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(Article begins on next page)

1 **Black Soldier Fly (*Hermetia illucens*) reared on roasted coffee by-product and *Schizochytrium***
2 ***sp.* as a sustainable terrestrial ingredient for aquafeeds production**

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19

20 **Key words:** insect meal; polyunsaturated fatty acids; circular economy; zebrafish; microbiota.

21

22 **Abstract**

23 Promoting circularity in the aquaculture sector through the conversion of great amount of organic by-
24 products produced on land is a valuable strategy for a further development of the aquaculture sector.

25 In this regard, insects represent a very promising example of bio-converting organisms; their
26 application in aquafeeds, however, still faces possible limitations because of their lack in
27 polyunsaturated fatty acids and the presence of chitin.

28 The aim of the present study was to apply circularity to Black Soldier Fly (BSF) (*Hermetia illucens*)
29 culture and to improve the insect's biomass fatty acid composition by culturing them on a land-
30 produced by-product (coffee silverskin) enriched with a 10% *Schizochytrium sp.* The insect biomass
31 was then used to formulate five fish diets containing 0, 25, 50, 75 and 100% of insect meal respect to
32 fish meal, respectively. Diets were used for a feeding trial during zebrafish (*Danio rerio*) larval
33 development (21 days) and a multidisciplinary approach including biometry, histology, gas
34 chromatography, spectroscopy (FTIR), microbiota analyses and molecular biology was applied to

35 better understand fish responses to the new diets. Results showed that the 50% substitution of fish
36 meal with insect meal represented the best compromise between ingredient sustainability and proper
37 fish growth and welfare. Fish fed with higher BSF inclusions (75 and 100%) showed a severe degree
38 of hepatic steatosis, microbiota modification, a higher lipid content (FTIR), fatty acid modification
39 and higher expression of both stress and immune response markers.

40

41 **1. Introduction**

42 Aquaculture is the fastest growing food production sector worldwide, and it is estimated that by 2030
43 62% of food-fish will come from aquaculture (FAO, 2018). The reason for this growth relies on
44 several factors, including wild capture fisheries decline, increase in global demand for seafood
45 products and the 9 billion people population expected on Earth by 2050, with the consequent doubling
46 of farmed production required to meet the mid-century demand for seafood (Gerland et al., 2014;
47 Guillen et al., 2018).

48 Because of the estimated increase in World's population, a significant rise in waste and by-products
49 production is expected. The EC Directive No. 2008/98, which establishes the order of priority in the
50 choice of by-products treatments (with their reuse as favoured option and their landfill disposal as
51 last option), will play a central role for further development of a European circular economy.

52 In consideration of this, aquaculture should be more responsible, sustainable, innovative, based on
53 the circular economy concept, and able to provide larger volumes of healthy food by using
54 environmentally friendly ingredients while promoting fish needs and welfare (Merino et al., 2012;
55 Tlusty and Thorsen, 2017; Stevens et al., 2018; Bohnes and Laurent, 2019).

56 For many years aquaculture has relied on the use of fish meal (FM) and fish oil (FO) as main
57 ingredients in aquafeeds (Tacon & Metian, 2008; Shepherd and Jackson, 2013). For its further
58 development, however, nutritious and sustainable ingredients must be identified and tested (Alhazzaa
59 et al., 2018; Sarker et al., 2018; Vargas et al., 2018).

60 Among such ingredients, several options have already been investigated and great attention has been
61 addressed to animal by-products (Processed Animals Proteins, PAPs), vegetable sources and
62 microalgae (Ayadi et al., 2012; Roy and Pal, 2014; Cardinaletti et al., 2018; Xu et al., 2019), but
63 unfortunately each of them has some downsides in the application for aquafeed formulation (Francis
64 et al., 2001; Naylor et al., 2009; Manceron et al., 2014; Bandara, 2018; Daniel, 2018).

65 Insects represent a very promising example of bio-converting organisms (Barroso et al., 2014; Henry
66 et al., 2015; Belghit et al., 2019).

67 Most insect species are farmed on land-produced by-products (van Huis, 2013; Čičková et al., 2015;
68 Webster et al., 2016; Spinelli et al., 2019;) and, in addition, farming of insects is sustainable in terms
69 of land use, water consumption and CO₂ production, because of their low environmental requirements
70 (Berggren et al., 2019; Smetana et al., 2019).

71 Land organic by-products are thus efficiently bio-converted in a highly nutritious biomass which, in
72 turn, can provide sustainable new ingredients for fish nutrition. Specifically, every year up to 200.000
73 tons of coffee silverskin are produced by the coffee industry as by-products (Murthy and Naidu, 2012;
74 Mussatto et al., 2011). Therefore, in a circular economy perspective, coffee silverskin may represent
75 a suitable substrate for insect production.

76 Among several insect species, the Black Soldier Fly (*Hermetia illucens*; BSF) has a promising role
77 for aquafeed production. From a nutritional point of view, the BSF accumulates good amounts of
78 proteins and lipids (307.5-588.0 g kg⁻¹ and 113.0-386.0 g kg⁻¹, respectively; Caligiani et al., 2018;
79 Nogales-Mérida et al., 2018). However, while the essential amino acid composition is approximately
80 similar to that of fish meal (Müller et al., 2017), the fatty acid (FA) profile is extremely different,
81 posing some limits in the full-fat BSF meal inclusion in aquafeeds (Zarantoniello et al., 2018;
82 Cardinaletti et al., 2019). In terms of fatty acid profile, the BSF is usually rich in saturated fatty acids
83 (SFAs) and poor in polyunsaturated (PUFAs) ones (Barroso et al., 2014), which are extremely
84 important for fish (Sargent et al., 1999). PUFAs deficiencies during fish farming can cause a general
85 decrease of fish health, poor growth, low feed efficiency, anaemia and high mortality (Tocher, 2010;

86 Olivotto et al., 2011; Piccinetti et al., 2012; Dumas et al., 2018). Because of their fatty acid profile,
87 insects are mainly used as protein source in fish nutrition and often undergo a defatting process which
88 represents an important cost in their manufacture (Jin et al., 2012; Li et al., 2015; Wang et al., 2017).
89 Therefore, in aquafeed production it is desirable to use full-fat insect meal. In this regard, it is known
90 that insects are able to modulate their fatty acid composition in relation to the growth substrate
91 (Komprda et al., 2013; Liland et al., 2017). Recently, some authors demonstrated that rearing BSF
92 larvae on an organic substrate containing proper amounts of omega-3 fatty acids was a suitable
93 procedure to improve the FAs profile of the final insect biomass (Barroso et al., 2017; St-Hilaire et
94 al., 2007b).

95 New ingredients to be introduced in aquafeeds must be carefully analysed, since it is well established
96 that different feed ingredients may have modulatory effects of on fish physiological responses and
97 gut microbiota (Li et al., 2019; Rimoldi et al., 2019). Besides zootechnical indexes, several molecular
98 markers involved in fish growth, stress response, lipid metabolism, appetite and immuno response
99 (Olivotto et al., 2002; Piccinetti et al., 2015; Cardinaletti et al., 2019; Vargas-Abúndez et al., 2019)
100 have been proposed as valid tools to precociously detect physiological responses in fish fed new diets
101 and represent an up-to-date and important approach.

102 In addition, it is already known that insects possess natural antibiotic properties, possibly modifying
103 fish microflora (Huyben et al., 2019; Terova et al., 2019), but knowledge concerning interactions
104 between insect-based diets, gut microbiota, and the aforementioned markers is still fragmentary and
105 incomplete.

106 The aim of the present study was to interconnect land and aquatic environment by: 1) recycling land-
107 produced organic by-products (coffee silverskin) to rear BSF larvae; 2) enriching the insects' growth
108 substrate with *Schizochytrium* sp. to improve their FAs profile; 3) producing highly nutritious full-fat
109 BSF prepupae meal; 4) testing the biological effects of diets including graded inclusions of BSF
110 prepupae meal in an aquatic experimental model organism, the zebrafish (*Danio rerio*) and 5)
111 applying a multidisciplinary approach integrating biometric, histological, gas chromatographic,

112 molecular, microbiological and spectroscopic analyses to better understand the physiological
113 responses of the fish. Zebrafish represents an extraordinary experimental model for aquaculture,
114 biomedical, developmental biology, genetics, toxicology studies, due to its high reproductive rate and
115 abundant information that has recently become available from genome sequencing (Lawrence, 2007;
116 Reed and Jennings, 2011). Particularly, zebrafish are used to generalize how several biological
117 processes take place in related organisms (like finfish species) and contribute to understand the
118 mechanisms involved in fish nutrition, welfare and growth, possibly providing useful information for
119 finfish production (Aleström et al., 2006; Dahm and Geisler, 2006; De-Santis and Jerry, 2007;
120 Johnston et al., 2008).

121

122 **2. Materials and methods**

123 *2.1. Ethics*

124 All procedures involving animals were conducted in line with the Italian legislation on experimental
125 animals and were approved by the Ethics Committee of the Università Politecnica delle Marche
126 (Ancona, Italy) and the Italian Ministry of Health (626/2018-PR). Optimal rearing conditions (see
127 further section for details) were applied throughout the study, and all efforts were made to minimize
128 animal suffering by using an anaesthetic (MS222; Sigma Aldrich, Saint Louis, Missouri, USA).

129

130 *2.2. Insect feeding substrate preparation*

131 The main component of the insect feeding substrate consisted of coffee silverskin, a coffee industry
132 by-product provided by Saccaria Caffè S.R.L. (Marina di Montemarçiano, Ancona, Italy). Coffee
133 by-product (moisture 44%) was collected and grinded in an Ariete 1769 food processor (De Longhi
134 Appliances Srl, Italy) to a 0.4 ± 2 mm particle size before the feeding substrate preparation. The insect
135 diet was formulated including a 10% (w/w) of *Schizochytrium* sp to the coffee by-product (for details,
136 please see Truzzi et al., in press). The freeze-dried *Schizochytrium* sp. was provided by AlghItaly

137 Società Agricola S.R.L. (Sommacampagna, VR, Italy). Insect feeding substrate was added with
138 distilled water to reach a final moisture of ~70% (Makkar et al., 2014).

139

140 *2.3. Insect rearing*

141 Insects were reared in a climatic chamber at a 27 ± 1 °C temperature, relative humidity of $65\pm 5\%$
142 (Spranghers et al., 2017), in continuous darkness. Six days old larvae [purchased from Smart Bugs
143 s.s. Ponzano Veneto (TV), Italy] were hand counted and divided in groups of 640 larvae per replicate
144 ($n=65$) for a total of 41,600 specimens. Each replicate consisted of a plastic box (57x38x16cm)
145 screened with fine-mesh cotton gauze and covered with a lid provided with 90 ventilation holes of
146 0.05cm Ø (Spranghers et al., 2017). Larvae were reared at a density of $0.3/\text{cm}^2$ (Barragan-Fonseca et
147 al., 2018). Each larva was provided with a feeding rate of 100 mg/day (Diener et al., 2009) that was
148 prepared and added once a week (448 g for each box). Insects were visually inspected every day and,
149 when prepupae were identified by the change in tegument colour from white to black (May, 1961),
150 they were collected, washed, dried and stored at -80 °C.

151

152 *2.4. Fish diets production*

153 Full-fat BSF prepupae were freeze dried, grinded with Retsch Centrifugal Grinding Mill ZM 1000
154 (Retsch GmbH, Haan, Germany) and used to prepare the experimental diets. A control diet (Hi0)
155 containing FM, wheat gluten, pea protein concentrates and FO as major ingredients, was prepared
156 according to a commercially available standard diet for zebrafish (Zebrafeed, Sparos ltd, Portugal).
157 The experimental diets were isonitrogenous (50%) and isolipidic (13%). Insect-based diets were
158 prepared by including graded levels of insect meal (25%, 50%, 75%, and 100%, referred to as Hi25
159 and Hi50, Hi75 and Hi100, respectively) in the Hi0 formulation. The conventional vegetable
160 ingredients (pea protein concentrate and wheat gluten) used to formulate the experimental diets were
161 maintained approximatively at constant (0.7:1 w:w) ratio in all diets. In summary, all the grounded
162 ingredients (0.5 mm) and fish oil were thoroughly blended (Kenwood kMix KMX53 stand Mixer;

163 Kenwood, De Longhi S.p.a., Treviso, Italy) for 20 min and then water was added to the mixture to
 164 obtain an appropriate consistency for pelleting. Pellets were obtained by using a 1 mm die meat
 165 grinder and dried at 40 °C for 48–72 h. The obtained diets were then grinded and stored in vacuum
 166 bags at –20 °C until used. Feed samples were analyzed for moisture (AOAC #950.46), crude protein,
 167 CP (AOAC #976.05), ash (AOAC #920.153) and ether extract (EE; AOAC #991.36) contents
 168 according to AOAC 2006.

169 The total lipid fraction of the test diets was extracted using chloroform-methanol (2:1 v:v) (Merck
 170 KGaA, Darmstadt, Germany) mixture according to Folch et al., 1957. Diet formulation and proximate
 171 composition are shown in Table 1.

172 TABLE 1. INGREDIENTS (g Kg⁻¹) AND PROXIMATE COMPOSITION (g 100 g⁻¹) OF THE
 173 EXPERIMENTAL DIETS USED IN THIS STUDY.

	Hi0 (Control)	Hi25	Hi50	Hi75	Hi100
Ingredients (g/kg)					
Fish meal ¹	470	400	250	110	-
Vegetable mix ²	220	230	298	385	440
Hi meal	-	150	275	350	460
Wheat flour ³	198	172	120	110	72
Fish oil	80	51	25	10	-
Soy lecithin	8	8	8	11	4
Mineral and Vitamin supplements [§]	14	14	14	14	14
Binder	10	10	10	10	10
Proximate composition (%)					
Dry Matter	97.08 ± 0.06	95.78 ± 0.13	94.93 ± 0.05	93.63 ± 0.05	92.70 ± 0.04
Crude protein, CP	51.57 ± 0.13	50.75 ± 2.57	50.39 ± 0.28	51.23 ± 1.49	50.50 ± 3.15
Ether extract, EE	14.38 ± 0.64	13.10 ± 0.42	12.93 ± 0.38	13.24 ± 0.46	12.99 ± 0.51
NFE	21.32 ± 0.34	20.82 ± 1.00	20.64 ± 0.55	19.03 ± 0.67	18.47 ± 1.26
Ash	9.81 ± 0.25	11.11 ± 0.01	10.97 ± 0.00	10.13 ± 0.06	10.74 ± 0.13

174 ¹ Raw ingredients kindly supplied by Skretting Italia; ² Vegetable mix (pea protein concentrate : wheat gluten, 0.7:1
 175 w/w) Lombarda trading srl, Casalbuttano & Uniti (CR, Italy) and Sacchetto spa (Torino, Italy); ³ Consorzio Agrario
 176 (Pordenone, Italy); [§] Mineral and Vitamin supplement composition (% mix): CaHPO₄·2H₂O, 78.9; MgO, 2.725;

177 KCl, 0.005; NaCl, 17.65; FeCO₃, 0.335; ZnSO₄.H₂O, 0.197; MnSO₄.H₂O, 0.094; CuSO₄.5H₂O, 0.027; Na₂SeO₃,
178 0.067; thiamine hydrochloride (vitamin B1), 0.16; riboflavin (vitamin B2), 0.39; pyridoxine hydrochloride (vitamin
179 B6), 0.21; cyanocobalamin (vitamin B12), 0.21; niacin (vitamin PP or B3), 2.12; calcium pantothenate, 0.63; folic
180 acid, 0.10; biotin (vitamin H), 1.05; myo-inositol (vitamin B7), 3.15; stay C Roche (vitamin C), 4.51; tocopherol
181 (vitamin E), 3.15; menadione (vitamin K3), 0.24; retinol (vitamin A 2500 UI kg⁻¹ diet), 0.026; cholecalciferol
182 (vitamin D3 2400 UI kg⁻¹ diet), 0.05; choline chloride, 83.99; * Values reported as mean of triplicate analyses; ⁶
183 n.d.: not determined

184

185 *2.5. Fish*

186 Zebrafish AB embryos were maintained for 48h in a Tecniplast system (Varese, Italy) in the following
187 conditions: 28 °C temperature, pH 7.0, NO₂ and NH₃ concentrations < 0.01 mg/L, NO₃ concentration
188 < 10 mg/L, and photoperiod 12L/12D, respectively (Randazzo et al., 2017). After this period,
189 embryos were gently collected, counted under a stereomicroscope (Leica Wild M3B, Leica
190 Microsystems, Nussloch, Germany) and randomly divided in five experimental groups (in triplicate)
191 according to the five test diets.

192

193 *2.6. Experimental design*

194 Zebrafish larvae were maintained in fifteen 20L tanks to set up the five experimental dietary
195 treatments; each experimental group was composed of 1500 larvae (500 larvae per tank). The water
196 in the larval tanks had the same chemical-physical characteristics of the parent's tank and was gently
197 replaced 10 times a day by a dripping system (Olivotto et al., 2004). The sides of each tank were
198 covered with black panels to reduce light reflection. All tanks were siphoned 30 min after feeding
199 (twice a day) to remove possible feed excess and dead larvae. The required larvae were sampled 20
200 days after fertilization (dpf), euthanized with a lethal dose of MS222 (1g/L) and properly stored for
201 further analyses.

202

203 *2.7. Feeding schedule*

204 Starting from 5 dpf to 20 dpf, zebrafish larvae were fed as follows: Control group: larvae fed on diet
205 0% insect meal (Hi0 diet); Group A: larvae fed on the diet including 25% BSF full-fat prepupae meal

206 (Hi25 diet); Group B: larvae fed on the diet including 50% of BSF full-fat prepupae meal (Hi50 diet);
207 Group C: larvae fed on the diet including 75% BSF full-fat prepupae meal (Hi75 diet); Group D:
208 larvae fed on the diet including 100% BSF full-fat prepupae meal (Hi100 diet). Zebrafish larvae were
209 fed on the experimental diets (2% body weight, BW; 100-250 μm size) twice a day and, in addition,
210 from 5 to 10 dpf, all groups were fed (one feeding in the morning) on the rotifer *Brachionus plicatilis*
211 (5 ind/mL) according to Lawrence et al. (2012) and Piccinetti et al. (2014).

212

213 2.8. Biometry

214 Five zebrafish larvae (15 per dietary group) were randomly collected from the different tanks of each
215 experimental group at hatching (3 dpf) and at the end of the experiment (20 dpf). The standard length
216 was determined using a sliding calliper (Measy 2000 Typ 5921, Swiss; precision: 0.1 mm) and the
217 wet weight using an OHAUS Explorer (OHAUS Europe GmbH, Greifensee, Switzerland) analytical
218 balance (precision: 0.1 mg) according to Zarantoniello et al. (2018). At 3 dpf, wet weight was
219 measured on pools of five larvae in triplicate. For each experimental group, specific growth rate
220 (SGR) was calculated as follows: $\text{SGR}\% = [(\ln W_f - \ln W_i)/t] \times 100$, where W_f is the final wet weight,
221 W_i , the initial wet weight, and t , the number of days (17). Survival rate in all experimental groups
222 was about 85%.

223

224 2.9. Fatty acid composition

225 The experimental diets and fish larvae samples were analyzed for fatty acid composition. Samples
226 were minced and homogenized (homogenizer MZ 4110, DCG Eltronic, Monza, Italy), and larvae
227 were also freeze-dried (Edwards EF4, Crawley, Sussex, England). Aliquots of 200 mg of each sample
228 were added with 100 μl of Internal Standard (methyl ester of nonadecanoic acid, 99.6%, Dr.
229 Ehrenstorfer GmbH, Germany), and extracted overnight following the method of Folch et al., 1957.
230 Analyses were carried out on three aliquots *per* sample. All lipid extracts were evaporated under
231 laminar flow inert gas (N_2) until constant weight and re-suspended in 0.5 ml of n-epthane.

232 Fatty acid methyl esters (FAMES) were prepared according to Canonico et al., 2016 using methyl
233 ester of nonadecanoic acid (19:0; Dr. Ehrenstorfer GmbH, Augsburg, Germany) as internal standard.
234 FAMES were determined using an Agilent-6890 GC System (Milano, Italy) coupled to an Agilent-
235 5973N quadrupole Mass Selective Detector (MSD) (Milano, Italy). A CPS ANALITICA CC-wax-
236 MS (30 m × 0.25 mm ID, 0.25 µm film thickness) capillary column was used to separate FAMES.
237 Instrumental conditions for the studied matrices were set up according to Truzzi et al. (2017, 2018).
238 For each analysed aliquot of sample, at least three runs were performed on the GCMS.

239

240 *2.10. Histology*

241 Nine zebrafish larvae per dietary group (3 larvae per tank, at 20 dpf), randomly collected from the
242 different tanks, were fixed by immersion in Bouin's solution (Sigma-Aldrich, Milano, Italy) and then
243 stored at 4°C for 24h. Larvae were washed three times in ethanol (70%) for ten minutes and preserved
244 in the same ethanol solution. Larvae were then dehydrated in increasing ethanol solutions (80, 95 and
245 100%), washed in xylene (Bio-Optica, Milano, Italy) and embedded in paraffin (Bio-Optica).
246 Solidified paraffin blocks were cut with a microtome (Leica RM2125 RTS, Nussloch, Germany) and
247 5 µm sections were stained with Mayer hematoxylin and eosin Y (Sigma-Aldrich, Milano, Italy).
248 Sections were observed using a Zeiss Axio Imager.A2 (Oberkochen, Germany) microscope in order
249 to study the hepatic parenchyma and intestine morphology. Images were acquired by mean of a
250 combined color digital camera Axiocam 503 (Zeiss, Oberkochen, Germany). Moreover, to ascertain
251 the extent of fat accumulation in liver, a quantitative analysis was performed on a significant number
252 of histological sections from each experimental group in triplicate (n=9). No-n-evaluable areas, such
253 as blood vessels were not considered. The percentage of fat fraction (PFF) on the total tissue areas
254 was calculated using the ImageJ software setting a homogeneous threshold value.

255

256

257 *2.11. FTIR analysis*

258 Fifteen zebrafish larvae per dietary group (5 per tank) were randomly collected at 20 dpf. Samples
259 were minced, homogenized (homogenizer MZ 4110, DCG Eltronic, Monza, Italy) and freeze-dried
260 (Edwards EF4, Crawley, Sussex, England) for FTIR analysis. For each group, five aliquots of 5 mg
261 each were analysed. FTIR analysis was performed using a Spectrum GX1 Spectrometer equipped with
262 a U-ATR accessory and a diamond/SeZn crystal (Perkin Elmer, Waltham, Massachusetts, USA).
263 Measurements were carried out in reflectance in the MIR region from 4000 to 800 cm^{-1} (spectral
264 resolution 4 cm^{-1}). Each spectrum was the result of 64 scans. Before each sample acquisition, a
265 background spectrum was collected on the clean surface of the crystal. Raw IR spectra were converted
266 in absorbance, two-points baseline linear fitted in the 4000-800 cm^{-1} spectral range and vector
267 normalized in the same interval (OPUS 7.1 software package).

268 For all experimental groups, the average absorbance spectra were calculated together with their standard
269 deviation spectra (average absorbance spectrum \pm standard deviation spectra) and analysed to identify
270 the most featuring IR peaks (in terms of position/wavenumbers). Then, average absorbance spectra and
271 their standard deviation spectra were curve-fitted in the 3050-2800 cm^{-1} and 1790-900 cm^{-1} regions
272 upon two-points baseline correction and vector normalization. A Gaussian algorithm was adopted, and
273 the number and position of the underlying peaks was defined by second derivative analysis of the
274 spectra (GRAMS/AI 9.1, Galactic Industries, Inc., Salem, NH). In the 3050-2800 cm^{-1} region, the
275 following underlying peaks were identified: $\sim 3010 \text{ cm}^{-1}$ (=CH moieties in unsaturated lipid alkyl
276 chains); $\sim 2959 \text{ cm}^{-1}$ and $\sim 2872 \text{ cm}^{-1}$ (CH_3 groups in lipid alkyl chains); $\sim 2925 \text{ cm}^{-1}$ and $\sim 2854 \text{ cm}^{-1}$
277 (CH_2 groups in lipid alkyl chains). In the 1790-900 cm^{-1} region, the following underlying peaks were
278 identified: $\sim 1744 \text{ cm}^{-1}$ (C=O moiety in lipids and fatty acids); ~ 1639 and $\sim 1536 \text{ cm}^{-1}$ (respectively
279 Amide I and II bands of proteins); $\sim 1457 \text{ cm}^{-1}$ (proteins side chains); $\sim 1390 \text{ cm}^{-1}$ (COO^- groups in
280 aspartate and glutamate amino acids); $\sim 1234 \text{ cm}^{-1}$ (collagen); $\sim 1157 \text{ cm}^{-1}$ (glycosylated compounds);
281 $\sim 1080 \text{ cm}^{-1}$ (phosphate groups), and $\sim 1055 \text{ cm}^{-1}$ (mucin). The integrated areas of the most relevant
282 peaks were calculated and used to evaluate the following band area ratios: LIP/TBM (representing the

283 overall amount of lipids), 1744/LIP (representing the amount of fatty acids compared to lipids),
284 2928/LIP (representing the amount of saturated fatty acids with respect to lipids), and 3010/LIP
285 (representing the amount of unsaturated fatty acids compared to lipids); PRT/TBM (representing the
286 overall amount of proteins), 1234/PRT (representing the amount of collagen compared to proteins), and
287 1055/PRT (representing the amount of mucin compared to proteins). TBM was the sum of the
288 integrated areas of all peaks in the 3050-2800 cm^{-1} and 1790-900 cm^{-1} regions; LIP was the sum of the
289 integrated areas of all peaks in the 3050-2800 cm^{-1} region, while PRT was the sum of the integrated
290 areas of the bands at 1639 and 1536 cm^{-1} .

291 2.12. Microbiome

292 *RNA extraction and cDNA synthesis.* Prior to analysis, zebrafish larvae (60 larvae per dietary group)
293 were disinfected by washing in 50 mL of ethanol (70%) on a laboratory shaker (VDRL Stirrer with
294 thermostatic cupola, ASAL s.r.l, Milan, Italy) at 150 rpm for 1 min at room temperature, in order to
295 analyse only the gut microbiome. These samples were thus identified as G (gut) Hi0, 25, 50, 75 and
296 100 for these analyses. After discharging the ethanol, the samples were rinsed in two additional
297 washing steps in 50 mL of sterile deionized water. Subsequently, 31.5 mL of sterile peptone water
298 (peptone, 1 g/L) was added to each sample, which was then homogenized in a Stomacher apparatus
299 (400 Circulator, International PBI, Milan, Italy) for 3 min at 260 rpm. Then, 1.5 mL of each tenfold
300 diluted (10-1 dilution) homogenate were centrifuged at 14,000 rpm for 10 min, the supernatants were
301 discarded, the obtained cell pellets covered with RNA later Stabilization Solution (Ambion, Foster
302 City, CA, USA) and stored at -80°C until use. The Quick-RNA Fungal/Bacterial Microprep kit (Zymo
303 Research, CA, USA) was used for the extraction of total microbial RNA from the cell pellets
304 following the manufacturer's instructions. The extracted RNAs were checked for quantity, purity and
305 integrity as previously described by Garofalo et al. (2017). Moreover, the extracts were amplified
306 using the PCR universal prokaryotic primers 27f and 1495r (Weisburg et al., 1991) to exclude the
307 presence of bacterial DNA contamination. Five μL of each RNA sample were reverse- transcribed in

308 cDNA using the SensiFAST cDNA Synthesis Kit for RT-qPCR (Bioline, London, UK) following the
309 manufacturer's instructions.

310

311 *16S rRNA amplicon target sequencing.* cDNA was used as template in the PCR amplifying the V3-
312 V4 region of the 16S rRNA gene using the primers and protocols described by Klindworth et al.
313 (2013).

314 PCR amplicons were cleaned using the Agencourt AMPure kit (Beckman Coulter, Milan, Italy) and
315 tagged using the Nextera XT Index Kit (Illumina Inc. San Diego, CA) according to the manufacturer's
316 instructions. After the 2nd clean-up step, amplicons were quantified using a QUBIT dsDNA Assay kit
317 and an equimolar amount of amplicons from different samples were pooled. The library was
318 denatured with 0.2 N NaOH, diluted to 12 pM, and combined with 20% (vol/vol) denatured 12 pM
319 PhiX, prepared according to Illumina guidelines. The sequencing was performed with a MiSeq
320 Illumina instrument (Illumina) with V3 chemistry and generated 250 bp paired-end reads according
321 to the manufacturer's instructions.

322

323 *2.13. Molecular analyses*

324 *RNA extraction and cDNA synthesis.* Total RNA extraction from 5 zebrafish larvae from each tank
325 (15 larvae per dietary group) was optimized using the RNazol RT reagent (Sigma-Aldrich, R4533)
326 according to Piccinetti et al. (2013). The total RNA extracted was eluted in 40 µl of RNase-free water
327 (Qiagen). The final RNA concentration was determined using a NanoPhotometer P-Class (Implen,
328 München, Germany). RNA integrity was verified by GelRedTM staining of 28S and 18S ribosomal
329 RNA bands on 1% agarose gel. RNA was stored at -80°C until use. Finally, 2 µg of RNA were used
330 for cDNA synthesis, using the High Capacity cDNA Reverse Transcription Kit (Bio-Rad, Milan,
331 Italy) following the manufacturer's instructions.

332

333 *Real-Time PCR*. PCRs were performed with SYBER green method in an iQ5 iCycler thermal cycler
334 (Bio-Rad Laboratories) following Vargas et al. (2018).
335 Relative quantification of the expression of genes involved in fish growth (*igf1*, *igf2a* and *mstnb*),
336 stress response (*nr3c1* and *hsp70.1*), long-chain polyunsaturated fatty acids biosynthesis (*elovl2*,
337 *elovl5* and *fads2*), appetite response (*ghrl*, *npy*, *cnr1* and *lepa*), immune response (*il1b*, *il10* and *tnfa*)
338 and enzymatic hydrolysis of chitin (*chia.2* and *chia.3*) was performed. Actin-related protein 2/3
339 complex, subunit 1A (*arpc1a*) and ribosomal protein, large, 13 (*rpl13*) were used as internal standards
340 in each sample in order to standardize the results by eliminating variation in mRNA and cDNA
341 quantity and quality. No amplification products were observed in negative controls and no primer-
342 dimer formations were observed in the control templates. Amplification products were sequenced,
343 and homology was verified. The data obtained were analysed using the iQ5 optical system software
344 version 2.0 (Bio-Rad) including GeneEx Macro iQ5 Conversion and genex Macro iQ5 files. Primer
345 sequences were designed using Primer3 (210 v. 0.4.0) starting from zebrafish sequences available in
346 ZFIN Primer sequences used were reported in Table 2.

347
348 TABLE 2. PRIMER SEQUENCES AND THE ZEBRAFISH INFORMATION NETWORK (ZFIN)
349 USED IN THIS STUDY

<i>Gene</i>	<i>Forward primer (5'- 3')</i>	<i>Reverse primer (5'- 3')</i>	<i>ZFIN ID</i>
<i>igf1</i>	5'-GGCAAATCTCCACGATCTCTAC-3'	5'-CGGTTTCTCTTGTCTCTCTCAG-3'	ZDB-GENE-010607-2
<i>igf2a</i>	5'-GAGTCCCATTCCATTCTGTTG-3'	5'-GTGGATTGGGGTTTGTATGTG-3'	ZDB-GENE-991111-3
<i>mstnb</i>	5'-GGACTGGACTGCGATGAG-3'	5'-GATGGGTGTGGGGATACTTC-3'	ZDB-GENE-990415-165
<i>nr3c1</i>	5'-AGACCTTGGTCCCCTTCACT-3'	5'-CGCCTTTAATCATGGGAGAA-3'	ZDB-GENE-050522-503
<i>hsp70.1</i>	5'-TGTTTCAGTTCTCTGCCGTTG-3'	5'-AAAGCACTGAGGGACGCTAA-3'	ZDB-GENE-990415-91
<i>elovl2</i>	5'-CACTGGACGAAGTTGGTGAA-3'	5'-GTTGAGGACACACCACCAGA-3'	ZDB-GENE-060421-5612
<i>elovl5</i>	5'-TGGATGGGACCGAAATACAT-3'	5'-GTCTCCTCCACTGTGGGTGT-3'	ZDB-GENE-040407-2
<i>fads2</i>	5'-CATCACGCTAAACCCAACA-3'	5'-GGGAGGACCAATGAAGAAGA-3'	ZDB-GENE-011212-1
<i>ghrl</i>	5'-CAGCATGTTTCTGCTCCTGTG-3'	5'TCTTCTGCCACTCTTGGTG-3'	ZDB-GENE-070622-2
<i>npy</i>	5'-GTCTGCTTGGGACTCTCAC-3'	5'CGGGACTCTGTTTCACCAAT-3'	ZDB-GENE-980526-438
<i>cnr1</i>	5'-AGCAAAAAGGAGCAACAGGCA-3'	5'GTTGGTCTGGTACTTTCACCTTGAC-3'	ZDB-GENE-040312-3
<i>lepa</i>	5'-CTCCAGTGACGAAGGCAACTT-3'	5'GGGAAGGAGCCGGAAATGT-3'	ZDB-GENE-081001-1

<i>illb</i>	5'-GCTGGGGATGTGGACTTC-3'	5'-GTGGATTGGGGTTTGATGTG-3'	ZDB-GENE-040702-2
<i>ill0</i>	5'-ATTTGTGGAGGGCTTTCCTT-3'	5'AGAGCTGTTGGCAGAATGGT-3'	ZDB-GENE-051111-1
<i>infa</i>	5'-TTGTGGTGGGGTTTGATG-3'	5'-TTGGGGCATTTTATTTTGTAAG-3'	ZDB-GENE-050317-1
<i>chia.2</i>	5'-GGTGCTCTGCCACCTTGCCTT-3'	5'-GGCATGGTTGATCATGGCGAAAGC-3'	ZDB-GENE-040426-2014
<i>chia.3</i>	5'-TCGACCCTTACCTTTGCACACACCT-3'	5'-ACACCATGATGGAGAACTGTGCCGA-3'	ZDB-GENE-040426-2891
<i>arpc1a</i>	5'-CTGAACATCTCGCCCTTCTC-3'	5'-TAGCCGATCTGCAGACACAC-3'	ZDB-GENE-040116-1
<i>rpl13</i>	5'-TCTGGAGGACTGTAAGAGGTATGC-3'	5'-AGACGCACAATCTTGAGAGCAG-3'	ZDB-GENE-031007-1

350

351 2.14. Statistical analysis

352 All data (except for microbiome) were analyzed by one-way ANOVA, with diet as the explanatory
353 variable. All ANOVA tests were followed by Tukey's post-hoc tests. The statistical software package
354 Prism5 (GraphPad Software) was used. Significance was set at $p < 0.05$ and all results are presented
355 as mean \pm SD. For microbiome analyses, paired-end reads were first merged using the FLASH
356 software (Magoč and Salzberg, 2011). Joint reads were quality filtered (at Phred $<$ Q20) by QIIME
357 1.9.0 software (Caporaso et al., 2010) and the pipeline recently described (Osimani et al., 2019).
358 Briefly, the USEARCH software version 8.1 (Edgar et al., 2011) was used for chimera filtering and
359 clean sequences were clustered into Operational Taxonomic Units (OTUs) at 97% of similarity by
360 UCLUST algorithms. Centroids sequences of each cluster were used for taxonomic assignment using
361 the Greengenes 16S rRNA gene database. OTU tables were rarefied at 44412 sequences. The OTU
362 table displays the higher taxonomy resolution that was reached.

363

364 3. Results

365 3.1. Biometry

366 The increasing inclusion levels of BSF full-fat prepupae meal resulted in a statistically significant
367 ($p < 0.05$) increase in the larval specific growth rate. In particular, no significant differences ($p > 0.05$)
368 were detected between Control ($25.4 \pm 0.7\%$) and Hi25 group ($25.7 \pm 1.0\%$), while Hi50, Hi75 and

369 Hi100 groups (27.6 ± 0.5 , 27.8 ± 0.4 , 28.4 ± 0.3 %, respectively) showed significantly ($p<0.05$) higher
370 values compared to both Control and Hi25 ones.

371

372 *3.2. Fatty acid content and composition*

373 *Diets.* The FAs classes percentages of the five experimental diets are presented in Figure 2a. Insect-
374 based diets showed significantly higher ($p<0.05$) percentages of SFAs (40.9 ± 0.7 , 40.0 ± 2.0 , 35.9 ± 0.7
375 and 37.6 ± 2.8 % for Hi25, Hi50, Hi75 and Hi100 diets, respectively) compared to Control diet
376 ($27.8\pm 1.3\%$). Considering mono-unsaturated fatty acids (MUFAs), all insect-based diets showed
377 significantly lower percentages ($p<0.05$) compared to Control (24.7 ± 0.6 , 19.8 ± 0.3 , 19.0 ± 0.9 ,
378 21.5 ± 0.2 and 20.0 ± 1.0 % for Control, Hi25, Hi50, Hi75 and Hi100, respectively). Finally, insect-
379 based diets showed significantly ($p<0.05$) lower (39.3 ± 1.0 , 41.0 ± 1.0 , 42.6 ± 0.3 and 42.2 ± 3.2 % for
380 Hi25, Hi50, Hi75 and Hi100, respectively) percentages of PUFAs compared to Control diet (47.4 ± 1.4
381 %). In addition, increasing inclusion levels of BSF full-fat prepupae meal in the diets resulted in a
382 significant decrease ($p<0.05$) of n3 percentages (from $38.8\pm 1.4\%$ for Control diet to 11.1 ± 3.1 % for
383 Hi100 diet) and a parallel significant ($p<0.05$) increase in n6 percentages (from 8.6 ± 0.1 for Control
384 diet to 31.3 ± 0.9 % for Hi100 diet; Fig. 2a). Consequently, the n6/n3 ratio showed significant
385 differences ($p<0.05$) among experimental diets, increasing from Control diet (0.22 ± 0.01) to Hi100
386 diet (2.8 ± 0.2) diets (Fig. 2b). Finally, considering the insect-based diets, the higher was the BSF meal
387 dietary inclusion, the higher was the n9 content (10.7 ± 0.2 , 12.1 ± 0.7 , 14.6 ± 0.2 and 15.2 ± 0.7 % for
388 Hi25, Hi50, Hi75 and Hi100, respectively). The control diet ($13.9\pm 0.3\%$) showed an intermediate n9
389 content between Hi50 and Hi75 diets.

390 *Zebrafish larvae.* Figure 2c illustrates the FAs classes percentages of zebrafish larvae fed on the
391 different diets. The FA classes of zebrafish larvae fed on the different diets was deeply influenced by
392 the BSF meal dietary inclusion. In particular, SFAs percentage increased with the inclusion of insect
393 meal in the diets, while both MUFAs and PUFAs generally decreased with the increasing BSF

394 inclusion levels in the diets (Fig. 2c). Specifically, the higher was the dietary BSF meal inclusion
 395 level, the lower was the n3 and the higher was the n6 percentage detected. Considering the n6/n3 ratio
 396 (Fig. 2d), no significant differences ($p>0.05$) were detected among Control, Hi25 and Hi50 groups
 397 (0.5 ± 0.1 , 0.5 ± 0.1 and 0.6 ± 0.1 , respectively), while Hi75 and Hi100 (0.8 ± 0.1 and 1.0 ± 0.1 ,
 398 respectively) showed a significantly ($p<0.05$) higher value compared to the other experimental
 399 groups.

400 Finally, in terms of n9 percentage, only Hi25 and Hi50 groups (17.7 ± 0.4 and 17.9 ± 0.3 %,
 401 respectively) showed significantly ($p<0.05$) higher values compared to Control group (17.3 ± 0.2 %).

402 Table 3 shows the FA composition of total lipids of zebrafish larvae. The FA profile of larvae reared
 403 on the Control diet was characterized by high percentages of 16:0, 18:1n9, 22:6n3 (docosahexaenoic
 404 acid, DHA), and 20:5n3 (eicosapentaenoic acid, EPA) fatty acids, followed by 18:2n6, 16:1n7 and
 405 18:0. The increasing inclusion levels of BSF meal in the diets triggered some changes in the FA
 406 composition of the larvae and the major changes are reported here.

407 A substantial (up to ~30-folds) increase in the lauric acid (12:0) and a significant decrease in EPA
 408 (20:5n3) percentage was detected in fish fed diets with increasing BSF meal inclusion levels. In terms
 409 of DHA, a similar, but milder trend was observed respect to EPA. Because of this the DHA/EPA ratio
 410 significantly increased with the increasing BSF meal inclusion levels in the diets (Tab. 3).

411
 412 **TABLE 3: FATTY ACID COMPOSITION (AS % OF TOTAL FAS) OF ZEBRAFISH LARVAE FED DIETS**
 413 **WHERE FM WAS REPLACED WITH 25, 50, 75 AND 100 % OF BSF MEAL (Hi25, Hi50, Hi75 AND Hi100**
 414 **GROUPS).**

Zebrafish larvae					
	CTRL	Hi25	Hi50	Hi75	Hi100
10:0	0.02±0.01	0.05±0.01	0.10±0.01	0.08±0.01	0.09±0.02
12:0	0.20±0.03 ^a	2.6±0.1 ^b	4.5±0.1 ^c	4.7±0.4 ^c	5.7±0.4 ^d
13:0	0.06±0.01	0.06±0.01	0.07±0.01	0.06±0.01	0.07±0.01
14:0	4.0±0.1 ^{a,b}	4.0±0.1 ^a	4.2±0.1 ^b	3.9±0.2 ^a	4.3±0.2 ^b
15:0	0.86±0.04	0.80±0.03	0.82±0.01	0.79±0.02	0.84±0.04
16:0	14.7±1.0 ^c	14.0±0.7 ^{a,b}	13.8±0.5 ^a	14.6±0.1 ^{b,c}	15.5±0.9 ^d
16:1n9	1.2±0.1 ^a	1.3±0.1 ^b	1.3±0.1 ^c	1.5±0.1 ^d	1.5±0.1 ^d
16:1n7	8.7±0.6 ^c	9.0±0.3 ^c	8.3±0.1 ^b	7.6±0.4 ^a	7.9±0.1 ^a

17:0	1.2±0.1 ^b	1.1±0.1 ^a	1.1±0.1 ^a	1.1±0.1 ^a	1.2±0.1 ^b
18:0	6.3±0.2 ^c	5.5±0.1 ^a	5.8±0.2 ^b	6.3±0.4 ^c	6.7±0.4 ^d
18:1n9	14.7±0.1 ^a	15.3±0.3 ^b	15.7±0.7 ^c	15.1±0.3 ^b	15.3±0.1 ^b
18:1n7	5.2±0.1 ^d	4.9±0.2 ^c	4.6±0.2 ^b	4.3±0.1 ^a	4.5±0.2 ^b
18:2n6	9.6±0.1 ^a	10.6±0.2 ^b	11.0±0.2 ^c	12.6±0.3 ^d	12.7±0.7 ^d
18:3n6	0.43±0.01	0.41±0.02	0.50±0.01	0.47±0.03	0.53±0.03
18:3n3	3.7±0.1 ^c	3.3±0.2 ^b	3.5±0.2 ^c	2.7±0.2 ^a	2.8±0.3 ^a
20:0	0.22±0.01	0.18±0.01	0.19±0.01	0.23±0.02	0.21±0.01
20:1n9	0.90±0.05	0.75±0.01	0.63±0.02	0.48±0.02	0.39±0.02
20:2n6	0.29±0.02	0.29±0.01	0.31±0.01	0.35±0.02	0.38±0.02
20:3n6	0.46±0.04 ^a	0.52±0.02 ^b	0.70±0.04 ^c	0.98±0.06 ^d	1.1±0.1 ^e
20:4n6	2.3±0.2 ^a	2.3±0.1 ^a	2.4±0.1 ^b	3.2±0.3 ^c	3.5±0.1 ^d
20:3n3	0.21±0.03	0.19±0.01	0.21±0.01	0.17±0.01	0.17±0.02
20:5n3	10.2±0.2 ^c	8.7±0.7 ^d	7.2±0.3 ^c	4.6±0.1 ^b	3.5±0.2 ^a
22:1n9	0.31±0.02	0.28±0.01	0.18±0.02	0.12±0.01	0.04±0.01
22:6n3	14.0±1.3 ^c	13.9±1.2 ^c	12.9±0.8 ^b	14.0±1.3 ^c	11.2±1.0 ^a
24:1n9	0.18±0.02	0.09±0.01	0.08±0.01	0.05±0.01	0.03±0.01
DHA/EPA	1.4±0.1 ^a	1.6±0.2 ^b	1.8±0.1 ^b	3.0±0.3 ^c	3.2±0.2 ^c

415 Means within rows bearing different letters are significantly different ($p < 0.05$). Statistical analysis was performed only
416 for FAs > 1%. FAs with a percentage < 1% were excluded from any statistical analyses because their concentrations were
417 close to the limit of detection.
418

419

420

421

422

423 3.3. Histology

424 Histological analyses were performed in order to detect possible inflammatory events in the intestine

425 and to evaluate lipid accumulation or steatosis in the liver. The intestine mucosa appeared unaltered

426 and did not show any appreciable inflammatory influx in all experimental groups and fish fed on

427 BFS-based diets showed a normal intestinal morphology, comparable to Control (Fig. 3a-j).

428 Conversely, results evidenced a variable degree of lipid accumulation in the liver of the experimental

429 groups (Fig. 3k-o). The most appreciable visual differences were observed in the Hi75 and Hi100

430 groups. Indeed, while all experimental groups presented a modest fat liver parenchima, Hi75 and

431 Hi100 showed a severe degree of steatosis with swollen hepatocytes and abundant intracytoplasmic
432 lipid accumulation (Fig. 3n,o). These results were confirmed by the statistical quantification of the
433 fat percentage fraction (PPF) on liver sections that showed a significant increase in response to dietary
434 treatments (Fig. 4). In particular, Hi50 showed a significant increase in PPF (48.7 ± 2.4) compared to
435 Control (41.7 ± 2.1) and Hi25 (42.5 ± 1.2) groups. However, the highest values in PPF were detected
436 in Hi75 and Hi100 with a significant difference (58.7 ± 0.9 and 60.6 ± 1.5 respectively) compared to
437 the other groups.

438

439 *3.4. FTIR analysis*

440 The average absorbance spectra of Control, Hi25, Hi50, Hi75 and Hi100 larval groups are shown in
441 Figure 5. The analysis of IR spectra allowed to distinguish the vibrational modes of lipids, proteins,
442 carbohydrates and phosphates (see Materials and Methods section) (Vargas et al., 2018).

443 In terms of biochemical composition of zebrafish larvae samples belonging to the different dietary
444 groups, the following considerations can be drawn: (i) statistically significant higher amounts of
445 overall lipids (LIP/TBM, Fig. 6a) were detected in Hi50, Hi75 and Hi100 zebrafish larvae compared
446 to Control and Hi25 ones; (ii) in all zebrafish larvae fed on diets with different inclusion levels of
447 BSF meal, a significant increase in saturated fatty acids (1744/LIP and 2925/LIP, Fig. 6b and 6c) and
448 a significant decrease in unsaturated ones (3010/LIP, Fig. 6d) were observed; (iii) in terms of protein
449 composition, no statistically significant changes were detected in the overall amount of proteins
450 (PRT/TBM, Fig. 7a) among the experimental groups; (iv) a slight but statistically significant decrease
451 of collagen (1234/PRT, Fig. 7b) was observed in all zebrafish larvae fed on diets including BFS meal,
452 and (v) a higher amount of mucin (1055/PRT, Fig. 7c) was found in Hi50, Hi75 and Hi100 zebrafish
453 larvae.

454 *3.5. Microbiome*

455 A total of 247,654 reads passed the filters applied through QIIME, with an average value of 49,530
456 reads/sample, and a mean sequence length of 464 bp. The Good's coverage indicated also satisfactory

457 coverage for all samples (>99%) (Supplementary Table 1A). Alpha-diversity indicated the highest
458 number of OTUs after 50% of BSF inclusion if compared with control and with 25% of inclusion.
459 Beta diversity calculation based on weighted and on unweight UniFrac distance matrix showed a
460 clear separation of the control samples if compared with those including BSF (Fig. 8). Furthermore,
461 50 and 75% of BSF meal inclusion in the diets showed a similar effect on microbial composition.
462 As shown in Figure 9, the main OTUs shared among the data sets were *Cetobacterium* that reached
463 50% of the relative abundance in sample GHi50 and about 30% in the other samples. *Vibrio* and
464 *Mycoplasmataceae* decreased with the increasing inclusion of BSF meal in the diets; *Flavobacterium*
465 and *Plesiomonas* were present at very low abundance in GHi100 sample, whereas in the other samples
466 the same two genera showed a remarkable presence. Finally, among other bacteria, *Aeromonadaceae*,
467 *Ochrobactrum* and *Tetrathiobacter* were also detected.

468

469 3.6. Real-time PCR results

470 Real-time PCR analyses were performed on genes involved in fish growth (*igf1*, *igf2a* and *mstnb*),
471 stress response (*hsp70.1* and *nr3c1*), long-chain polyunsaturated fatty acids biosynthesis (*elovl2*,
472 *elovl5* and *fads2*), appetite (*ghrl*, *npy*, *cnr1* and *lepa*), immune response (*il1b*, *il10* and *tnfa*) and
473 enzymatic hydrolysis of chitin (*chia.2* and *chia.3*).

474 *Growth factors.* Higher mean levels in the expression of the *igf1* and *igf2a* genes were detected in all
475 experimental groups fed on BSF-based diets compared to Control (with the exception of *igf1* gene
476 expression in Hi25 group). In particular, the Hi75 and Hi100 groups showed significantly ($p<0.05$)
477 higher levels in comparison to the Hi25 group in terms of *igf1* gene expression, and to Control in
478 terms of *igf2a* gene expression. Only the Hi50 and Hi100 groups showed significantly ($p<0.05$) higher
479 levels of expression of the *mstn* gene than Control (Fig. 10c).

480 *Stress response.* Considering stress markers (*nr3c1* and *hsp70.1*; Fig. 10d,e), all groups fed on BSF-
481 based diets showed a significantly ($p<0.05$) higher gene expression compared to Control (with the
482 exception of *nr3c1* gene expression in Hi25 group). Furthermore, no significant differences ($p>0.05$)

483 were evident among the Hi50, Hi75 and Hi100 groups, while the Hi25 group showed significantly
484 ($p < 0.05$) lower values compared to groups fed on diets with higher BSF meal inclusion level (with
485 the exception of *nr3c1* gene expression in Hi50 group).

486

487 *Lipid metabolism.* The highest BSF meal inclusion (Hi100) caused the highest expression of the genes
488 involved in long-chain polyunsaturated fatty acid elongation (*elovl2* and *elovl5*) and desaturation
489 (*fads2*). As illustrated in Figure 10f-h, the Hi100 group presented the highest gene expression
490 ($p < 0.05$) compared to all the other experimental groups. More specifically, no significant differences
491 ($p > 0.05$) in the expression of the *elovl2* gene were detected between Control and Hi25 group (Fig.
492 10f), while both Hi50 and Hi75 had a significantly ($p < 0.05$) higher expression than Control. No
493 significant differences ($p > 0.05$) in the expression of the *elovl5* and *fads2* genes (Fig. 10g,h) were
494 observed among the Control, Hi25, Hi50 and Hi75 groups.

495

496 *Appetite.* As shown in Figure 10i,l, the higher was the BSF meal inclusion in the diets, the higher was
497 the *ghrl* and *npv* gene expression. However, no significant differences ($p > 0.05$) were observed
498 between Control and Hi25 in terms of *ghrl* gene expression and among Control, Hi25 and Hi50 in
499 terms of *npv* gene expression. Figure 10m shows *cnr1* gene expression. No BSF meal dose
500 dependency was observed in the expression of this specific gene, since only Hi25 and Hi50 groups
501 evidenced significantly ($p < 0.05$) higher values than Control. , Groups fed on the highest BSF
502 inclusion (Hi75 and Hi100) showed significantly ($p < 0.05$) higher gene expression of the *lepa* gene
503 (Fig. 10n) compared to the other groups, while no significant differences ($p < 0.05$) were detected
504 among Control, Hi25 and Hi50 groups.

505

506 *Immune response.* Considering genes involved in the immune response, higher BSF meal dietary
507 inclusions resulted in a significantly ($p < 0.05$) higher gene expression compared to Control.
508 Specifically, no significant differences ($p > 0.05$) were evident in the expression of the *illb* gene (Fig

509 10o) among the Control, Hi25 and Hi50 groups, while the Hi75 and Hi100 groups showed
510 significantly ($p < 0.05$) higher values compared to the other experimental groups. Similarly, the Hi50,
511 Hi75 and Hi100 groups showed a significantly ($p < 0.05$) higher expression of the *ill0* gene (Fig 10p)
512 compared to the Control and Hi25 groups, which did not differ significantly from each other ($p > 0.05$).
513 Finally, as reported in Figure 10q, the increasing levels of inclusion of BSF meal in the diets resulted
514 in a statistically significant ($p < 0.05$) dose-dependent increase in *tnfa* gene expression.

515

516 *Chitinases*. All the experimental groups fed on BSF-based diets showed an increase in chitinases gene
517 expression (Fig. 10r,s). Specifically, no significant differences ($p > 0.05$) were evident in the
518 expression of the *chia.2* and *chia.3* genes among all the experimental groups, with the exception of
519 the Hi100 group for *chia.2* and both Hi75 and Hi100 groups for *chia.3*, which showed a significantly
520 ($p < 0.05$) higher gene expression than Control.

521

522 **4. Discussion**

523 The use of insects in aquafeed still faces possible limitations because of their lack in PUFAs (Barroso
524 et al., 2017) and presence of chitin (Kroeckel et al., 2012; Xiao et al., 2018).

525 Insects are usually farmed on vegetable organic by-products, and previous studies showed that the
526 final insect biomass had a very low PUFAs and a high SFA content (Vargas et al., 2018; Zarantoniello
527 et al., 2018). However, it is now well established that the quantity and quality of FAs in insects can
528 be modified by the growth substrate (Barroso et al., 2014; Spranghers et al., 2017; St-Hilaire et al.,
529 2007a). Specifically, in the present study we demonstrated that the addition of a 10% (w/w) of
530 *Schyzochitrium sp.* to the growth substrate is an efficient approach to increase the PUFAs content of
531 the final insect biomass.

532 This “enrichment” procedure of the insect biomass caused a progressive increase in PUFAs content
533 in the Hi25, 50, 75 and 100 diets, respectively, highlighting that the enrichment method used is
534 appropriate and represents, in this sense, an interesting and innovative approach on how food by-

535 products produced on land (coffee silverskin) can be enriched and bio-converted into a valuable
536 biomass for aquafeed production (Meneguz et al., 2018; Pinotti et al., 2019).

537 Compared to previous insect-based diets already tested on zebrafish (Vargas et al., 2018;
538 Zarantonello et al., 2019), the diets tested in this study showed good performances on fish growth,
539 since higher BSF meal inclusion levels in the experimental diets resulted in higher larval SGR%. This
540 result was supported by the analysed molecular markers of growth but is in contrast with previous
541 studies reporting that BSF meal inclusion levels higher than 40% often resulted in negative effects on
542 fish growth and welfare (Sánchez-Muros et al., 2014; Barragan-Fonseca et al., 2018; Secci et al.,
543 2019). In particular, previous studies evidenced that such high inclusion levels affected both intestine
544 and liver integrity and, thus, dietary nutrients absorption (Henry et al., 2015; Li et al., 2017;
545 Cardinaletti et al., 2019).

546 The different diets tested in the present study also affected larval fish FA composition. However,
547 differences among FA classes were less evident in zebrafish larvae compared to those detected in the
548 diets. In fact, as a freshwater species, zebrafish are able to convert shorter-chain FAs in highly
549 unsaturated ones through the elongation and desaturation pathways (Tocher, 2010). This was clearly
550 documented in the present study by a higher *elovl2*, *elovl5* and *fads* gene expression in all the
551 experimental groups fed on BSF meal-based diets (with emphasis on the Hi100 group).

552 Gas chromatographic results were also confirmed and integrated by FTIR analyses which provided
553 data about the macromolecular composition of the analysed biological samples (Giorgini et al., 2018).
554 Specifically, FTIR analyses showed that there were no differences in the total amount of proteins
555 (PRT/TBM ratio) (with the exception of collagen (1234/PRT ratio) that slightly decreased) among
556 the experimental groups, highlighting that BSF meal is a valuable protein source for aquafeeds (Al-
557 Qazzaz and Ismail, 2016; Lock et al., 2016; Nogales-Mérida et al., 2018).

558 Concerning lipids, the same analyses revealed that the increase of BSF meal percentage in the diets
559 was associated with a general increase in total larval lipid content. Additionally, analysing specific

560 wavelengths, FTIR confirmed the gas chromatographic results, evidencing an overall increase in
561 saturated FA (1744/LIP and 2925/LIP ratios) and a decrease in unsaturated (3010/LIP ratio).

562 Molecular markers related to the appetite stimulus were fully supported by the biometric results. Fish
563 food intake is regulated by specific regions in the brain that interpret and integrate positive
564 (orexigenic) and negative (anorexigenic) signals derived from the hypothalamic area
565 (neurohormones) and from the body periphery (Copeland et al., 2011; Sobrino Crespo et al., 2014).

566 Specifically, the orexigenic signals *ghrl* and *npy* analysed in the present study, showed a dose-
567 dependent gene expression increase related to the BSF meal inclusion levels in the diets (the higher
568 the BSF meal inclusion level, the higher their gene expression), while the *cnrl* gene expression was
569 always higher in all the larval groups fed on BSF-based diets. Conversely, the results obtained from
570 the anorexigenic signal *lepa* was not obvious. Leptin is usually involved in the inhibition of
571 orexigenic pathways and in the stimulation of anorexigenic (Piccinetti et al., 2010; Volkoff, 2006);
572 in the present study, however, an opposite activity was evidenced.

573 This anomalous pattern could be explained by the FTIR analyses that revealed an increasing total
574 lipid content (LIP/TBM ratio) in larvae fed on diets with increasing BSF meal levels. In fact, previous
575 studies, performed both on mammals and fish species, demonstrated a positive correlation between
576 leptin levels and amount of adipose tissue (Chisada et al., 2014; Park and Ahima, 2015; Li et al.,
577 2016).

578 Alternatively, an interconnection between leptin and proinflammatory cytokines exist (Lafrance et
579 al., 2010). Leptin is known to enhance the production of proinflammatory cytokines, as well as
580 proinflammatory cytokines are able to trigger leptin release (Carlton et al., 2012). In the present study,
581 the increasing *lepa* gene expression observed in larvae fed on insect-based diets might have promoted
582 the immune-response observed in the same experimental groups. However, it should be mentioned
583 that all insect-based diets contain chitin (Borrelli et al., 2017; Gasco et al., 2018), a molecule which
584 still has a controversial role in aquafeeds. Indeed, some studies reported a beneficial modulatory role
585 of chitin on fish immune system and microbiota (Ringø et al., 2012; Zhang et al., 2012; Zhou et al.,

586 2013; Bruni et al., 2018; Henry et al., 2018), while others showed that, especially at high inclusion
587 levels, chitin may induce inflammation of the intestinal tract and a reduction in nutrient assimilation
588 (Kroeckel et al., 2012; Magalhães et al., 2017; Su et al., 2017; Xiao et al., 2018). In support of this
589 conclusion, Hi50, Hi75 and Hi100 larvae showed a significant increase of mucin (1055/PRT ratio),
590 possibly associated to a higher intestine lubrication necessary for a proper intestinal transit of these
591 diets.

592 In the present study, no specific inflammatory events were detected through the histological analysis
593 of the intestine samples, suggesting: 1) a possible digestion of chitin by zebrafish larvae through
594 specific chitinases (*chia.2* and *chia.3*); 2) a possible intestinal anti-inflammatory role of lauric acid
595 (C12). This fatty acid, which in the present study increased its dietary amount with the increasing
596 BSF meal inclusion in the diets, has been demonstrated to have beneficial effects on fish gut's welfare
597 by mitigating inflammatory conditions (Aleström et al., 2006; Dahm and Geisler, 2006; De-Santis
598 and Jerry, 2007; Zarantoniello et al., 2019).

599 The histological analysis of the liver showed a lipid accumulation in the hepatic parenchyma in all
600 analysed samples, regardless of the dietary treatment. Specifically, the Hi75 and Hi100 groups
601 showed a severe level of hepatic steatosis, a pathological condition that has previously been related
602 to a high n-6/n-3 ratio (Di Minno et al., 2012; Leamy et al., 2013; Zarantoniello et al., 2018, 2019).
603 This hepatic disorder is probably the cause of the higher gene expression of the stress markers (*nr3c1*
604 and *hsp70.1*). Finally, insects are known to possess natural bioactive molecules that are known to
605 modulate fish microbiota (Huyben et al., 2019; Terova et al., 2019) and, therefore, the bacterial
606 community is expected to impact host metabolism and health status.

607 Although autochthonous bacterial communities were dominated by the same phyla regardless of the
608 diet, the present study showed that bacterial populations are dynamic and can be modulated by dietary
609 inclusion of insect meal.

610 Some bacteria were almost present in all the samples analysed, while others were mainly modulated
611 by the insect-based diets. Among the ubiquitous species, the massive presence of *Cetobacterium* in

612 all gut samples suggested that graded BSF inclusion levels in the diets did not influence the presence
613 of such a fish associated microorganism. In fact, *Cetobacterium* has already been found as core genus
614 in the gut of zebrafish with beneficial effect on fish health because of its ability to produce cobalamin
615 (vitamin B12) (Roeselers et al., 2011; Merrifield et al., 2013; Ghanbari et al., 2015; Earley et al.,
616 2018;). Mycoplasmataceae were also dominant in the present study, and have already been found
617 among the dominant microorganisms in fish intestines (e.g. rainbow trout), with a possible beneficial
618 action on host health by producing lactic acid with antibacterial effect (Rimoldi et al., 2019). Finally,
619 Aeromonadaceae were also detected in all analysed gut samples, irrespective of the diet used. This
620 bacterial family has already been found as naturally associated with the gut of fish or insects
621 (Grabowski and Klein, 2017; Udayangani et al., 2017; Rimoldi et al., 2019), suggesting a contribution
622 of BSF-based diets in the occurrence of such a bacterial family in the gut of the analysed zebrafish.
623 BSF meal inclusion in the diets modified the presence of some bacteria. Specifically, the presence of
624 *Vibrio* was negatively influenced by the addition of increasing graded levels of BSF meal in the diets.
625 As reported by Brugman et al., (2015), in the zebrafish intestine T lymphocytes can control the
626 outgrowth of *Vibrio* species. As shown by the Real Time PCR analyses, the supply of BSF-based
627 diets stimulated immune-response related genes that might be implied in *Vibrio* species control.
628 However, further research is needed to better understand the involvement of BSF in this aspect.
629 Finally, both *Ochrobactrum* and *Tetrathiobacter* were detected in the samples GHi100. The
630 occurrence of *Ochrobactrum* was already reported in zebrafish gut (Cantas et al., 2012) as well as in
631 the microbiota of insects as coleoptera and lepidoptera (Grabowski and Klein, 2017), while
632 *Tetrathiobacter* has already been isolated from the midgut of ticks (Li et al., 2014). These data suggest
633 that the inclusion of BSF meal in fish diets can contribute to modify the fish microbiota with a specific
634 possible involvement in the allochthonous microbiota modification.
635 As a general remark about gut microbiota, bacterial diversity is considered a positive indicator of gut
636 health while a reduced diversity is often associated to dysbiosis and risk of disease in fish (Terova et
637 al., 2019). In the present study, the PCA analyses revealed a difference between the Hi0-25 and Hi50-

638 75-100 groups, thus suggesting a possible influence of BFS-based diets on the zebrafish gut
639 microbiota, to be further investigated.

640

641

642 **5. Conclusion**

643 In order to meet aquafeed requirements, BSF biomass should be enriched in PUFAs. The present
644 study showed that the addition of a 10% (W/W) *Schizochytrium* sp to the growth substrate is a valid
645 method to achieve this objective. The present study evidenced that a 50% BSF meal inclusion level
646 in the diet sustains a better fish growth and does not have any major negative effects on the fish.
647 Higher inclusion levels affect larval liver histology and induce a general increase in lipid
648 accumulation and stress response. This novel approach represents an interesting example of how, in
649 the long term, a circular economy applied to the aquaculture sector may sustain animal's welfare and
650 encourage sustainability and competitiveness.

651

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656 **7. References**

- 657 Al-Qazzaz, M.F., Ismail, D.B., 2016. Insect meal as a source of protein in animal diet. *Anim. Nutr.*
658 *Feed Technol.* 16, 527–547. <https://doi.org/10.5958/0974-181X.2016.00038.X>
- 659 Aleström, P., Holter, J.L., Nourizadeh-Lillabadi, R., 2006. Zebrafish in functional genomics and
660 aquatic biomedicine. *Trends Biotechnol.* <https://doi.org/10.1016/j.tibtech.2005.11.004>
- 661 Alhazzaa, R., Nichols, P.D., Carter, C.G., 2018. Sustainable alternatives to dietary fish oil in tropical
662 fish aquaculture. *Rev. Aquac.* <https://doi.org/10.1111/raq.12287>
- 663 Ayadi, F.Y., Rosentrater, K.A., Muthukumar, K., 2012. Alternative Protein Sources for Aquaculture
664 Feeds. *J. Aquac. Feed Sci. Nutr.* 4, 1–26. <https://doi.org/10.3923/joafsnu.2012.1.26>
- 665 Bandara, T., 2018. Alternative feed ingredients in aquaculture: Opportunities and challenges. *J.*
666 *Entomol. Zool. Stud.* 6, 3087–3094.
- 667 Barragan-Fonseca, K.B., Dicke, M., van Loon, J.J.A., 2018. Influence of larval density and dietary

- 668 nutrient concentration on performance, body protein, and fat contents of black soldier fly larvae
669 (*Hermetia illucens*). *Entomol. Exp. Appl.* <https://doi.org/10.1111/eea.12716>
- 670 Barroso, F.G., de Haro, C., Sánchez-Muros, M.J., Venegas, E., Martínez-Sánchez, A., Pérez-Bañón,
671 C., 2014. The potential of various insect species for use as food for fish. *Aquaculture* 422–423,
672 193–201. <https://doi.org/10.1016/j.aquaculture.2013.12.024>
- 673 Barroso, F.G., Sánchez-Muros, M.J., Segura, M., Morote, E., Torres, A., Ramos, R., Guil, J.L., 2017.
674 Insects as food: Enrichment of larvae of *Hermetia illucens* with omega 3 fatty acids by means
675 of dietary modifications. *J. Food Compos. Anal.* <https://doi.org/10.1016/j.jfca.2017.04.008>
- 676 Belghit, I., Liland, N.S., Gjesdal, P., Biancarosa, I., Menchetti, E., Li, Y., Waagbø, R., Krogdahl, Å.,
677 Lock, E.J., 2019. Black soldier fly larvae meal can replace fish meal in diets of sea-water phase
678 Atlantic salmon (*Salmo salar*). *Aquaculture*. <https://doi.org/10.1016/j.aquaculture.2018.12.032>
- 679 Berggren, Å., Jansson, A., Low, M., 2019. Approaching Ecological Sustainability in the Emerging
680 Insects-as-Food Industry. *Trends Ecol. Evol.* <https://doi.org/10.1016/j.tree.2018.11.005>
- 681 Bohnes, F.A., Laurent, A., 2019. LCA of aquaculture systems: methodological issues and potential
682 improvements. *Int. J. Life Cycle Assess.* <https://doi.org/10.1007/s11367-018-1517-x>
- 683 Borrelli, L., Coretti, L., Dipineto, L., Bovera, F., Menna, F., Chiariotti, L., Nizza, A., Lembo, F.,
684 Fioretti, A., 2017. Insect-based diet, a promising nutritional source, modulates gut microbiota
685 composition and SCFAs production in laying hens. *Sci. Rep.* 7, 16269.
686 <https://doi.org/10.1038/s41598-017-16560-6>
- 687 Brugman, S., Schneeberger, K., Witte, M., Klein, M.R., van den Bogert, B., Boekhorst, J.,
688 Timmerman, H.M., Boes, M.L., Kleerebezem, M., Nieuwenhuis, E.E.S., 2015. T lymphocytes
689 control microbial composition by regulating the abundance of *Vibrio* in the zebrafish gut. *Gut*
690 *Microbes*. <https://doi.org/10.4161/19490976.2014.972228>
- 691 Bruni, L., Pastorelli, R., Viti, C., Gasco, L., Parisi, G., 2018. Characterisation of the intestinal
692 microbial communities of rainbow trout (*Oncorhynchus mykiss*) fed with *Hermetia illucens*
693 (black soldier fly) partially defatted larva meal as partial dietary protein source. *Aquaculture*
694 487, 56–63. <https://doi.org/10.1016/J.AQUACULTURE.2018.01.006>
- 695 Caligiani, A., Marseglia, A., Leni, G., Baldassarre, S., Maistrello, L., Dossena, A., Sforza, S., 2018.
696 Composition of black soldier fly prepupae and systematic approaches for extraction and
697 fractionation of proteins, lipids and chitin. *Food Res. Int.*
698 <https://doi.org/10.1016/j.foodres.2017.12.012>
- 699 Canonico, L., Ashoor, S., Taccari, M., Comitini, F., Antonucci, M., Truzzi, C., Scarponi, G., Ciani,
700 M., 2016. Conversion of raw glycerol to microbial lipids by new *Metschnikowia* and *Yarrowia*
701 *lipolytica* strains. *Ann. Microbiol.* <https://doi.org/10.1007/s13213-016-1228-0>
- 702 Cantas, L., Sørby, J.R.T., Aleström, P., Sørum, H., 2012. Culturable Gut Microbiota Diversity in
703 Zebrafish. *Zebrafish*. <https://doi.org/10.1089/zeb.2011.0712>
- 704 Caporaso, J.G., Kuczynski, J., Stombaugh, J., Bittinger, K., Bushman, F.D., Costello, E.K., Fierer,
705 N., Peña, A.G., Goodrich, J.K., Gordon, J.I., Huttley, G.A., Kelley, S.T., Knights, D., Koenig,
706 J.E., Ley, R.E., Lozupone, C.A., McDonald, D., Muegge, B.D., Pirrung, M., Reeder, J.,
707 Sevinsky, J.R., Turnbaugh, P.J., Walters, W.A., Widmann, J., Yatsunenko, T., Zaneveld, J.,
708 Knight, R., 2010. QIIME allows analysis of high-throughput community sequencing data. *Nat.*
709 *Methods*. <https://doi.org/10.1038/nmeth.f.303>
- 710 Cardinaletti, G., Messina, M., Bruno, M., Tulli, F., Poli, B.M., Giorgi, G., Chini-Zittelli, G., Tredici,

- 711 M., Tibaldi, E., 2018. Effects of graded levels of a blend of *Tisochrysis lutea* and *Tetraselmis*
712 *suecica* dried biomass on growth and muscle tissue composition of European sea bass
713 (*Dicentrarchus labrax*) fed diets low in fish meal and oil. *Aquaculture* 485, 173–182.
714 <https://doi.org/10.1016/J.AQUACULTURE.2017.11.049>
- 715 Cardinaletti, G., Randazzo, B., Messina, M., Zarantoniello, M., Giorgini, E., Zimbelli, A., Bruni, L.,
716 Parisi, G., Olivotto, I., Tulli, F., 2019. Effects of Graded Dietary Inclusion Level of Full-Fat
717 *Hermetia illucens* Prepupae Meal in Practical Diets for Rainbow Trout (*Oncorhynchus mykiss*).
718 *Animals* 9, 251. <https://doi.org/10.3390/ani9050251>
- 719 Carlton, E.D., Demas, G.E., French, S.S., 2012. Leptin, a neuroendocrine mediator of immune
720 responses, inflammation, and sickness behaviors. *Horm. Behav.* 62, 272–279.
721 <https://doi.org/10.1016/j.yhbeh.2012.04.010>
- 722 Chisada, S., Kurokawa, T., Murashita, K., Rønnestad, I., Taniguchi, Y., Toyoda, A., Sakaki, Y.,
723 Takeda, S., Yoshiura, Y., 2014. Leptin receptor-deficient (knockout) medaka, *Oryzias latipes*,
724 show chronic up-regulated levels of orexigenic neuropeptides, elevated food intake and stage
725 specific effects on growth and fat allocation. *Gen. Comp. Endocrinol.* 195, 9–20.
726 <https://doi.org/10.1016/J.YGCEN.2013.10.008>
- 727 Čičková, H., Newton, G.L., Lacy, R.C., Kozánek, M., 2015. The use of fly larvae for organic waste
728 treatment. *Waste Manag.* 35, 68–80. <https://doi.org/10.1016/J.WASMAN.2014.09.026>
- 729 Copeland, D.L., Duff, R.J., Liu, Q., Prokop, J., Londraville, R.L., 2011. Leptin in teleost fishes: An
730 argument for comparative study. *Front. Physiol.* <https://doi.org/10.3389/fphys.2011.00026>
- 731 Dahm, R., Geisler, R., 2006. Learning from small fry: The zebrafish as a genetic model organism for
732 aquaculture fish species. *Mar. Biotechnol.* <https://doi.org/10.1007/s10126-006-5139-0>
- 733 Daniel, D., 2018. A review on replacing fish meal in aqua feeds using plant protein sources. *Int. J.*
734 *Fish. Aquat. Stud.*
- 735 De-Santis, C., Jerry, D.R., 2007. Candidate growth genes in finfish - Where should we be looking?
736 *Aquaculture.* <https://doi.org/10.1016/j.aquaculture.2007.08.036>
- 737 Diener, S., Zurbrügg, C., Tockner, K., 2009. Conversion of organic material by black soldier fly
738 larvae: Establishing optimal feeding rates. *Waste Manag. Res.*
739 <https://doi.org/10.1177/0734242X09103838>
- 740 Dumas, A., Raggi, T., Barkhouse, J., Lewis, E., Weltzien, E., 2018. The oil fraction and partially
741 defatted meal of black soldier fly larvae (*Hermetia illucens*) affect differently growth
742 performance, feed efficiency, nutrient deposition, blood glucose and lipid digestibility of
743 rainbow trout (*Oncorhynchus mykiss*). *Aquaculture.*
744 <https://doi.org/10.1016/j.aquaculture.2018.03.038>
- 745 Earley, A.M., Graves, C.L., Shiau, C.E., 2018. Critical Role for a Subset of Intestinal Macrophages
746 in Shaping Gut Microbiota in Adult Zebrafish. *Cell Rep.*
747 <https://doi.org/10.1016/j.celrep.2018.09.025>
- 748 Edgar, R.C., Haas, B.J., Clemente, J.C., Quince, C., Knight, R., 2011. UCHIME improves sensitivity
749 and speed of chimera detection. *Bioinformatics.* <https://doi.org/10.1093/bioinformatics/btr381>
- 750 FAO, 2018. State of Fisheries and Aquaculture in the world. *State World Fish. Aquac.* 2018 - Meet.
751 *Sustain. Dev. goals.* <https://doi.org/issn 10>
- 752 FOLCH, J., LEES, M., SLOANE STANLEY, G.H., 1957. A simple method for the isolation and
753 purification of total lipides from animal tissues. *J. Biol. Chem.*

- 754 Francis, G., Makkar, H.P.S., Becker, K., 2001. Antinutritional factors present in plant-derived
755 alternate fish feed ingredients and their effects in fish. *Aquaculture*.
756 [https://doi.org/10.1016/S0044-8486\(01\)00526-9](https://doi.org/10.1016/S0044-8486(01)00526-9)
- 757 Garofalo, C., Bancalari, E., Milanović, V., Cardinali, F., Osimani, A., Sardaro, M.L.S., Bottari, B.,
758 Bernini, V., Aquilanti, L., Clementi, F., Neviani, E., Gatti, M., 2017. Study of the bacterial
759 diversity of foods: PCR-DGGE versus LH-PCR. *Int. J. Food Microbiol.*
760 <https://doi.org/10.1016/j.ijfoodmicro.2016.11.008>
- 761 Gasco, L., Finke, M., van Huis, A., 2018. Can diets containing insects promote animal health? *J.*
762 *Insects as Food Feed* 4, 1–4. <https://doi.org/10.3920/JIFF2018.x001>
- 763 Gerland, P., Raftery, A.E., Ševčíková, H., Li, N., Gu, D., Spoorenberg, T., Alkema, L., Fosdick, B.K.,
764 Chunn, J., Lalic, N., Bay, G., Buettner, T., Heilig, G.K., Wilmoth, J., 2014. World population
765 stabilization unlikely this century. *Science* (80-.). <https://doi.org/10.1126/science.1257469>
- 766 Ghanbari, M., Kneifel, W., Domig, K.J., 2015. A new view of the fish gut microbiome: Advances
767 from next-generation sequencing. *Aquaculture*.
768 <https://doi.org/10.1016/j.aquaculture.2015.06.033>
- 769 Giorgini, E., Randazzo, B., Gioacchini, G., Cardinaletti, G., Vaccari, L., Tibaldi, E., Olivotto, I.,
770 2018. New insights on the macromolecular building of rainbow trout (*O. mykiss*) intestine: FTIR
771 Imaging and histological correlative study. *Aquaculture* 497, 1–9.
772 <https://doi.org/10.1016/j.aquaculture.2018.07.032>
- 773 Grabowski, N.T., Klein, G., 2017. Bacteria encountered in raw insect, spider, scorpion, and centipede
774 taxa including edible species, and their significance from the food hygiene point of view. *Trends*
775 *Food Sci. Technol.* <https://doi.org/10.1016/j.tifs.2017.01.007>
- 776 Guillen, J., Natale, F., Carvalho, N., Casey, J., Hofherr, J., Druon, J.N., Fiore, G., Gibin, M., Zanzi,
777 A., Martinsohn, J.T., 2018. Global seafood consumption footprint. *Ambio*.
778 <https://doi.org/10.1007/s13280-018-1060-9>
- 779 Henry, M., Gasco, L., Piccolo, G., Fountoulaki, E., 2015. Review on the use of insects in the diet of
780 farmed fish: Past and future. *Anim. Feed Sci. Technol.*
781 <https://doi.org/10.1016/j.anifeedsci.2015.03.001>
- 782 Henry, M.A., Gasco, L., Chatzifotis, S., Piccolo, G., 2018. Does dietary insect meal affect the fish
783 immune system? The case of mealworm, *Tenebrio molitor* on European sea bass, *Dicentrarchus*
784 *labrax*. *Dev. Comp. Immunol.* 81, 204–209. <https://doi.org/10.1016/J.DCI.2017.12.002>
- 785 Huyben, D., Vidaković, A., Werner Hallgren, S., Langeland, M., 2019. High-throughput sequencing
786 of gut microbiota in rainbow trout (*Oncorhynchus mykiss*) fed larval and pre-pupae stages of
787 black soldier fly (*Hermetia illucens*). *Aquaculture*.
788 <https://doi.org/10.1016/j.aquaculture.2018.10.034>
- 789 Jin, G., Yang, F., Hu, C., Shen, H., Zhao, Z.K., 2012. Enzyme-assisted extraction of lipids directly
790 from the culture of the oleaginous yeast *Rhodospiridium toruloides*. *Bioresour. Technol.*
791 <https://doi.org/10.1016/j.biortech.2012.01.152>
- 792 Johnston, I.A., Macqueen, D.J., Watabe, S., 2008. Molecular Biotechnology of Development and
793 Growth in Fish Muscle, in: *Fisheries for Global Welfare and Environment*.
- 794 Klindworth, A., Pruesse, E., Schweer, T., Peplies, J., Quast, C., Horn, M., Glöckner, F.O., 2013.
795 Evaluation of general 16S ribosomal RNA gene PCR primers for classical and next-generation
796 sequencing-based diversity studies. *Nucleic Acids Res.* <https://doi.org/10.1093/nar/gks808>

- 797 Komprda, T., Zorníková, G., Rozíková, V., Borkovcová, M., Przywarová, A., 2013. The effect of
798 dietary *Salvia hispanica* seed on the content of n-3 long-chain polyunsaturated fatty acids in
799 tissues of selected animal species, including edible insects. *J. Food Compos. Anal.*
800 <https://doi.org/10.1016/j.jfca.2013.06.010>
- 801 Kroeckel, S., Harjes, A.G.E., Roth, I., Katz, H., Wuertz, S., Susenbeth, A., Schulz, C., 2012. When a
802 turbot catches a fly: Evaluation of a pre-pupae meal of the Black Soldier Fly (*Hermetia illucens*)
803 as fish meal substitute - Growth performance and chitin degradation in juvenile turbot (*Psetta*
804 *maxima*). *Aquaculture*. <https://doi.org/10.1016/j.aquaculture.2012.08.041>
- 805 Lafrance, V., Inoue, W., Kan, B., Luheshi, G.N., 2010. Leptin modulates cell morphology and
806 cytokine release in microglia. *Brain. Behav. Immun.* 24, 358–365.
807 <https://doi.org/10.1016/j.bbi.2009.11.003>
- 808 Lawrence, C., 2007. The husbandry of zebrafish (*Danio rerio*): A review. *Aquaculture* 269, 1–20.
809 <https://doi.org/10.1016/j.aquaculture.2007.04.077>
- 810 Lawrence, C., Adatto, I., Best, J., James, A., Maloney, K., 2012. Generation time of zebrafish (*Danio*
811 *rerio*) and medakas (*Oryzias latipes*) housed in the same aquaculture facility. *Lab Anim. (NY)*.
812 <https://doi.org/10.1038/labani0612-158>
- 813 Li, A., Yuan, X., Liang, X.-F., Liu, L., Li, Jie, Li, B., Fang, J., Li, Jiao, He, S., Xue, M., Wang, J.,
814 Tao, Y.-X., 2016. Adaptations of lipid metabolism and food intake in response to low and high
815 fat diets in juvenile grass carp (*Ctenopharyngodon idellus*). *Aquaculture* 457, 43–49.
816 <https://doi.org/10.1016/J.AQUACULTURE.2016.01.014>
- 817 Li, C.H., cao, J., Zhou, Y.Z., Zhang, H.S., Gong, H.Y., Zhou, J.L., 2014. The midgut bacterial flora
818 of laboratory-reared hard ticks, *haemaphysalis longicornis*, *hyalomma asiaticum*, and
819 *rhipicephalus haemaphysaloides*. *J. Integr. Agric.* [https://doi.org/10.1016/S2095-3119\(13\)60517-1](https://doi.org/10.1016/S2095-3119(13)60517-1)
- 821 Li, S., Ji, H., Zhang, B., Zhou, J., Yu, H., 2017. Defatted black soldier fly (*Hermetia illucens*) larvae
822 meal in diets for juvenile Jian carp (*Cyprinus carpio* var. Jian): Growth performance, antioxidant
823 enzyme activities, digestive enzyme activities, intestine and hepatopancreas histological
824 structure. *Aquaculture* 477, 62–70. <https://doi.org/10.1016/J.AQUACULTURE.2017.04.015>
- 825 Li, W., Li, Q., Zheng, L., Wang, Y., Zhang, J., Yu, Z., Zhang, Y., 2015. Potential biodiesel and biogas
826 production from corncob by anaerobic fermentation and black soldier fly. *Bioresour. Technol.*
827 <https://doi.org/10.1016/j.biortech.2015.06.112>
- 828 Li, Y., Kortner, T.M., Chikwati, E.M., Munang'andu, H.M., Lock, E.J., Krogdahl, Å., 2019. Gut
829 health and vaccination response in pre-smolt Atlantic salmon (*Salmo salar*) fed black soldier fly
830 (*Hermetia illucens*) larvae meal. *Fish Shellfish Immunol.*
831 <https://doi.org/10.1016/j.fsi.2018.12.057>
- 832 Liland, N.S., Biancarosa, I., Araujo, P., Biemans, D., Bruckner, C.G., Waagbø, R., Torstensen, B.E.,
833 Lock, E.J., 2017. Modulation of nutrient composition of black soldier fly (*Hermetia illucens*)
834 larvae by feeding seaweed-enriched media. *PLoS One*.
835 <https://doi.org/10.1371/journal.pone.0183188>
- 836 Lock, E.R., Arsiwalla, T., Waagbø, R., 2016. Insect larvae meal as an alternative source of nutrients
837 in the diet of Atlantic salmon (*Salmo salar*) postsmolt. *Aquac. Nutr.* 22, 1202–1213.
838 <https://doi.org/10.1111/anu.12343>
- 839 Magalhães, R., Sánchez-López, A., Leal, R.S., Martínez-Llorens, S., Oliva-Teles, A., Peres, H., 2017.
840 Black soldier fly (*Hermetia illucens*) pre-pupae meal as a fish meal replacement in diets for

- 841 European seabass (*Dicentrarchus labrax*). *Aquaculture* 476, 79–85.
842 <https://doi.org/10.1016/J.AQUACULTURE.2017.04.021>
- 843 Magoč, T., Salzberg, S.L., 2011. FLASH: Fast length adjustment of short reads to improve genome
844 assemblies. *Bioinformatics*. <https://doi.org/10.1093/bioinformatics/btr507>
- 845 Makkar, H.P.S., Tran, G., Heuzé, V., Ankers, P., 2014. State-of-the-art on use of insects as animal
846 feed. *Anim. Feed Sci. Technol.* <https://doi.org/10.1016/j.anifeedsci.2014.07.008>
- 847 Manceron, S., Ben-Ari, T., Dumas, P., 2014. Feeding proteins to livestock: Global land use and food
848 vs. feed competition . OCL. <https://doi.org/10.1051/ocl/2014020>
- 849 May, B.M., 1961. The occurrence in New Zealand and the life-history of the soldier fly *Hermetia*
850 *illucens* (L.) (Diptera: Stratiomyidae). *New Zeal. J. Sci.*
- 851 Meneguz, M., Schiavone, A., Gai, F., Dama, A., Lussiana, C., Renna, M., Gasco, L., 2018. Effect of
852 rearing substrate on growth performance, waste reduction efficiency and chemical composition
853 of black soldier fly (*Hermetia illucens*) larvae. *J. Sci. Food Agric.*
854 <https://doi.org/10.1002/jsfa.9127>
- 855 Merino, G., Barange, M., Blanchard, J.L., Harle, J., Holmes, R., Allen, I., Allison, E.H., Badjeck,
856 M.C., Dulvy, N.K., Holt, J., Jennings, S., Mullon, C., Rodwell, L.D., 2012. Can marine fisheries
857 and aquaculture meet fish demand from a growing human population in a changing climate?
858 *Glob. Environ. Chang.* <https://doi.org/10.1016/j.gloenvcha.2012.03.003>
- 859 Merrifield, D.L., Shaw, B.J., Harper, G.M., Saoud, I.P., Davies, S.J., Handy, R.D., Henry, T.B., 2013.
860 Ingestion of metal-nanoparticle contaminated food disrupts endogenous microbiota in zebrafish
861 (*Danio rerio*). *Environ. Pollut.* <https://doi.org/10.1016/j.envpol.2012.11.017>
- 862 Müller, A., Wolf, D., Gutzeit, H.O., 2017. The black soldier fly, *Hermetia illucens* – a promising
863 source for sustainable production of proteins, lipids and bioactive substances. *Zeitschrift für*
864 *Naturforsch. C* 72, 351–363. <https://doi.org/10.1515/znc-2017-0030>
- 865 Murthy, P.S., Naidu, M.M., 2012. Resources , Conservation and Recycling Sustainable management
866 of coffee industry by-products and value addition — A review. *Resour. Conserv. Recycl.* 66,
867 45–58. <https://doi.org/10.1016/j.resconrec.2012.06.005>
- 868 Mussatto, S.I., Machado, E.M.S., Martins, S., Teixeira, J.A., 2011. Production, Composition, and
869 Application of Coffee and Its Industrial Residues. *Food Bioprocess Technol.* 4, 661–672.
870 <https://doi.org/10.1007/s11947-011-0565-z>
- 871 Naylor, R.L., Hardy, R.W., Bureau, D.P., Chiu, A., Elliott, M., Farrell, A.P., Forster, I., Gatlin, D.M.,
872 Goldberg, R.J., Hua, K., Nichols, P.D., 2009. Feeding aquaculture in an era of finite resources.
873 *Proc. Natl. Acad. Sci.* <https://doi.org/10.1073/pnas.0905235106>
- 874 Nogales-Mérida, S., Gobbi, P., Józefiak, D., Mazurkiewicz, J., Dudek, K., Rawski, M., Kierończyk,
875 B., Józefiak, A., 2018. Insect meals in fish nutrition. *Rev. Aquac.* 1–24.
876 <https://doi.org/10.1111/raq.12281>
- 877 Official Methods of Analysis of the Association of Official Analytical Chemists, 2006. . *J. Pharm.*
878 *Sci.* <https://doi.org/10.1002/jps.2600600253>
- 879 Olivotto, I., Di Stefano, M., Rosetti, S., Cossignani, L., Pugnali, A., Giantomassi, F., Carnevali,
880 O., 2011. Live prey enrichment, with particular emphasis on HUFAs, as limiting factor in false
881 percula clownfish (*Amphiprion ocellaris*, Pomacentridae) larval development and
882 metamorphosis: Molecular and biochemical implications. *Comp. Biochem. Physiol. - A Mol.*
883 *Integr. Physiol.* <https://doi.org/10.1016/j.cbpa.2011.02.004>

- 884 Olivotto, I., Mosconi, G., Maradonna, F., Cardinali, M., Carnevali, O., 2002. Diplodus sargus
885 interrenal-pituitary response: Chemical communication in stressed fish. *Gen. Comp. Endocrinol.*
886 [https://doi.org/10.1016/S0016-6480\(02\)00024-2](https://doi.org/10.1016/S0016-6480(02)00024-2)
- 887 Olivotto, I., Yasumasu, S., Gioacchini, G., Maradonna, F., Cionna, C., Carnevali, O., 2004. Cloning
888 and expression of high choriolytic enzyme, a component of the hatching enzyme system, during
889 embryonic development of the marine ornamental fish *Chrysiptera parasema*. *Mar. Biol.*
890 <https://doi.org/10.1007/s00227-004-1404-9>
- 891 Osimani, A., Ferrocino, I., Agnolucci, M., Cocolin, L., Giovannetti, M., Cristani, C., Palla, M.,
892 Milanović, V., Roncolini, A., Sabbatini, R., Garofalo, C., Clementi, F., Cardinali, F., Petruzzelli,
893 A., Gabucci, C., Tonucci, F., Aquilanti, L., 2019. Unveiling hákarl: A study of the microbiota
894 of the traditional Icelandic fermented fish. *Food Microbiol.*
895 <https://doi.org/10.1016/j.fm.2019.03.027>
- 896 Park, H.-K., Ahima, R.S., 2015. Physiology of leptin: energy homeostasis, neuroendocrine function
897 and metabolism. *Metabolism* 64, 24–34. <https://doi.org/10.1016/J.METABOL.2014.08.004>
- 898 Piccinetti, C.C., Donati, M., Radaelli, G., Caporale, G., Mosconi, G., Palermo, F., Cossignani, L.,
899 Salvatori, R., Lopez, R.P., Olivotto, I., 2015. The effects of starving and feeding on Dover sole
900 (*Solea solea*, Soleidae, Linnaeus, 1758) stress response and early larval development. *Aquac.*
901 *Res.* <https://doi.org/10.1111/are.12410>
- 902 Piccinetti, C.C., Migliarini, B., Olivotto, I., Coletti, G., Amici, A., Carnevali, O., 2010. Appetite
903 regulation: The central role of melatonin in *Danio rerio*. *Horm. Behav.* 58, 780–785.
904 <https://doi.org/10.1016/J.YHBEH.2010.07.013>
- 905 Piccinetti, C.C., Migliarini, B., Olivotto, I., Simoniello, M.P., Giorgini, E., Carnevali, O., 2013.
906 Melatonin and Peripheral Circuitries: Insights on Appetite and Metabolism in *Danio Rerio* .
907 Zebrafish. <https://doi.org/10.1089/zeb.2012.0844>
- 908 Piccinetti, C.C., Montis, C., Bonini, M., Laurà, R., Guerrera, M.C., Radaelli, G., Vianello, F.,
909 Santinelli, V., Maradonna, F., Nozzi, V., Miccoli, A., Olivotto, I., 2014. Transfer of Silica-
910 Coated Magnetic (Fe₃O₄) Nanoparticles Through Food: A Molecular and Morphological
911 Study in Zebrafish. *Zebrafish.* <https://doi.org/10.1089/zeb.2014.1037>
- 912 Piccinetti, C.C., Ricci, L.A., Tokle, N., Radaelli, G., Pascoli, F., Cossignani, L., Palermo, F.,
913 Mosconi, G., Nozzi, V., Raccanello, F., Olivotto, I., 2012. Malnutrition may affect common sole
914 (*Solea solea* L.) growth, pigmentation and stress response: Molecular, biochemical and
915 histological implications. *Comp. Biochem. Physiol. - A Mol. Integr. Physiol.*
916 <https://doi.org/10.1016/j.cbpa.2011.12.009>
- 917 Pinotti, L., Giromini, C., Ottoboni, M., Tretola, M., Marchis, D., 2019. Review: Insects and former
918 foodstuffs for upgrading food waste biomasses/streams to feed ingredients for farm animals.
919 *Animal* 1–11. <https://doi.org/10.1017/s1751731118003622>
- 920 Randazzo, B., Chemello, G., Tortarolo, I., Chiarello, G.L., Zalas, M., Santini, A., Liberatore,
921 Margaret, Liberatore, Marco, Selli, E., Olivotto, I., 2017. A Novel Photocatalytic Purification
922 System for Fish Culture. *Zebrafish.* <https://doi.org/10.1089/zeb.2017.1448>
- 923 Reed, B., Jennings, M., 2011. Guidance on the housing and care of zebrafish *Danio rerio*. *Guid. Hous.*
924 *care zebrafish Danio rerio.*
- 925 Rimoldi, S., Gini, E., Iannini, F., Gasco, L., Terova, G., 2019. The Effects of Dietary Insect Meal
926 from *Hermetia illucens* Prepupae on Autochthonous Gut Microbiota of Rainbow Trout
927 (*Oncorhynchus mykiss*). *Animals.* <https://doi.org/10.3390/ani9040143>

- 928 Ringø, E., Zhou, Z., Olsen, R.E., Song, S.K., 2012. Use of chitin and krill in aquaculture - the effect
929 on gut microbiota and the immune system: a review. *Aquac. Nutr.* 18, 117–131.
930 <https://doi.org/10.1111/j.1365-2095.2011.00919.x>
- 931 Roeselers, G., Mittge, E.K., Stephens, W.Z., Parichy, D.M., Cavanaugh, C.M., Guillemin, K., Rawls,
932 J.F., 2011. Evidence for a core gut microbiota in the zebrafish. *ISME J.*
933 <https://doi.org/10.1038/ismej.2011.38>
- 934 Roy, S. Sen, Pal, R., 2014. Microalgae in Aquaculture: A Review with Special References to
935 Nutritional Value and Fish Dietetics. *Proc. Zool. Soc.* [https://doi.org/10.1007/s12595-013-0089-](https://doi.org/10.1007/s12595-013-0089-9)
936 9
- 937 Sánchez-Muros, M.J., Barroso, F.G., Manzano-Agugliaro, F., 2014. Insect meal as renewable source
938 of food for animal feeding: A review. *J. Clean. Prod.*
939 <https://doi.org/10.1016/j.jclepro.2013.11.068>
- 940 Sargent, J., Mcevoy, L., Estevez, A., Bell, G., Bell, M., Henderson, J., Tocher, D., 1999. Lipid
941 nutrition of marine fish during early development: current status and future directions,
942 *Aquaculture.*
- 943 Sarker, P.K., Kapuscinski, A.R., Bae, A.Y., Donaldson, E., Sitek, A.J., Fitzgerald, D.S., Edelson,
944 O.F., 2018. Towards sustainable aquafeeds: Evaluating substitution of fishmeal with lipid-
945 extracted microalgal co-product (*nannochloropsis oculata*) in diets of juvenile nile tilapia
946 (*oreochromis niloticus*). *PLoS One.* <https://doi.org/10.1371/journal.pone.0201315>
- 947 Secci, G., Mancini, S., Iaconisi, V., Gasco, L., Basto, A., Parisi, G., 2019. Can the inclusion of black
948 soldier fly (*Hermetia illucens*) in diet affect the flesh quality/nutritional traits of rainbow trout
949 (*Oncorhynchus mykiss*) after freezing and cooking? *Int. J. Food Sci. Nutr.* 70, 161–171.
950 <https://doi.org/10.1080/09637486.2018.1489529>
- 951 Shepherd, C.J., Jackson, A.J., 2013. Global fishmeal and fish-oil supply: Inputs, outputs and
952 markets. *J. Fish Biol.* <https://doi.org/10.1111/jfb.12224>
- 953 Smetana, S., Schmitt, E., Mathys, A., 2019. Sustainable use of *Hermetia illucens* insect biomass for
954 feed and food: Attributional and consequential life cycle assessment. *Resour. Conserv. Recycl.*
955 <https://doi.org/10.1016/j.resconrec.2019.01.042>
- 956 Sobrino Crespo, C., Perianes Cachero, A., Puebla Jiménez, L., Barrios, V., Arilla Ferreiro, E.,
957 2014. Peptides and Food Intake. *Front. Endocrinol. (Lausanne).* 5, 58.
958 <https://doi.org/10.3389/fendo.2014.00058>
- 959 Spinelli, R., Neri, P., Pini, M., Barbi, S., Montorsi, M., Ferrari, A.M., 2019. Using black soldier flies
960 (*Hermetia illucens*) to bioconvert waste from the livestock production chain: A life cycle
961 assessment case study, in: *WIT Transactions on Ecology and the Environment.*
962 <https://doi.org/10.2495/WM180051>
- 963 Spranghers, T., Ottoboni, M., Klootwijk, C., Oryn, A., Deboosere, S., De Meulenaer, B., Michiels,
964 J., Eeckhout, M., De Clercq, P., De Smet, S., 2017. Nutritional composition of black soldier fly
965 (*Hermetia illucens*) prepupae reared on different organic waste substrates. *J. Sci. Food Agric.*
966 <https://doi.org/10.1002/jsfa.8081>
- 967 St-Hilaire, S., Cranfill, K., McGuire, M.A., Mosley, E.E., Tomberlin, J.K., Newton, L., Sealey, W.,
968 Sheppard, C., Irving, S., 2007a. Fish offal recycling by the black soldier fly produces a foodstuff
969 high in omega-3 fatty acids. *J. World Aquac. Soc.* [https://doi.org/10.1111/j.1749-](https://doi.org/10.1111/j.1749-7345.2007.00101.x)
970 7345.2007.00101.x

- 971 St-Hilaire, S., Sheppard, C., Tomberlin, J.K., Irving, S., Newton, L., McGuire, M.A., Mosley, E.E.,
 972 Hardy, R.W., Sealey, W., 2007b. Fly prepupae as a feedstuff for rainbow trout, *Oncorhynchus*
 973 *mykiss*. *J. World Aquac. Soc.* <https://doi.org/10.1111/j.1749-7345.2006.00073.x>
- 974 Stevens, J.R., Newton, R.W., Tlusty, M., Little, D.C., 2018. The rise of aquaculture by-products:
 975 Increasing food production, value, and sustainability through strategic utilisation. *Mar. Policy.*
 976 <https://doi.org/10.1016/j.marpol.2017.12.027>
- 977 Su, J., Gong, Y., Cao, S., Lu, F., Han, D., Liu, H., Jin, J., Yang, Y., Zhu, X., Xie, S., 2017. Effects of
 978 dietary *Tenebrio molitor* meal on the growth performance, immune response and disease
 979 resistance of yellow catfish (*Pelteobagrus fulvidraco*). *Fish Shellfish Immunol.* 69, 59–66.
 980 <https://doi.org/10.1016/J.FSI.2017.08.008>
- 981 Tacon A. G. J., Metian, M., 2008. Global overview on the use of fish meal and fish oil in industrially
 982 compounded aquafeeds. *Aquaculture.* <https://doi.org/10.1016/j.aquaculture.2008.08.015>
- 983 Terova, G., Rimoldi, S., Ascione, C., Gini, E., Ceccotti, C., Gasco, L., 2019. Rainbow trout
 984 (*Oncorhynchus mykiss*) gut microbiota is modulated by insect meal from *Hermetia illucens*
 985 prepupae in the diet. *Rev. Fish Biol. Fish.* <https://doi.org/10.1007/s11160-019-09558-y>
- 986 Tlusty, M.F., Thorsen, Ø., 2017. Claiming seafood is ‘sustainable’ risks limiting improvements. *Fish*
 987 *Fish.* <https://doi.org/10.1111/faf.12170>
- 988 Tocher, D.R., 2010. Fatty acid requirements in ontogeny of marine and freshwater fish. *Aquac. Res.*
 989 41, 717–732. <https://doi.org/10.1111/j.1365-2109.2008.02150.x>
- 990 Truzzi, C., Illuminati, S., Annibaldi, A., Antonucci, M., Scarponi, G., 2017. Quantification of fatty
 991 acids in the muscle of Antarctic fish *Trematomus bernacchii* by gas chromatography-mass
 992 spectrometry: Optimization of the analytical methodology. *Chemosphere.*
 993 <https://doi.org/10.1016/j.chemosphere.2016.12.140>
- 994 Truzzi, C., Illuminati, S., Antonucci, M., Scarponi, G., Annibaldi, A., 2018. Heat shock influences
 995 the fatty acid composition of the muscle of the Antarctic fish *Trematomus bernacchii*. *Mar.*
 996 *Environ. Res.* <https://doi.org/10.1016/j.marenvres.2018.03.017>
- 997 Truzzi, C., Giorgini, E., Annibaldi, A., Antonucci, M., Illuminati, S., Scarponi, G., Riolo P., Isidoro,
 998 N., Conti C., Zarantoniello, M., Cipriani, R., Olivotto, I. (In press). **Fatty acids profile of black**
 999 **soldier fly (*Hermetia illucens*): influence of feeding substrate based on coffee-waste silverskin**
 1000 **enriched with microalgae. *Animal Feed Science and Technology, In press***
- 1001 Udayangani, R.M.C., Dananjaya, S.H.S., Nikapitiya, C., Heo, G.J., Lee, J., De Zoysa, M., 2017.
 1002 Metagenomics analysis of gut microbiota and immune modulation in zebrafish (*Danio rerio*) fed
 1003 chitosan silver nanocomposites. *Fish Shellfish Immunol.*
 1004 <https://doi.org/10.1016/j.fsi.2017.05.018>
- 1005 van Huis, A., 2013. Potential of Insects as Food and Feed in Assuring Food Security. *Annu. Rev.*
 1006 *Entomol.* 58, 563–583. <https://doi.org/10.1146/annurev-ento-120811-153704>
- 1007 Vargas-Abúndez, A.J., Randazzo, B., Foddai, M., Sanchini, L., Truzzi, C., Giorgini, E., Gasco, L.,
 1008 Olivotto, I., 2019. Insect meal based diets for clownfish: Biometric, histological, spectroscopic,
 1009 biochemical and molecular implications. *Aquaculture.*
 1010 <https://doi.org/10.1016/j.aquaculture.2018.08.018>

- 1011 Vargas, A., Randazzo, B., Riolo, P., Truzzi, C., Gioacchini, G., Giorgini, E., Loreto, N., Ruschioni,
1012 S., Zarantoniello, M., Antonucci, M., Polverini, S., Cardinaletti, G., Sabbatini, S., Tulli, F.,
1013 Olivotto, I., 2018. Rearing Zebrafish on Black Soldier Fly (*Hermetia illucens*): Biometric,
1014 Histological, Spectroscopic, Biochemical, and Molecular Implications. *Zebrafish* 15, 404–419.
1015 <https://doi.org/10.1089/zeb.2017.1559>
- 1016 Volkoff, H., 2006. The role of neuropeptide Y, orexins, cocaine and amphetamine-related transcript,
1017 cholecystokinin, amylin and leptin in the regulation of feeding in fish. *Comp. Biochem. Physiol.*
1018 *Part A Mol. Integr. Physiol.* 144, 325–331. <https://doi.org/10.1016/J.CBPA.2005.10.026>
- 1019 Wang, C., Qian, L., Wang, W., Wang, T., Deng, Z., Yang, F., Xiong, J., Feng, W., 2017. Exploring
1020 the potential of lipids from black soldier fly: New paradigm for biodiesel production (I). *Renew.*
1021 *Energy.* <https://doi.org/10.1016/j.renene.2017.04.063>
- 1022 Webster, C.D., Rawles, S.D., Koch, J.F., Thompson, K.R., Kobayashi, Y., Gannam, A.L., Twibell,
1023 R.G., Hyde, N.M., 2016. Bio-Ag reutilization of distiller’s dried grains with solubles (DDGS)
1024 as a substrate for black soldier fly larvae, *Hermetia illucens*, along with poultry by-product meal
1025 and soybean meal, as total replacement of fish meal in diets for Nile tilapia, *Oreochr. Aquac.*
1026 *Nutr.* 22, 976–988. <https://doi.org/10.1111/anu.12316>
- 1027 Weisburg, W.G., Barns, S.M., Pelletier, D.A., Lane, D.J., 1991. 16S ribosomal DNA amplification
1028 for phylogenetic study. *J. Bacteriol.*
- 1029 Xiao, X., Jin, P., Zheng, L., Cai, M., Yu, Z., Yu, J., Zhang, J., 2018. Effects of black soldier fly
1030 (*Hermetia illucens*) larvae meal protein as a fishmeal replacement on the growth and immune
1031 index of yellow catfish (*Pelteobagrus fulvidraco*). *Aquac. Res.* <https://doi.org/10.1111/are.13611>
- 1032 Xu, W.J., Jin, J.Y., Zou, T., Han, D., Liu, H.K., Zhu, X.M., Yang, Y.X., Xie, S.Q., 2019. Growth,
1033 feed utilization and metabolic responses of three gibel carp (*Carassius gibelio*) strains to fishmeal
1034 and plant protein-based diets. *Aquac. Nutr.* <https://doi.org/10.1111/anu.12855>
- 1035 Zarantoniello, M., Bruni, L., Randazzo, B., Vargas, A., Gioacchini, G., Truzzi, C., Annibaldi, A.,
1036 Riolo, P., Parisi, G., Cardinaletti, G., Tulli, F., Olivotto, I., 2018. Partial Dietary Inclusion of
1037 *Hermetia illucens* (Black Soldier Fly) Full-Fat Prepupae in Zebrafish Feed: Biometric,
1038 Histological, Biochemical, and Molecular Implications. *Zebrafish* 15, 519–532.
1039 <https://doi.org/10.1089/zeb.2018.1596>
- 1040 Zarantoniello, M., Randazzo, B., Truzzi, C., Giorgini, E., Marcellucci, C., Vargas-Abúndez, J.A.,
1041 Zimbelli, A., Annibaldi, A., Parisi, G., Tulli, F., Riolo, P., Olivotto, I., 2019. A six-months study
1042 on Black Soldier Fly (*Hermetia illucens*) based diets in zebrafish. *Sci. Rep.* 9, 8598.
1043 <https://doi.org/10.1038/s41598-019-45172-5>
- 1044 Zhang, Y., Feng, S., Chen, J., Qin, C., Lin, H., Li, W., 2012. Stimulatory effects of chitinase on
1045 growth and immune defense of orange-spotted grouper (*Epinephelus coioides*). *Fish Shellfish*
1046 *Immunol.* 32, 844–854. <https://doi.org/10.1016/J.FSI.2012.02.009>
- 1047 Zhou, Z., Karlsen, Ø., He, S., Olsen, R.E., Yao, B., Ringø, E., 2013. The effect of dietary chitin on
1048 the autochthonous gut bacteria of Atlantic cod (*Gadus morhua* L.). *Aquac. Res.* 44, 1889–1900.
1049 <https://doi.org/10.1111/j.1365-2109.2012.03194.x>

1050

1051 8. Appendix

1052 TABLE 1A. OBSERVED DIVERSITY, GOOD’S COVERAGE AND NUMBER OF SEQUENCES FOR
1053 THE 16S RRNA AMPLICONS OBTAINED FROM ZEBRAFISH GUT (G) SAMPLES.

1054

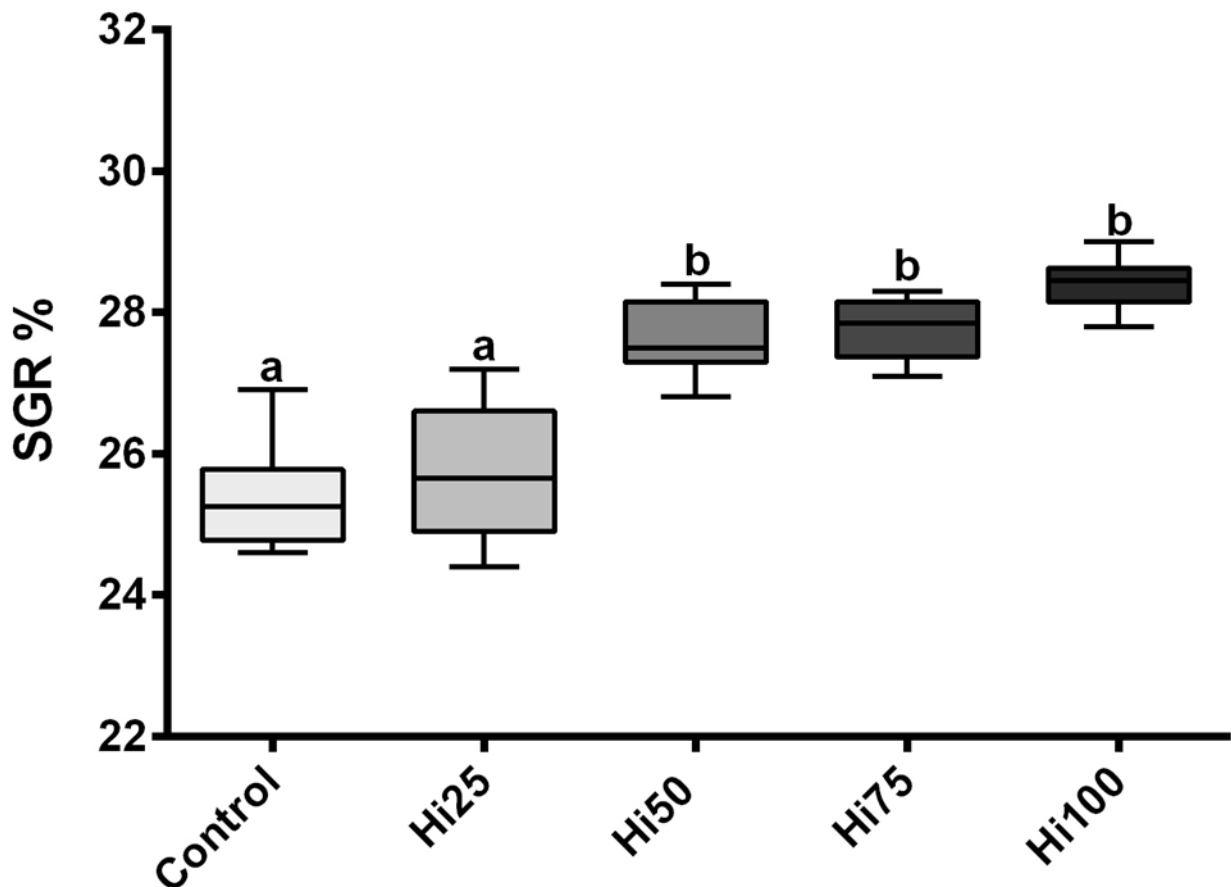
Sample	Goods_coverage	PD_whole_tree	chao1	Observed_species	Shannon
GHi0	99.77	19.37	418.40	299	3.41
GHi25	99.88	13.26	294.02	259	4.01
GHi50	99.80	20.37	462.88	380	3.25
GHi75	99.75	21.23	470.88	388	3.49
GHi100	99.75	19.97	419.85	332	3.72

1055 Zebrafish insect-based diets were prepared by including graded levels of insect meal (25, 50, 75, and 100 %,
1056 referred to as Hi25 and Hi50, Hi75 and Hi100, respectively) in the Hi0 formulation (control diet without insect
1057 addition).
1058

1059

1060 Figure Legends

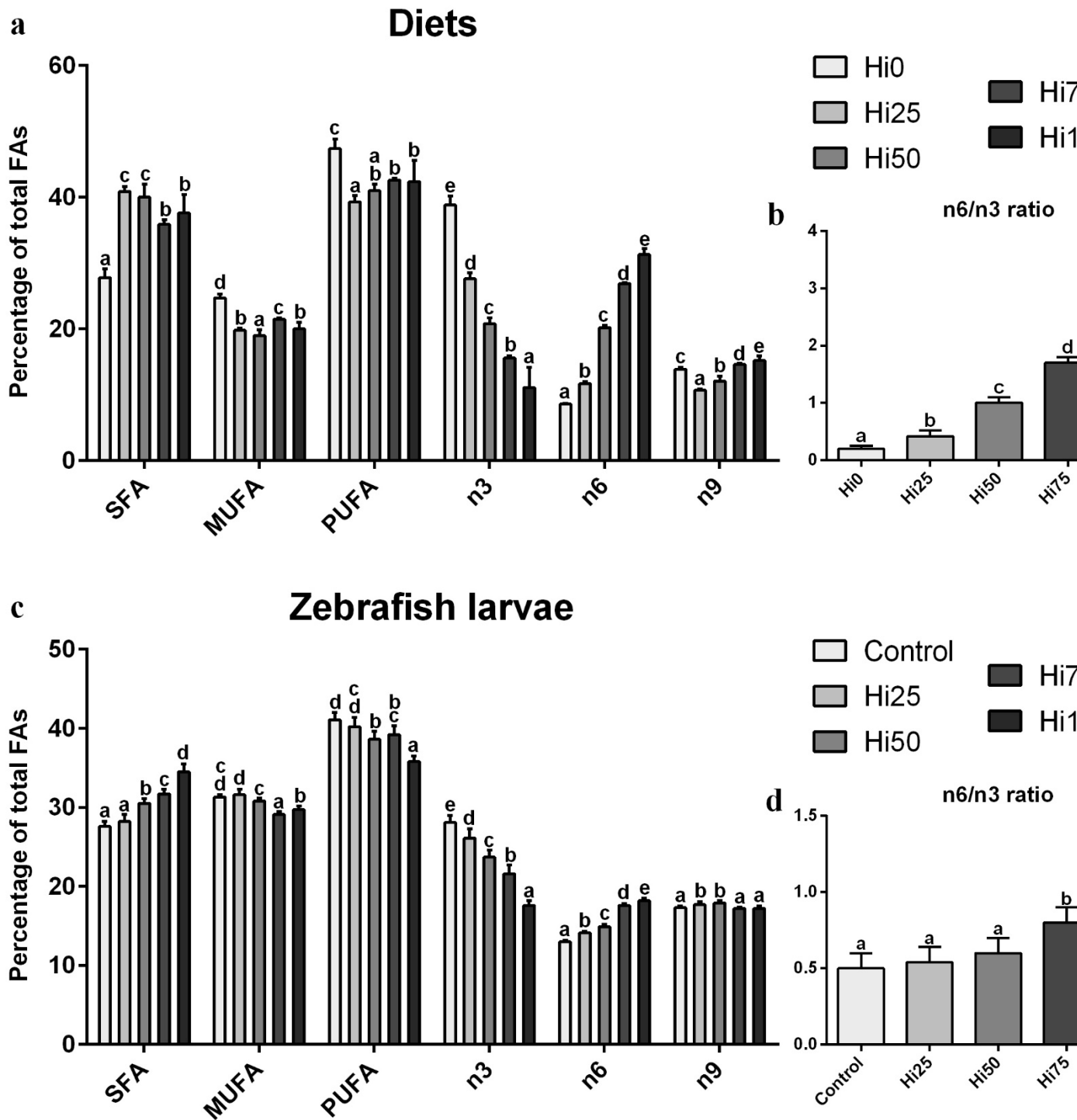
1061 **Figure 1. Specific Growth Rate (% weight growth day⁻¹) of zebrafish larvae.** Zebrafish larvae fed diets including 0,
1062 25, 50, 75 and 100% of BSF meal (Control, Hi25, Hi50, Hi75 and Hi100). Boxplots show minimum and maximum
1063 (whiskers), first quartile, median and third quartile (box). Different letters denote statistically significant differences
1064 among experimental groups.



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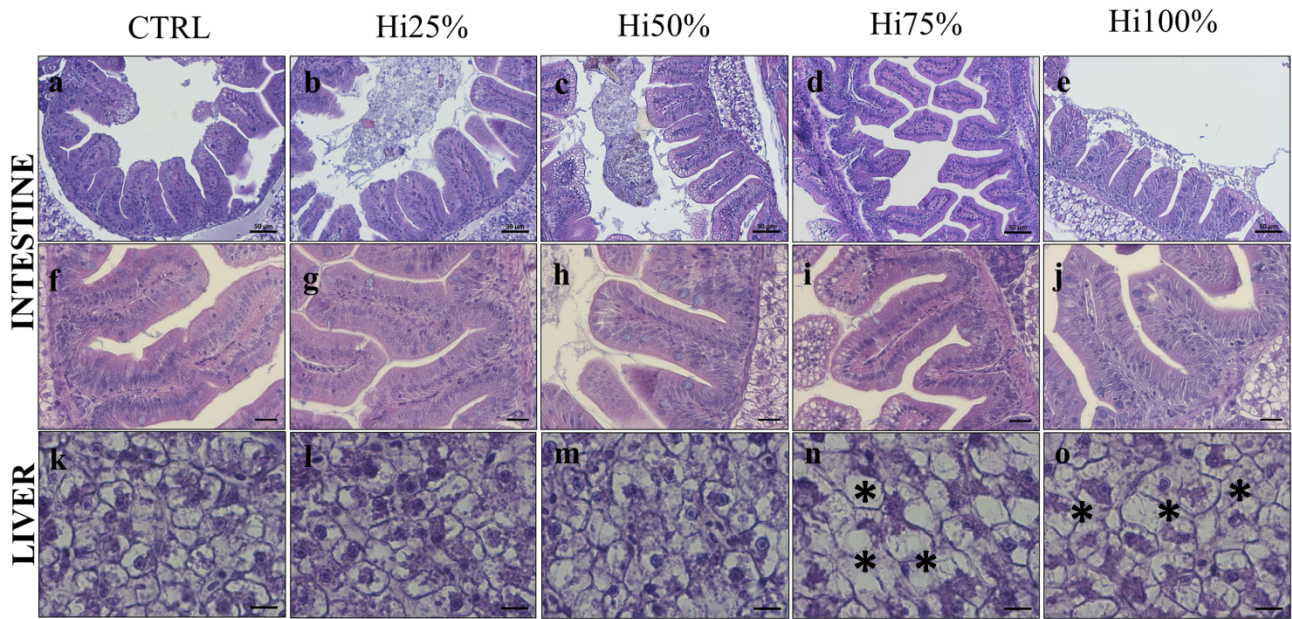
Figure 2. Content of SFA, MUFA and PUFA (as % of total FA) and contribution of omega 3 (n3), omega 6 (n6) and omega 9 (n9) fatty acids to lipid profile. (a,b) experimental diets; (c,d) zebrafish larvae. Control, Hi25, Hi50, Hi75 and Hi100 diets were characterized by 0, 25, 50, 75 or 100% inclusion of BSF meal. Zebrafish larvae fed diets including 0, 25, 50, 75 and 100% of BSF meal (Control, Hi25, Hi50, Hi75 and Hi100). Different letters indicate statistically significant differences among experimental groups compared within the same fatty acid class (p<0.05). Values are presented as mean ± SD (n = 12).



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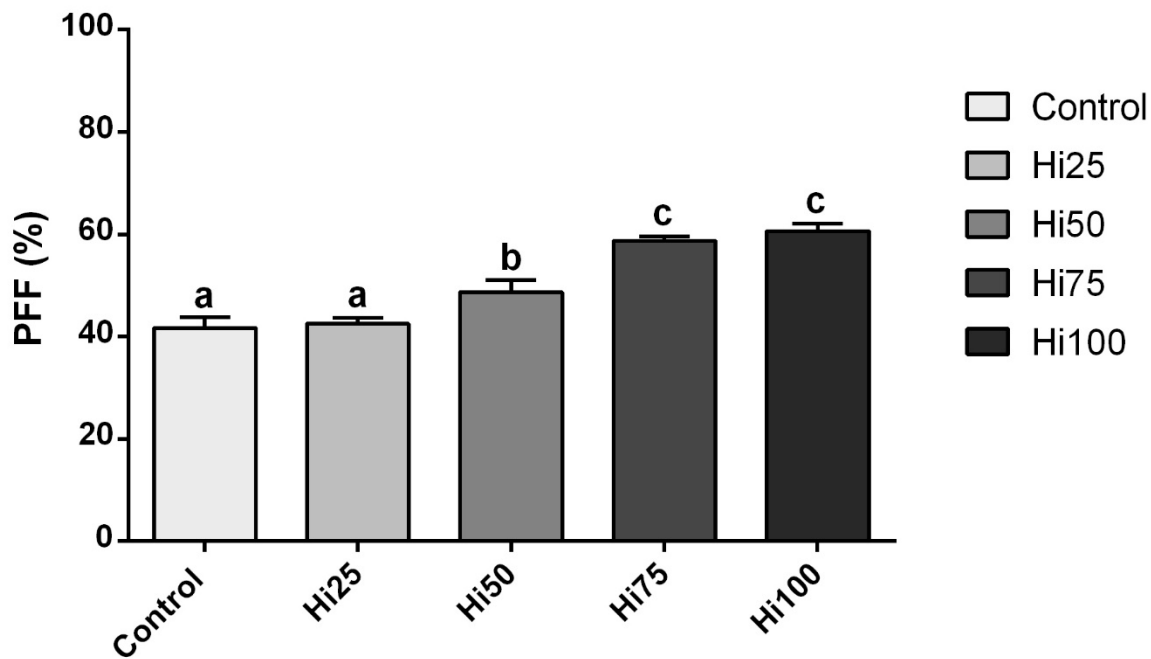
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Figure 3. Example of histomorphology of intestine and liver of zebrafish larvae. (a-j) intestine; (k-o) liver. Zebrafish larvae fed diets including 0, 25, 50, 75 and 100% of BSF meal (Control, Hi25, Hi50, Hi75 and Hi100). Scale bars: (a-e) 50 μ m; (f-j) 20 μ m; (k-o) 10 μ m. * indicates lipid accumulation.



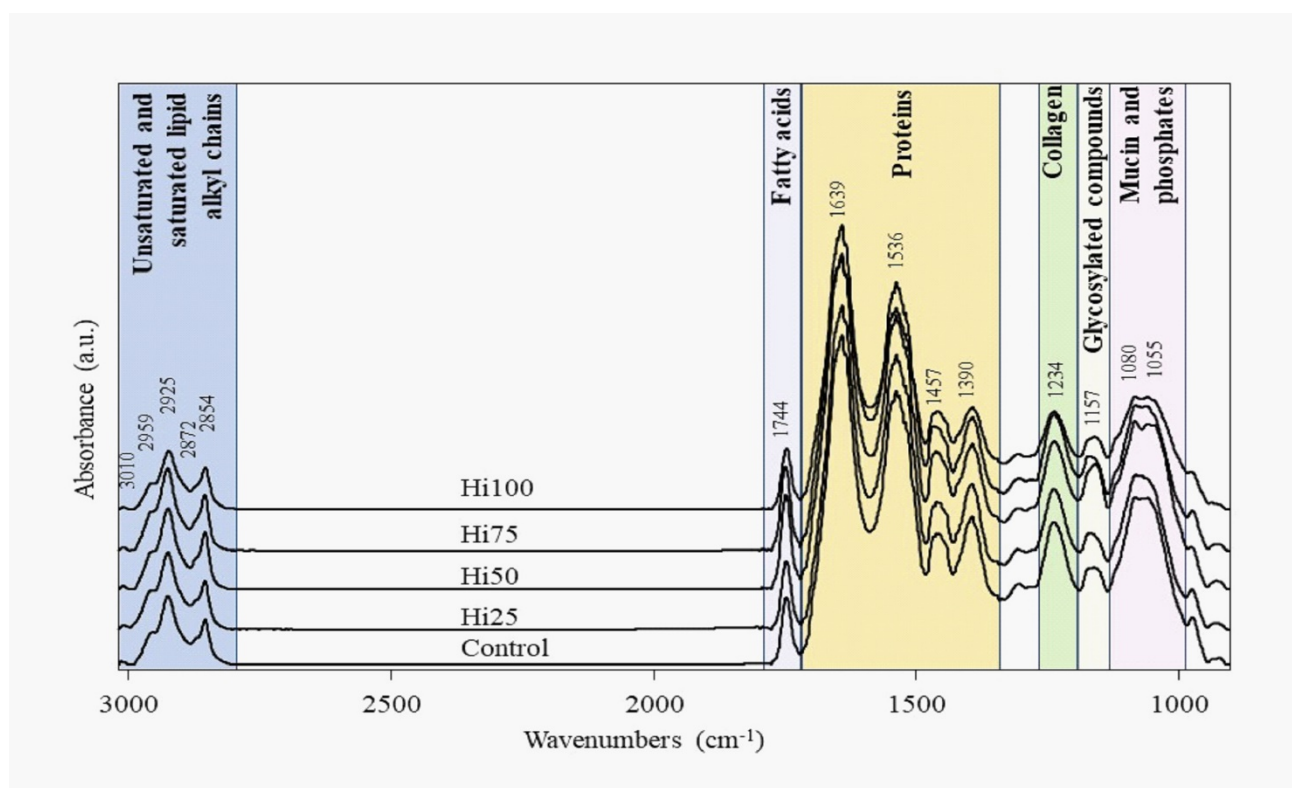
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Figure 4. Percentage of fat fraction (PFF) in liver tissue calculated on histological sections. in control, Hi25, Hi50, Hi75 and Hi100 groups. Values are presented as mean \pm SD (n=9). Different letters indicate statistically significant differences among the experimental groups.



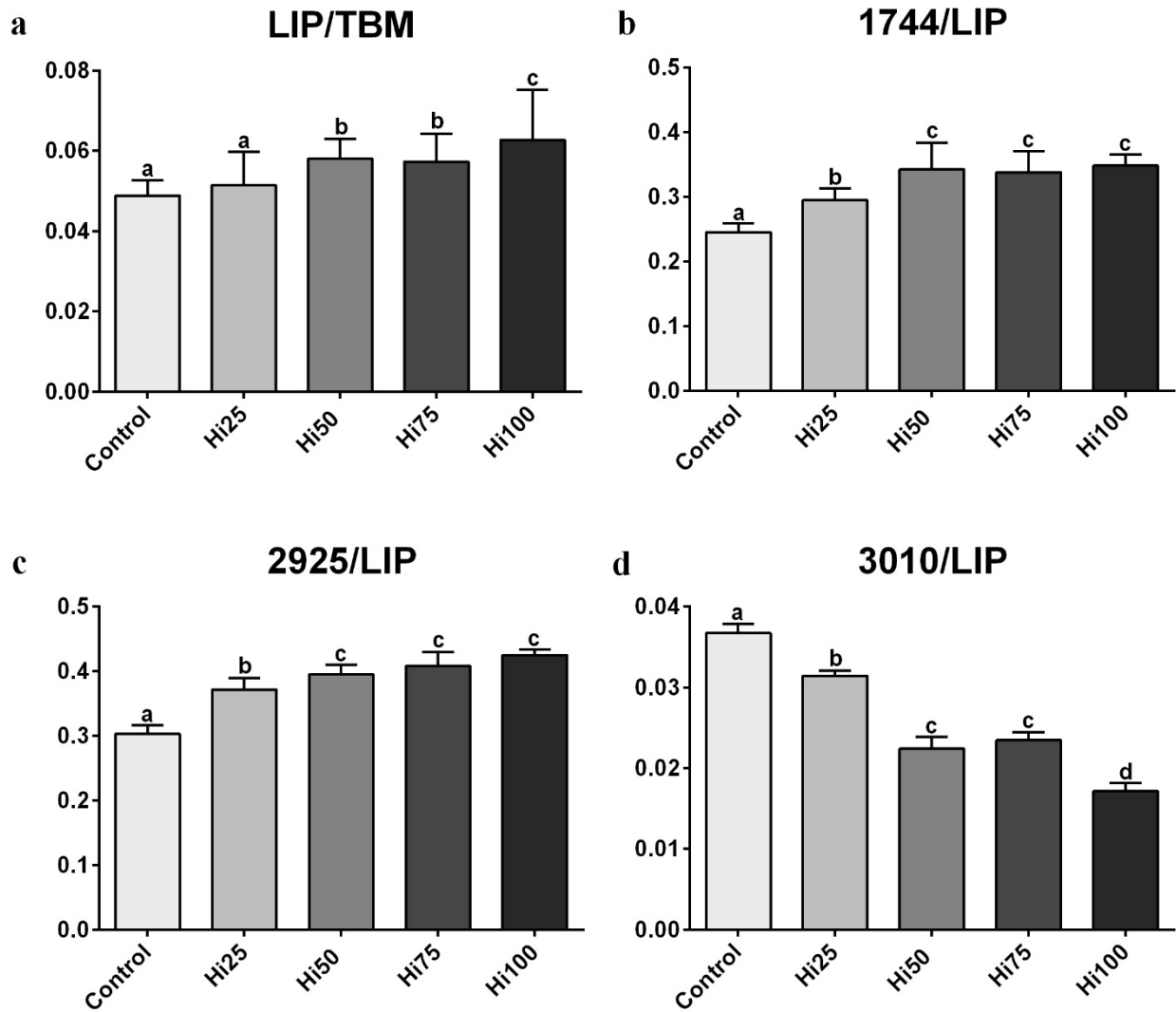
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Figure 5. IR spectra of zebrafish larvae. Average absorbance spectra of zebrafish larvae fed diets including 0, 25, 50, 75 and 100% of BSF meal (Control, Hi25, Hi50, Hi75 and Hi100). For clarity reasons, spectra are shifted along y-axis. The position (in terms of wavenumbers) of the most featuring IR peaks is reported, together with (colored boxes) the corresponding biochemical meaning.



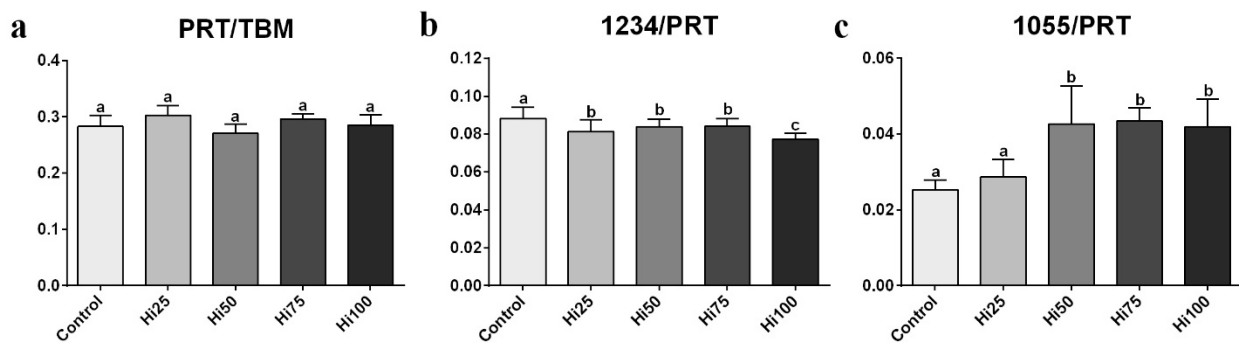
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Figure 6. Statistical analysis of lipid composition. Numerical variation of IR band area ratios in zebrafish larvae fed diets including 0, 25, 50, 75 and 100% of BSF meal (Control, Hi25, Hi50, Hi75 and Hi100): (a) LIP/TBM (overall amount of lipids); (b) 1744/LIP (amount of fatty acids with respect to lipids); (c) 2925/LIP (amount of saturated fatty acids with respect to lipids) and (d) 3010/LIP (amount of unsaturated fatty acids with respect to lipids). Different letters above histograms indicate statistically significant differences among groups ($p < 0.05$).



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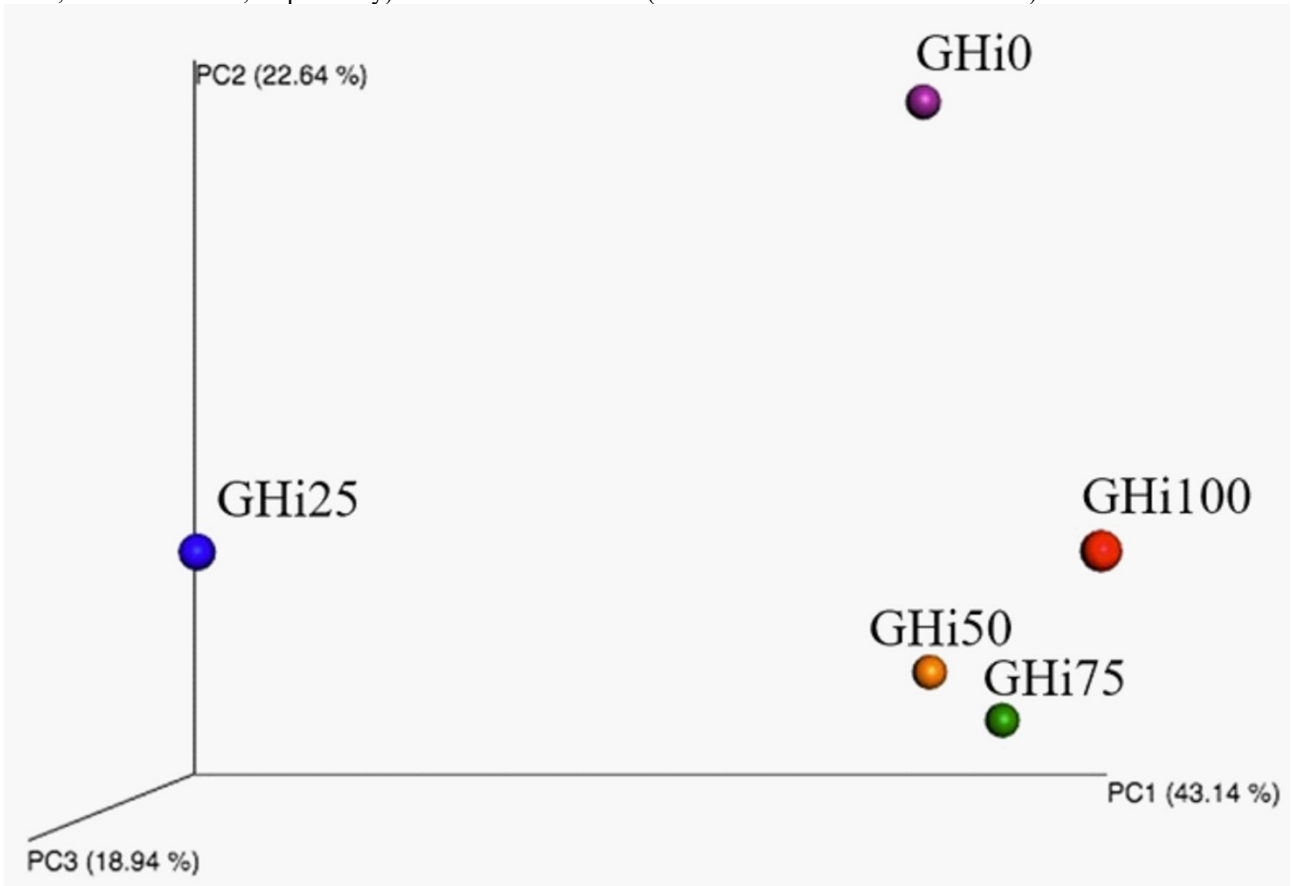
Figure 7. Statistical analysis of protein composition. Numerical variation of IR band area ratios in Zebrafish larvae fed diets including 0, 25, 50, 75 and 100% of BSF meal (Control, Hi25, Hi50, Hi75 and Hi100): (a) PRT/TBM (overall amount of proteins); (b) 1234/PRT (amount of collagen with respect to proteins) and (c) 1055/PRT (amount of mucin with respect to proteins). Different letters above histograms indicate statistically significant differences among groups ($p < 0.05$).



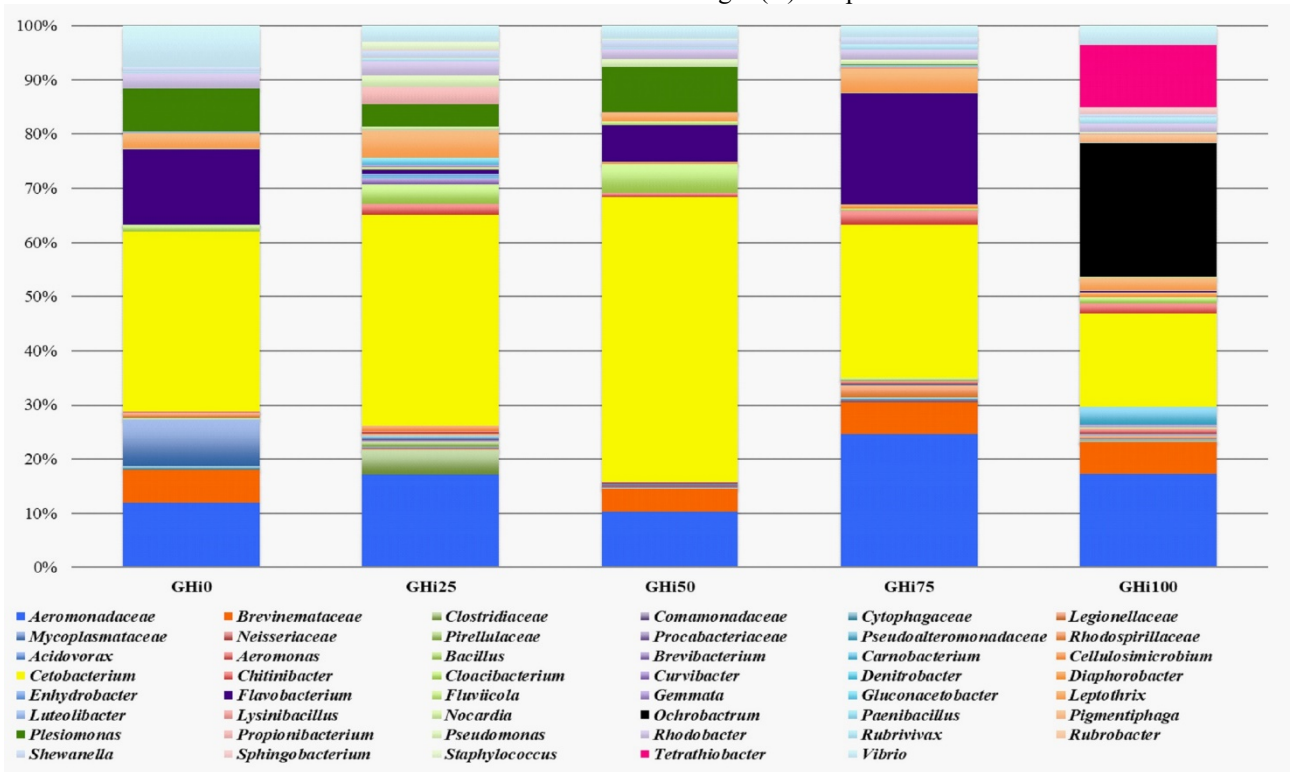
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Figure 8. PCA based on the OTU abundance of the zebrafish samples grouped as a function of the amount of BSF added to the diet. The first component (PC1) accounts for the 43.14% of the variance, the second component (PC2) accounts for the 22.64 % of the variance, the third component (PC3) accounts for the 18.94% of the variance. Zebrafish

1109 insect-based diets were prepared by including graded levels of BSF meal (25, 50, 75 and 100%, referred to as Hi25 and
 1110 Hi50, Hi75 and Hi100, respectively) in the Hi0 formulation (control diet without insect addition).

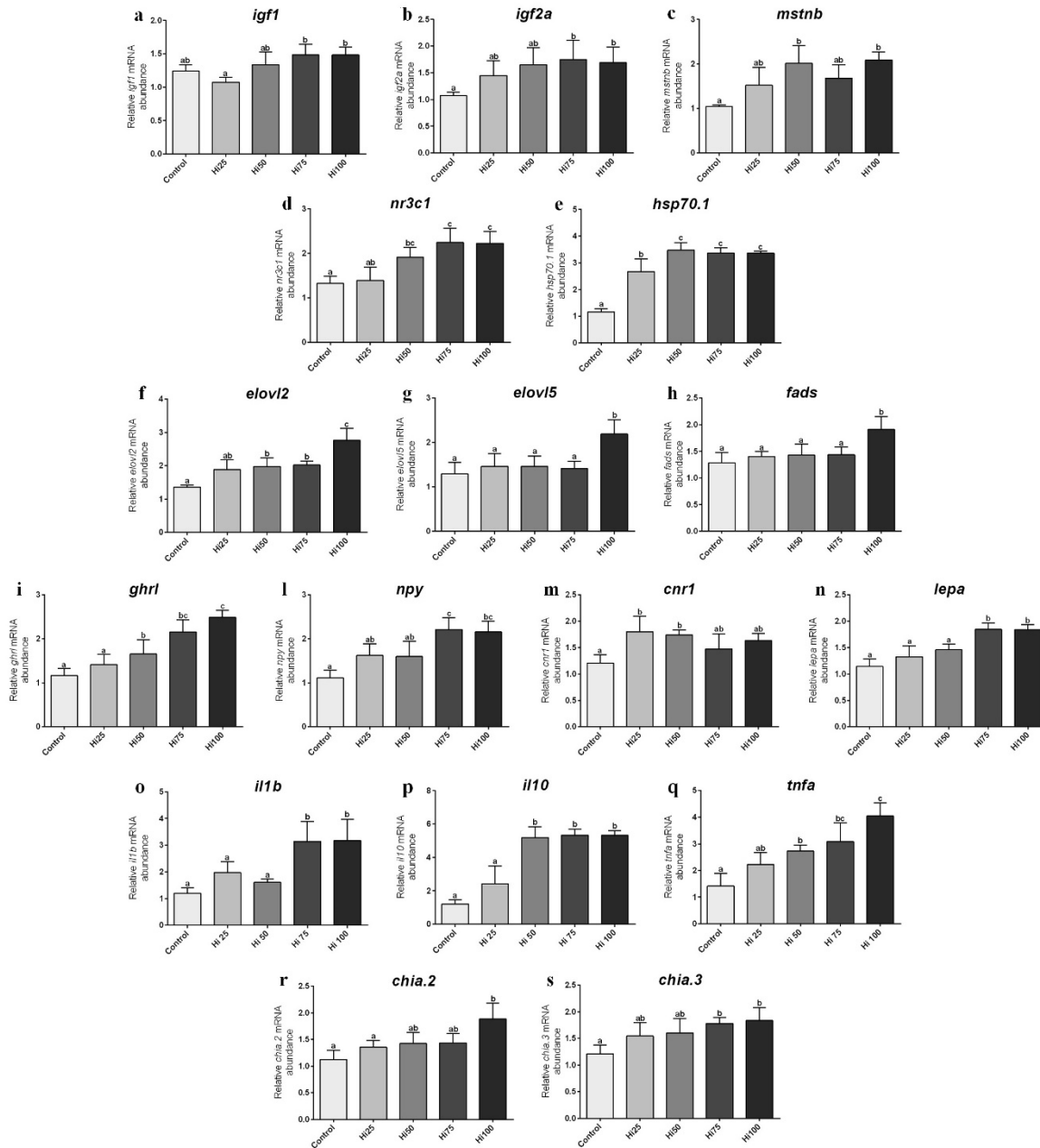


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 1113 **Figure 9.** Relative abundances as identified by MiSeq Illumina expressed as the percent ratio between the sum of reads
 1114 of each OTUs and the total number of reads found in the zebrafish gut (G) samples.



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1117 **Figure 10. Relative mRNA levels of genes analyzed in zebrafish larvae.** (a) *igf1*, (b) *igf2a*, (c) *mstnb*, (d) *nr3c1*, (e)
 1118 *hsp70.1*, (f) *elovl2*, (g) *elovl5*, (h) *fads*, (i) *ghrl*, (l) *npy*, (m) *cnr1*, (n) *lepa*, (o) *il1b*, (p) *il10*, (q) *tnfa*, (r) *chia.2*, (s) *chia.3*.
 1119 Different letters indicate statistically significant differences among experimental groups compared within the same
 1120 sampling time ($p < 0.05$). Values are presented as mean \pm SD ($n = 5$). Zebrafish larvae fed diets including 0, 25, 50, 75 and
 1121 100% of BSF meal (Control, Hi25, Hi50, Hi75 and Hi100).
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