.J. López, M.A. Elorrieta, M.C. Vargas-García, F. Suárez-Estrella, J. Moreno, The effect  
928 of aeration on the biotransformation of lignocellulosic wastes by white-rot fungi,  
929 *Bioresour. Technol.* 81 (2002) 123–129. [https://doi.org/10.1016/S0960-8524\(01\)00112-2](https://doi.org/10.1016/S0960-8524(01)00112-2).
- 930 [119] N.A. Dzung, T.T. Dzung, V. Thi, P. Khanh, Evaluation of Coffee Husk Compost for  
931 Improving Soil Fertility and Sustainable Coffee Production in Rural Central Highland of  
932 Vietnam, 3 (2013) 77–82. <https://doi.org/10.5923/j.re.20130304.03>.
- 933 [120] D. Sekhar, P.B.P. Kumar, K.T. Rao, Effect of Coffee Husk Compost on Growth and Yield  
934 of Paddy, *J. Acad. Ind. Res.* 3 (2014) 195–197. [http://jairjp.com/SEPTEMBER 2014/09](http://jairjp.com/SEPTEMBER 2014/09 SEKHAR.pdf)  
935 [SEKHAR.pdf](http://jairjp.com/SEPTEMBER 2014/09 SEKHAR.pdf).
- 936 [121] C.C. Bidappa, Organic manure from coffee husk: comparison of technologies for organic  
937 manure from plantation wastes, *J. Planta. Cr.* 26 (1998) 120–126.
- 938 [122] B. Tuan, Efficiency of using coffee husk to applied for Robusta coffee in Central  
939 Highland, *J. Soil Sci.* 22 (2005) 10–15.
- 940 [123] G.H. Bitew, *Evaluation of On-farm Composting and Compost Quality at Ilala Gojo*  
941 *Welmera Woreda, Oromiya Region*, 2008.
- 942