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Long-lasting synaptic regulation of dopamine neurons by astrocytes in the Ventral Tegmental Area

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Long-lasting synaptic regulation of dopamine neurons by astrocytes in the Ventral Tegmental Area

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ABSTRACT

The plasticity of glutamatergic transmission in the Ventral Tegmental Area (VTA) represents a fundamental mechanism in the modulation of dopamine neuron burst firing and the phasic dopamine release at VTA target regions. These processes encode basic behavioral responses, including locomotor activity, learning and motivated-behaviors. Here we describe a hitherto unidentified mechanism of long-lasting potentiation of glutamatergic synapses on DA neurons. We found that VTA astrocytes respond to dopamine neuron bursts with Ca^{2+} elevations that require activation of endocannabinoid CB1 and dopamine D2 receptors colocalized at the same astrocytic process. Astrocytes, in turn, release glutamate that, through presynaptic metabotropic glutamate receptor activation coupled with neuronal nitric oxide production, induces long-lasting potentiation of excitatory synapses on adjacent dopamine neurons. Consistent with this finding, selective activation of VTA astrocytes increases dopamine neuron bursts *in vivo* and induces locomotor hyperactivity. Astrocytes play, therefore, a key role in the modulation of VTA dopamine neuron activity.

INTRODUCTION

Dopamine (DA) neurons in the Ventral Tegmental Area (VTA) regulate a wide array of physiological functions including locomotor activity, attention, motivation and reward-based learning¹⁻³. A fundamental step in these DA-dependent functions is the transition in the spiking activity of VTA DA neurons from tonic, low frequency firing at rest, to high frequency bursts that modulate the action of DA by determining the synaptic phasic release of DA at VTA target areas such as nucleus accumbens (NAc), medial prefrontal cortex, hippocampus, and amygdala⁴⁻⁷. This transition and the bursting activity of DA neurons are under crucial control of glutamatergic afferent inputs to the VTA originating from various brain regions^{4,7,8}. Importantly, the enduring changes in the strength of these glutamatergic synapses exert profound effects on DA neurons regulating their burst firing mode and the

49 release of dopamine at target regions^{1,9}. The plasticity of these glutamatergic synapses
50 represents, therefore, a key mechanism in the modulation of DA transmission and DA-
51 dependent behaviors. While extensive studies highlighted the role of neuronal signals in the
52 synaptic plasticity of VTA circuits^{1,10}, the role of astrocytes has been poorly investigated.

53 A recent study reported that optogenetic stimulation of channelrhodopsin-expressing
54 VTA astrocytes alters glutamate transport, favoring DA neuron inhibition and avoidance
55 behavior¹¹. However, this type of stimulation depolarizes astrocytes, leading to a significant
56 increase in extracellular K⁺ and a subsequent neuronal excitation¹². Whether astrocytes are
57 functionally recruited to the VTA circuitry by neuronal signals and influence the plasticity of
58 glutamatergic synaptic transmission to VTA DA neurons remains totally unexplored.

59 Astrocytes are active components of brain circuits. Besides their support and
60 metabolic functions, they respond with Ca²⁺ elevations to neurotransmitters and, in turn,
61 release gliotransmitters that regulate synaptic transmission and plasticity^{13–15}. Astrocytes
62 are similarly activated by local signals, such as endocannabinoids (eCBs), released by
63 neurons at somatodendritic levels. In various brain areas, including the VTA, eCBs act as
64 retrograde signals that induce depression of synaptic neurotransmitter release upon
65 presynaptic type-1 cannabinoid receptor (CB1R) activation^{16,17}. Studies in hippocampus and
66 dorsal striatum revealed that eCBs also target astrocytic CB1Rs evoking Ca²⁺ elevations
67 and glutamate release that potentiates distant excitatory synapses^{18–20}. Whether this lateral
68 potentiation of synaptic transmission is also operative in the VTA is unknown.

69 Using *ex vivo* and *in vivo* approaches, we investigated whether eCBs released by
70 bursting discharges of VTA DA neurons²¹ induce a potentiation of glutamatergic
71 transmission to nearby DA neurons and whether this action is mediated by astrocytes.
72 Because VTA DA neurons, beside eCBs, release DA at somatodendritic levels²², we
73 investigated whether DA is also involved in DA neuron-to-astrocyte signalling. Finally, we
74 evaluated the functional consequences of a specific activation of astrocytes *in vivo* at the

75 level of both VTA DA neuron firing and locomotor activity. Our results unveil a reciprocal
76 functional signaling between DA neurons and astrocytes in VTA circuits.

77

78 **RESULTS**

79 **Bursting activity of individual VTA DA neurons induces long-lasting potentiation of**
80 **glutamatergic transmission in adjacent DA neurons.** We investigated whether the

81 bursting activity in individual DA neurons evokes lateral potentiation of glutamatergic
82 synaptic transmission¹⁸. In VTA slices of postnatal day (P)14-17 female mice, we recorded

83 from pairs of neurons exhibiting the typical features of DA neurons (Fig. 1a and Extended
84 Data Fig. 1a-d). In one neuron of the pair, we monitored excitatory post-synaptic currents

85 (EPSCs) evoked by low frequency stimulation of the rostral glutamatergic afferents. To the
86 second neuron, located 70-120 μ m apart, we imposed through intracellular current pulses

87 the burst firing mode that characterizes *in vivo* DA neuron activity (bursts of five action
88 potentials at 20 Hz, 2 Hz interburst frequency, 5 min duration; Extended Data Fig. 1e)²³⁻²⁵.

89 Following this bursting activity, EPSC amplitude from the first DA neuron was significantly
90 increased and this potentiation was maintained for at least 45 min (Fig. 1b and 1d). The

91 long-lasting potentiation was not observed in age-matched male mice, in which EPSC
92 amplitude was only transiently increased 3 min after bursts (Fig. 1b and 1d). We defined

93 these two forms of lateral synaptic plasticity as burst-induced long- and short-term
94 potentiation (bLTP and bSTP, respectively). Evaluation of the paired-pulse ratio (PPR) in

95 female mice revealed a significant PPR reduction at 30 and 45 minutes after bursts,
96 suggesting a presynaptic mechanism in bLTP generation (Fig. 1c). In contrast, the bSTP in

97 male mice occurred without PPR changes (Fig. 1c). Importantly, the induction of a tonic-like
98 discharge that mimics DA neuron activity at resting conditions (2 Hz action potential

99 frequency for 5 minutes, Extended Data Fig. 1e), failed to modify evoked EPSCs in adjacent

100 DA neurons at any time point tested (Fig. 1d), indicating that bLTP and bSTP are strictly
101 dependent on DA neuron bursting activity.

102

103 **Generation of bLTP requires Ca^{2+} elevations in astrocytes.** We focused our study on
104 bLTP generation mechanism and asked whether astrocytes are involved. We performed
105 experiments in type-2 inositol 1,4,5-trisphosphate receptor knock-out ($\text{IP}_3\text{R}2^{-/-}$) female mice
106 in which G-protein coupled-mediated astrocyte Ca^{2+} elevations are largely impaired^{19,26,27}.
107 In VTA slices from these mice, DA neuron bursts evoked transient, but not long-lasting
108 potentiation of synaptic transmission (Fig. 1d), suggesting that bLTP induction depends on
109 $\text{IP}_3\text{R}2$ -mediated astrocytic Ca^{2+} elevations induced by signals generated by the DA neuron.
110 This hypothesis was directly tested in VTA slices from wild type (wt) and $\text{IP}_3\text{R}2^{-/-}$ mice loaded
111 with the Ca^{2+} fluorescent indicator Fluo-4 and the specific astrocytic marker SR101. To
112 monitor Ca^{2+} signals from astrocytes in proximity of soma and dendrites, through a patch
113 pipette we filled DA neurons with the fluorescence tracer neurobiotin (Fig. 1e). We observed
114 that the sustained bursting activity of the DA neuron evoked in astrocytes of female, but not
115 male mice, Ca^{2+} elevations that lasted for at least 25 min (Fig. 1f-h). Furthermore, DA neuron
116 bursts failed to evoke Ca^{2+} elevations in astrocytes of $\text{IP}_3\text{R}2^{-/-}$ female mice (Fig. 1h). Overall,
117 these data suggest that astrocyte $\text{IP}_3\text{R}2$ -mediated Ca^{2+} elevations are required for bLTP
118 generation.

119

120 **Generation of bLTP requires eCB and DA signalling to astrocytes, presynaptic**
121 **mGluR1 activation and nitric oxide release from DA neurons.** To gain further insights
122 into the molecular mechanism of bLTP generation, we investigated whether CB1 and/or DA
123 receptors (Rs), activated by eCBs and/or DA locally released by VTA DA neurons, are
124 involved. We found that applications of either the CB1R antagonist AM251 (2 - 4 μM) or the
125 D2-type receptor antagonist eticlopride (1 μM) prevented bLTP (but not bSTP), whereas the

126 D1-type receptor antagonist SCH-23390 hydrochloride (10 μ M) was ineffective (Fig. 2a). We
127 also evaluated the role of the glutamatergic N-methyl-D-Aspartate receptor (NMDAR), which
128 is a fundamental element in the plasticity of glutamatergic synaptic transmission in different
129 brain regions²⁸, including the VTA¹⁰. We found that in the presence of the NMDAR
130 antagonist D-AP5 (50 μ M), bLTP was unchanged (Fig. 2a), indicating that NMDAR is not
131 involved in bLTP. We then hypothesized an involvement of type 1 metabotropic glutamate
132 receptor (mGluR1) activation, as previously observed for astrocyte-mediated synaptic
133 plasticity in hippocampal^{18,19} and striatal circuitries²⁰. We found that bLTP was abolished in
134 the presence of the mGluR1 receptor antagonist LY-367385 (100 μ M; Fig. 2a). Then, we
135 investigated whether nitric oxide (NO), which contributes to long-term synaptic plasticity in
136 other brain circuits^{19,29}, is also involved. We found that blocking NO synthesis in the bursting
137 DA neuron using an NO synthase inhibitor (L-NAME)-containing patch pipette (100 μ M),
138 abolished bLTP (Fig. 2a). However, a transient EPSC potentiation, that was absent in the
139 presence of the other antagonists which block bLTP (Extended Data Fig. 2a), was observed
140 6 min after DA neuron bursts, suggesting that NO release by DA neurons contributes to the
141 sustained phase of bLTP.

142 We asked whether CB1 and D2R activation, which is required for bLTP generation,
143 is also required for DA neuron burst-induced astrocytic Ca^{2+} elevations. We found that the
144 astrocyte Ca^{2+} response to DA neuron bursts was abolished in the presence of either AM251
145 or eticlopride (Fig. 2b). We next asked whether mGluR1 activation, which is required for
146 bLTP generation, is mediated by astrocytic glutamate. In such a case, the mGluR1
147 antagonist LY-367385 should not block the astrocyte Ca^{2+} response to DA neuron bursts.
148 As expected, the Ca^{2+} response of astrocytes was unaffected by LY-367385 (Fig. 2b).
149 Therefore, mGluR1 activation plays a crucial role in bLTP generation downstream of
150 astrocyte Ca^{2+} signals. Finally, we found that after blocking NO synthesis, DA neuron bursts
151 evoked in astrocytes only a transient Ca^{2+} response (Fig. 2b and Extended Data Fig. 2b),

152 indicating that NO release by DA neurons contributes to astrocyte Ca^{2+} signal dynamics, as
153 previously reported³⁰.

154 In support of the role of astrocytic CB1 and D2-type receptors in bLTP generation,
155 pre-embedding electron microscope (EM) experiments showed that, besides neurons
156 (Extended Data Fig. 3a, Table 1), astrocytes express CB1 and D2-type receptors (D2, D3
157 and D4R, Fig. 2c, Extended Data Fig. 3b, Table 1 and 2). According to our post-embedding
158 quantitative EM analysis of CB1/D2R immunogold double-labelled astrocytic processes,
159 CB1 and D2Rs colocalize at perisynaptic processes at variable distances (Fig. 2d, e, mean
160 distance: 532.2 ± 49.7 nm), indicating that the same astrocyte can sense both eCBs and
161 DA. Pre-embedding EM analysis also revealed that the mGluR1 β isoform is expressed at
162 axon terminals making asymmetric synaptic contacts (Fig. 2c; Extended Data Fig. 3c, Table
163 2 and 3), consistent with a presynaptic mechanism of bLTP, as suggested by PPR reduction.

164 Taking all these results together, we propose that bLTP in the VTA circuitry of female
165 mice is evoked by the following sequence of events (Fig. 2f). Firstly, DA neuron bursting
166 activity induces the somatodendritic release of eCBs, DA and NO; secondly, activation of
167 CB1 and D2Rs in astrocytes triggers IP₃R2-dependent Ca^{2+} elevations, modulated by
168 neuronal NO, and a subsequent glutamate release; thirdly, presynaptic mGluR1 activation
169 by astrocytic glutamate, possibly coupled with NO actions on neurons, induces a sustained
170 increase in glutamate release probability leading to bLTP of excitatory transmission onto
171 adjacent DA neurons.

172

173 **Differential expression of DA, CB1 and mGluR1 receptors in the VTA of young male**
174 **and female mice.** The bLTP was absent in the VTA of young male mice where astrocytes
175 failed to respond with Ca^{2+} elevations to DA neuron burst. We hypothesized that the lack
176 of astrocyte Ca^{2+} responses is due to absence or lower levels of CB1 and/or D2Rs in male
177 with respect to female mice. Our pre-embedding EM experiments showed that astrocytes

178 from male mice express CB1, D2, D3 and D4Rs (Fig. 3a and Extended Data Fig. 3a, d).
179 Quantitative evaluation revealed, however, that the expression of CB1 and D2Rs is higher
180 in astrocytic processes of female than male mice ($P < 0.0001$ and $P = 0.033$, respectively;
181 Fig. 3a and Table 2), whereas that of D3Rs is higher in male than female mice ($P < 0.0001$,
182 Extended Data Fig. 3d and Table 2). Furthermore, the percentage of axon terminals
183 expressing mGluR1 β in female mice is twice that observed in male mice ($P = 0.016$; Fig. 3a,
184 Extended Data Fig. 3c and Table 2). Therefore, the lack of bLTP in male mice is likely due
185 to a defective astrocyte Ca^{2+} response to DA neuron signalling caused by reduced
186 expression of CB1Rs and different pattern and density of D2/D3Rs in the astrocytic
187 membrane, coupled to mGluR1 β reduction which affects the ability of astrocytic glutamate
188 to regulate release probability at glutamatergic axon terminals.

189

190 **Selective chemogenetic activation of astrocytes rescues bLTP in young male mice.** If
191 the lack of bLTP in male mice is due, at least in part, to a lack of astrocyte Ca^{2+} responses
192 to DA neuron bursts, we expect bLTP to be rescued by stimulating astrocyte Ca^{2+} elevations.
193 As a specific stimulus, we used chemogenetic activation of Gq protein-coupled designer
194 receptor exclusively activated by designer drugs (DREADDs, hM3D(Gq)) that were
195 selectively expressed in astrocytes (Fig. 3b, c and Extended Data Fig. 4). In VTA slices from
196 mice that express both DREADDs and GCaMP6f in VTA astrocytes, we observed that bath
197 perfusion with the hM3D(Gq) agonist clozapine N-oxide (CNO, 10 μM) evoked transient
198 Ca^{2+} elevations in hM3D(Gq)-expressing astrocytes (Fig. 3d). In agreement with this
199 transient astrocyte response, parallel experiments performed in the presence of the CB1
200 and D2R antagonists, AM251 and eticlopride, revealed that CNO evoked in male mice
201 expressing hM3Dq in astrocytes, but not in non-injected control mice, a short-lasting
202 potentiation of the excitatory transmission (Fig. 3e), which became a bLTP lasting at least
203 30 min after coupling CNO with DA neuron burst firing (Fig. 3f). These results further suggest

204 that NO is required for bLTP generation. Given that these experiments were performed in
205 the presence of CB1 and D2R antagonists, these results further support that bLTP
206 generation depends on astrocytic rather than neuronal CB1 and D2Rs.

207

208 **Astrocytes induce bLTP in the VTA of both female and male adult mice.** We next
209 investigated whether the astrocyte-mediated bLTP observed in young mice is also present
210 in young adulthood. We found that in VTA slices from adolescent/adult mice (P30-70, to
211 simplify hereafter adult mice), DA neuron bursts evoked in adjacent DA neurons a bLTP
212 that was maintained for at least 30 min after bursts (Fig. 4a). In contrast to data obtained
213 from young mice, the bLTP in adult mice was observed in both female and male (Fig. 4a).
214 The presence of bLTP in adult male mice could be due to a developmentally regulated
215 expression of CB1, D2 and/or mGluR1 β receptors. Quantitative analysis from pre-
216 embedded material of adult male mice showed that the levels of CB1Rs at astrocytic
217 processes and mGluR1 β at glutamatergic terminals are, indeed, higher in adult compared
218 to young male mice, while the levels of D2Rs are comparable (Fig. 4b, Table 4). In
219 agreement with the presence of bLTP in both female and male adult mice, quantitative
220 evaluation of mGluR1 β , CB1 and D2 receptor expression revealed comparable levels in
221 these mice (Fig. 4b, Extended Data Fig. 5, Table 4). Together with data presented in figure
222 3f, these results suggest that the absence of bLTP in young male mice is mainly due to
223 CB1R low expression in astrocytic membranes.

224 The mechanism of bLTP generation in adult mice is similar to that in young female
225 mice, because it was abolished by specific D2, CB1 or mGluR1 receptor antagonists
226 (Extended Data Fig. 6). To further confirm the role played by astrocytic D2 and CB1Rs, we
227 injected the AAV9/2-hGFAP-mCherry_iCre-WPRE-hGHp into the VTA of male mice
228 carrying a “floxed” version of either the D2 or the CB1R genes, to express the Cre
229 recombinase in VTA astrocytes (Fig. 4c). Immunohistochemical experiments showed that

230 the great majority of mCherry-Cre-immuno positive cells were also GFAP-positive and only
231 a very few mCherry-Cre-immunopositive cells were NeuN-positive (Fig. 4d, Extended Data
232 Fig. 7a, b). As a control, we injected the same AAV vector in wt male mice. We found that
233 bLTP was abolished when the Cre recombinase was expressed in astrocytes containing the
234 D2 or CB1R floxed gene, whereas it could still be evoked when the recombinase was
235 expressed in wt astrocytes (Fig. 4e; see also specific comments on Discussion). Finally, as
236 in young female mice, bLTP in adult male mice was abolished when the NO synthase
237 inhibitor L-NAME was included in the patch pipette (Extended Data Fig. 6), and only a
238 reduced transient potentiation lasting no more than 6 min (EPSC amplitude (%) $t_{6\text{min}} = 112.7$
239 ± 4.4 , $p = 0.028$, $n = 7$) was observed. Altogether, these data indicate that the astrocyte-
240 mediated bLTP observed in young female mice is preserved in older female mice and is
241 also present in adult male mice with similar cellular and molecular mechanisms.

242

243 **bLTP is abolished in adult $\text{IP}_3\text{R}2^{-/-}$ mice expressing a plasmatic Ca^{2+} pump in**
244 **astrocytes.** To further explore the role of astrocyte Ca^{2+} signals in bLTP during adulthood,
245 we performed DA neuron-paired recording experiments from VTA slices of adult $\text{IP}_3\text{R}2^{-/-}$
246 female and male mice. In both groups, the amplitude of evoked EPSCs showed an increase
247 at 30 min after DA neuron bursts that was not statistically significant (Extended Data Fig. 8).
248 However, after pooling results obtained from both female and male $\text{IP}_3\text{R}2^{-/-}$ mice together,
249 we observed a statistically significant potentiation of the excitatory transmission at 30 min
250 (Fig. 4g). Consistently, after pooling results obtained from female and male mice together
251 as wt or $\text{IP}_3\text{R}2^{-/-}$ groups, the percentage of experiments showing bLTP was different ($p <$
252 0.05 , Chi square test, 85.7% female/male wt mice, $n = 14$, vs 44.4% female/male $\text{IP}_3\text{R}2^{-/-}$
253 mice, $n = 18$). These results suggest that the deletion of $\text{IP}_3\text{R}2$ has a lower impact on
254 astrocyte Ca^{2+} signal dynamics in adult than young mice and additional experimental tools
255 affecting Ca^{2+} signalling are needed to impair the Ca^{2+} dependent astrocytic actions in adult

256 mice. To validate this idea, we expressed in astrocytes from IP₃R2^{-/-} mice the plasma
257 membrane Ca²⁺ pump isoform hPMCA2w/b that significantly reduced Ca²⁺ signals in these
258 cells³¹ (Fig. 4f and Extended Data Fig. 7c, d). We found that bLTP generation was fully
259 impaired both in female and male IP₃R2^{-/-} mice that expressed hPMCA2w/b in VTA
260 astrocytes (Fig. 4g and Extended Data Fig. 8). Therefore, Ca²⁺ signaling is crucial for bLTP
261 generation also in adult mice, but the impairment of the IP₃R2-mediated signalling in these
262 mice is not sufficient to block astrocyte Ca²⁺-dependent actions.

263

264 ***In vivo* activation of VTA astrocytes favors DA neuron bursts and induces long-lasting**
265 **motor hyperactivity.** Glutamatergic synapses in the VTA circuitry modulate the firing
266 activity of DA neurons and their potentiation, mainly mediated by NMDARs, enhances the
267 burst firing in DA neurons thereby playing a key role in DA-dependent function and
268 dysfunction^{1,32}. Because astrocytes, as we show here, also induce a potentiation of these
269 glutamatergic synapses, we next investigated whether *in vivo* astrocyte activation increases
270 the burst firing mode of VTA DA neurons and eventually affects behavior. We injected AAV9-
271 GFAP-hM3D(Gq)-mcherry or AAV5.GfaABC1D.cyto-tdTomato.SV40 in the VTA of adult
272 male mice (Fig. 5a), specifically targeting astrocytes via the GFAP promoter (Extended Data
273 Fig. 9). Astrocytes were activated through brief pressure pulses applied to a CNO-containing
274 glass pipette, while recording the firing activity from individual VTA neurons that exhibit the
275 typical features of DA neurons (Fig. 5a, b). Consistent with the astrocyte-mediated
276 enhancement of glutamatergic transmission to DA neurons observed in VTA slices, VTA
277 astrocyte activation by CNO increased the bursting discharges of all putative DA neurons
278 that persisted for at least 10 min (Fig. 5c, e-f) and it also increased the overall firing rate in
279 5 out of 7 DA neurons (Fig. 5g, h). In contrast, the firing mode of DA neurons in tdTomato-
280 expressing mice was unaffected by CNO, in terms of percentage of spikes in bursts and

281 firing rate (Fig. 5d-h). These *in vivo* data show that VTA astrocytes exert a direct control on
282 DA neuron firing activity.

283 We then asked whether the selective activation of VTA astrocytes, that increases the
284 burst firing of VTA DA neurons, could influence DA-dependent physiological functions. We
285 focused on motor activity, which is known to be controlled by DA. Notably, recent studies
286 revealed that DA neurons of the VTA, and not those of the Substantia Nigra (SN), play a
287 major role in the induction of motor hyperactivity^{3,33,34}. Locomotion in an open field arena
288 was tested in male mice given bilateral VTA injections of AAV9-GFAP-hM3D(Gq)-mCherry
289 (hM3D) or AAV8-GFAP-GFP (GFP) (Fig. 6a, b). Thirty min after intraperitoneal injections,
290 CNO induced a locomotor hyperactivity in hM3D-injected mice as compared to GFP control
291 mice (Fig. 6c-e). The time spent at the center was similar in the two groups (Fig. 6f),
292 suggesting no major effects on anxiety-like phenotypes. Interestingly, the sustained
293 locomotor hyperactivity was still observed in hM3D mice 48 hrs after the CNO treatment
294 (Fig. 6c-e). Overall, these *in vivo* data show that activation of astrocytes in the VTA
295 enhances DA neuron firing activity and induces long-lasting motor hyperactivity.

296

297

298 **DISCUSSION**

299 The present study shows that astrocytes of the VTA, a key area in DA-dependent functions,
300 respond with Ca²⁺ elevations to eCBs and DA released by DA neurons and, in turn, induce
301 a long-lasting potentiation of glutamatergic synapses onto adjacent DA neurons. The
302 induction of this novel form of astrocyte-mediated plasticity is independent on NMDAR
303 activation and requires astrocytic Ca²⁺ elevations, presynaptic mGluR1 activation and NO
304 signalling. We also show that astrocyte activation *in vivo* increases the burst and the overall
305 firing rate of DA neurons and induces locomotor hyperactivity. These results indicate that

306 astrocytes play a key role in the modulation of VTA DA neuron circuits that control DA-
307 dependent physiological functions.

308 Astrocytes have been shown to respond with Ca^{2+} elevations to synaptic
309 neurotransmitters and release gliotransmitters that contribute to sensory information
310 processing and behavioral responses^{13–15,35–38}. In different brain regions, including
311 hippocampus, dorsal striatum and neocortex, eCBs released by neurons at the somato-
312 dendritic level also recruit astrocytes evoking CB1R-mediated Ca^{2+} elevations and
313 gliotransmitter release that modulates synaptic transmission^{18–20,39}. The present results
314 show that in the VTA, activation of both CB1 and D2Rs is required for astrocytic Ca^{2+}
315 elevations in response to DA neuron bursts and that these Ca^{2+} elevations in astrocytes are
316 crucial for bLTP generation. This is based on the following observations: 1) CB1 and D2Rs
317 are expressed and closely localized, in the same astrocyte; 2) astrocyte Ca^{2+} response to
318 DA neuron bursts and bLTP induction are abolished in the presence of specific antagonists
319 that block either the CB1 or the D2R; 3) DA neuron bursts fail to evoke bLTP following
320 deletion in VTA astrocytes of either the CB1 or the D2R; 4) bLTP could not be evoked after
321 the impairment of astrocytic Ca^{2+} elevations downstream the activation of CB1 and D2Rs;
322 5) bLTP can be rescued in young male mice upon selective chemogenetic activation of
323 astrocytes coupled to DA burst firing, even in the presence of the CB1 and D2R antagonists.
324 Notably, the results reported above in points 3-5, provide evidence that activation of
325 neuronal CB1 and D2Rs is not required for bLTP induction.

326 It is worth to further comment on results reported in point 3. In our experiments on
327 mice carrying the “floxed” CB1 or D2 gene and injected in the VTA with AAV9/2-hGFAP-
328 mCherry_iCre, we observed that the great majority of Cre-positive cells were astrocytes and
329 only about 5% were neurons (Extended Data Fig. 7). However, this approach may lead to
330 undetectable expression levels of the Cre recombinase and result in CB1 or D2R deletion
331 in a higher percentage of neurons⁴⁰. It is noteworthy, however, that neuronal CB1 and D2Rs

332 in the VTA are inhibitory. Indeed, activation of presynaptic CB1 or D2Rs inhibits excitatory
333 transmission onto VTA DA neurons^{9,16} and activation of postsynaptic D2Rs induces a
334 hyperpolarization that reduces VTA DA neuron excitability⁹. Furthermore, D2R activation in
335 the VTA favors eCB-induced suppression of excitation¹⁶. These well-established inhibitory
336 actions of neuronal CB1 and D2Rs in VTA circuitry are not consistent with the CB1 and
337 D2R-dependent bLTP that we describe here and further support that activation of astrocytic
338 rather than neuronal CB1 and D2Rs is required for bLTP generation.

339 Recent studies reported that astrocytes in different brain regions, including the VTA,
340 express D1 and D2-type receptors and respond to bath applied DA stimuli with complex
341 Ca^{2+} dynamics including regulation of basal cytosolic Ca^{2+} and repetitive Ca^{2+} transients^{41–}
342 ⁴⁴. Most interestingly, astrocytes in the NAc respond to synaptically released DA with D1R-
343 rather than D2R-mediated Ca^{2+} elevations⁴⁴. These data confirm that astrocytes of different
344 brain regions and synaptic circuitries express different receptors that match the specific
345 signals generated by distinct neuronal activities¹³. Consistent with this view, through the low
346 affinity D1R, NAc astrocytes can sense the transient, high DA concentrations generated by
347 synaptic release of DA from VTA DA projections³². Conversely, through the high affinity
348 D2R³², VTA astrocytes can sense the lower DA concentrations mainly generated in the VTA
349 by somatodendritic rather than synaptic release and thus be functional targets of DA volume
350 transmission⁴⁵.

351 An additional specificity of VTA astrocytes is that a cooperativity between CB1 and
352 D2Rs is necessary for the Ca^{2+} response to DA neuron bursts, being activation of either
353 CB1 or D2Rs alone insufficient to induce astrocytic Ca^{2+} elevations. Our EM immunogold
354 experiments provide an ultrastructural background for this cooperativity revealing that CB1
355 and D2Rs are expressed in the same astrocytes, closely localized at astrocytic processes.
356 Quantitative analysis from CB1/D2R double-labelled astrocytic processes also reveals that
357 a group of couples exhibits an edge-to-edge separation ≤ 50 nm, which suggests physical

358 interactions between these two receptors and possible formation of heterodimers. Notably,
359 previous studies reported that D2 and CB1R coactivation in neurons enhances the formation
360 of CB1/D2R heterodimers^{17,46}. Furthermore, we recently showed that coactivation of GABAb
361 and Somatostatin receptors in neocortical astrocytes confers signalling specificity between
362 different interneuron subtypes and astrocytes⁴⁷. A cooperativity between different G-protein
363 coupled receptors may, therefore, be a general functional feature in the astrocyte response
364 to neuronal signals. Overall, CB1/D2R-expressing astrocytes in the VTA are fine-tuned to
365 sense eCB and dopamine releasing bursting neurons and extent excitation to neighboring
366 DA neurons through lateral potentiation of glutamatergic transmission. These results provide
367 further evidence for circuit- and synapse-specificity of neuron-astrocyte reciprocal signalling
368 in the brain¹³.

369 The astrocyte-mediated bLTP is absent in young male mice in which DA neuron
370 bursts fail to elevate Ca^{2+} in astrocytes. Our data suggest that this failure is mainly due to a
371 lower expression of astrocytic CB1Rs in young with respect to adult male mice showing
372 regular bLTP. Importantly, bLTP could be rescued in young male mice by coupling DA
373 neuron bursts with Ca^{2+} elevations evoked by CNO in hM3D-expressing astrocytes. These
374 results further support a crucial role of astrocytic Ca^{2+} signals in bLTP induction mechanism.
375 It is noteworthy that the expression of mGlu1R in excitatory terminals is also significantly
376 lower in young with respect to adult male mice, suggesting that a defective neuronal
377 response to astrocytic glutamate may also contribute to the absence of bLTP in young male
378 mice.

379 In hippocampus, transient enhancement of excitatory transmission induced by
380 astrocytic glutamate becomes a long-lasting event when coupled with neuronal NO
381 release¹⁹. A similar mechanism appears operative in the VTA where in the absence of NO
382 only a transient (6 min) enhancement of glutamatergic transmission was observed.

383 Therefore, NO released by burst firing DA neurons is required for the sustained phase of
384 bLTP.

385 Astrocytic Ca^{2+} signalling is required for bLTP generation in both young female and
386 adult mice. However, while the mechanism downstream the astrocyte Ca^{2+} signaling
387 remains similar, $\text{IP}_3\text{R2}$ deletion in adult mice is insufficient to abolish bLTP. This finding
388 reveals an increased complexity in the regulatory mechanisms of astrocytic Ca^{2+} dynamics
389 during development with contribution of signalling pathways other than the $\text{IP}_3\text{R2}$ -mediated
390 pathway^{27,48}. From these observations it also follows that negative results on the role of
391 astrocytic Ca^{2+} signaling in $\text{IP}_3\text{R2}^{-/-}$ adult mice must be interpreted with caution.

392 Transient and/or persistent potentiation of glutamatergic synapses, fundamentally
393 mediated by NMDAR activation, regulate the burst firing mode of VTA DA neurons that plays
394 a pivotal role in DA-dependent behaviors^{1,32}. The novel form of astrocyte-mediated
395 potentiation described here may integrate with these other forms of NMDAR-dependent
396 plasticity that favor the burst firing of DA neurons. Consistent with this view, we report that
397 *in vivo* astrocyte activation enhances DA neurons bursts and leads to a long-lasting
398 locomotor hyperactivity that recent studies revealed to depend on VTA rather than SN DA
399 neuron activity^{3,33,34}. Present results also suggest that activation of astrocytes by burst firing
400 DA neurons and the consequent lateral potentiation of glutamatergic synapses, may
401 represent a strategy used by individual DA neurons to expand the burst firing mode to
402 neighboring DA neurons. Hence, it is possible that, through the fundamental recruitment of
403 astrocytes, an isolated, high bursting DA neuron favors the formation of spatially defined
404 clusters of coactive DA neurons that have been proposed to convey essential information
405 about a specific subset of behavioral variables to target regions⁴⁹. Further specifically
406 designed experiments are necessary to validate this hypothesis.

407 Present results show that in the VTA circuitry, astrocyte signaling induces a long-
408 lasting potentiation of glutamatergic synapses to DA neurons, increases the burst firing

409 mode of DA neurons and favors locomotor hyperactivity thereby revealing an astrocyte-
410 mediated mechanism in the control of DA neuron activity and DA-dependent behaviors. Our
411 study also paves the way to future investigations examining whether dysregulations of DA
412 neuron-astrocyte reciprocal communication within the VTA may contribute to the
413 development of psychiatry disease states including motivation disorders, psychiatric
414 disorders with a strong motor component, such as attention-deficit/hyperactivity disorder,
415 and drug addiction.

416

417

METHODS

Animals and brain slice preparation. Animal care, handling and procedures were carried out in accordance with National (D.L. n.26, March 14, 2014) and European Community Council (2010/63/UE) laws, policies, and guidelines, and were approved by the local veterinary service. Horizontal VTA slices (240 μ m) were obtained from both male and female C57BL/6J wild type (wt) mice and inositol 1,4,5-triphosphate-type 2 receptor (IP₃R2) knockout mice (IP₃R2^{-/-})⁵⁰, and from male CB1R⁻⁵¹ and D2R-flox⁵² mice, at P14-17 (young mice) or P30-70 (adolescent/adult mice, to simplify adult mice). Animals were anesthetized with isoflurane, the brain removed and transferred into an ice-cold artificial cerebrospinal fluid (ACSF) containing (in mM): 125 NaCl, 2 KCl, 2 CaCl₂, 1 MgCl₂, 25 glucose, 25 NaHCO₃, 1.25 NaH₂PO₄, pH 7.4 with 95% O₂-5% CO₂. Slices were cut with a vibratome (Leica Vibratome VT1000S Mannheim, Germany) in the ice-cold solution described in Dugue et al. 2005⁵³ containing (in mM): 130 KGluconate, 15 KCl, 0.2 EGTA, 20 HEPES, 25 glucose, 2 Kynurenic acid. Slices were then transferred for 1 minute in a room-temperature solution containing (in mM): 225 D-mannitol, 2.5 KCl, 1.25 NaH₂PO₄, 26 NaHCO₃, 25 glucose, 0.8 CaCl₂, 8 MgCl₂, with 95% O₂-5% CO₂. Finally, slices were transferred in ACSF at 32°C for 15-20 minutes and then maintained at room temperature for the entire experiment.

Electrophysiological recordings and extracellular stimulation. Brain slices were continuously perfused in a submerged chamber with recording solution containing (in mM): NaCl 120; KCl 2; NaH₂PO₄ 1; NaHCO₃ 26; MgCl₂ 1; CaCl₂ 2; glucose 10; pH 7.4 (with 95% O₂-5% CO₂). Picrotoxin (50 μ M) was added to block GABA_A receptor currents. When indicated, other antagonists were bath applied with the recording solution. Cells were visualized with an Olympus FV1000 microscope (Olympus Optical, Tokyo, Japan). Conventional VTA DA neurons were recorded in the lateral part of the region medial to the

445 medial terminal nucleus of the accessory optical tract (mt, Fig. 1)^{5,54}. DA neurons from the
446 lateral VTA were identified on the basis of their distinct morphology characterized by a large
447 and elongated soma with no particular dendritic orientation and the presence of the following
448 electrophysiological properties: a low-frequency tonic firing, a large I_h current elicited by
449 hyperpolarizing steps¹⁶ and a slow depolarizing potential during current step injections⁵⁵
450 (Extended Data Fig. 1 b-d). Simultaneous electrophysiological whole-cell patch-clamp
451 recordings from two DA neurons (distance of the somata, 70-120 μ m) were made. Patch
452 electrodes (resistance, 3-4 M Ω) were filled with an internal solution containing (in mM): K-
453 gluconate, 135; KCl, 70; Hepes, 10; MgCl₂, 1; Na₂ATP 2 (pH 7.4 adjusted with KOH, 280-
454 290 mOsm/L). Recordings were obtained using a multiclamp-700B amplifier (Molecular
455 Device, Foster City, CA, USA). Signals were filtered at 1 kHz and acquired at 10 KHz
456 sampling rate with a DigiData 1440A interface board and pClamp 10 software. Series and
457 input resistances were monitored throughout the experiment using a 5 mV pulse.
458 Recordings were considered stable when the change of series and input resistances were
459 below 20%. Cells that do not meet these criteria were discharged. Theta capillaries filled
460 with recording solution were used for bipolar stimulation. To stimulate glutamatergic
461 afferents, electrodes were connected to an S-900 stimulator through an isolation unit and
462 placed 100-200 μ m rostral to the recording electrode (Extended Data Fig. 1 a). Paired pulses
463 (50 ms intervals) were delivered at 0.33 Hz. Excitatory post-synaptic currents (EPSCs) were
464 recorded while holding the membrane potential at -70 mV. Stimulus intensity was adjusted
465 to evoke 30-50 % maximal EPSCs. The EPSC amplitude was measured as the peak current
466 amplitude (2-9 ms after stimulus) minus the mean baseline current (100 ms before stimulus).
467 To illustrate the mean EPSCs time course, values were grouped in 3-min bins (i. e. mean
468 EPSCs from 60 stimuli). Changes in mean EPSCs in the first DA neuron were monitored
469 after imposing a burst or a tonic firing pattern to the second DA neuron (70-120 mm apart).
470 Burst firing pattern was imposed in current-clamp mode, through injections of intracellular

471 current pulses, with 5-pulse 20 Hz burst, every 500 ms for 5 min (Extended Data Fig. 1e)²³.
472 Tonic firing was imposed with individual current pulses applied at 2 Hz for 5 minutes²³
473 (Extended Data Fig. 1e). During the burst/tonic firing the extracellular stimulation was
474 switched off. In electrophysiological experiments, time 0 indicates the end of the burst/tonic
475 firing. For statistical analysis of short-term effects, mean EPSCs from 120 stimuli applied
476 before (basal) and mean EPSCs from 60 stimuli applied during the first 3 min after the firing
477 protocol (burst or tonic firing pattern) were compared. For statistical analysis of long-term
478 effects, mean EPSCs from 120 stimuli applied before (basal), 24-30 min (indicated as 30
479 min time point in the bar chart) or 39-45 min (indicated as 45 min time point in the bar chart)
480 after the firing protocol were compared. In young adult mice only the long-term effect at the
481 time point of 30 min was analyzed due to the difficulty of obtaining long-lasting recordings
482 in tissues from these mice. Paired-pulse ratio (PPR) was calculated as 2nd EPSC/1st EPSC
483 and evaluation of the PPR before and after the burst firing protocol was used to identify the
484 pre- or postsynaptic locus of the bLTP.

485

486 **Dye loading and Ca²⁺ imaging experiments.** Slices were loaded with the astrocyte specific
487 marker Sulforhodamine 101 (SR101) (0.3 μ M, Sigma Aldrich, Milano) in ACSF at 32°C for
488 15 minutes. Then, slices were loaded for 45 min at room temperature with the Ca²⁺ sensitive
489 dye Fluo4-AM (7,6 μ M, Life Technologies, Monza, IT) in an ACSF solution containing
490 pluronic F-127 (0.0067%, Sigma Aldrich, Milano, IT) and bubbled with 95% O₂-5% CO₂.
491 Ca²⁺ imaging experiments were conducted with a confocal laser scanning microscope TCS-
492 SP5-RS (Leica Microsystems, GmbH, Wetzlar, Germany) equipped with two solid state
493 lasers tuned at 448 nm and 543 nm (to image Fluo-4 and SR101 fluorescence, respectively)
494 and a 20x objective (NA, 1.0). Images were acquired with a 0.5 Hz frame rate for 90s, with
495 time intervals of 5 min between recordings. Image sequences were processed with ImageJ.
496 Regions of interest (ROIs) were drawn around cellular somata using the red SR101 signal.

Ca²⁺ events were estimated as changes of the Fluo-4 fluorescence signal over baseline ($\Delta F/F_0 = (F(t)-F_0)/F_0$). A fluorescence increase was considered a significant event when it exceeded two times the standard deviation from the baseline. Astrocyte Ca²⁺ responses were quantified by analyzing the probability of occurrence of Ca²⁺ spike by detecting the onset of Ca²⁺ elevations (Ca²⁺ spikes) during the recording period. To investigate the astrocyte response to the burst firing of DA neurons, a DA neuron was patch with an intracellular solution (see details before) containing the fluorescent tracer Neurobiotin 488 (65 μ M, Vector Laboratories, Inc, USA) to visualize neuronal soma and dendrites. To obtain the time course of the Ca²⁺ spike probability index reported in figures 1 and 2, the number of astrocytic Ca²⁺ spikes for each recording period was divided by the number of SR101-positive astrocytes in proximity (around 50 μ m) to Neurobiotin 488-filled DA neuron soma and dendrites. After three basal recordings, a burst firing pattern was imposed to the DA neuron (in current-clamp mode, through injections of intracellular current pulses, with 5-pulse 20 Hz burst, every 500 ms for 5 min) and the quantification of the Ca²⁺ spike probability was resumed 4.5 min after the initiation of the burst firing. In Ca²⁺ imaging experiments, time 0 indicates the start of the burst firing. For statistical analysis, a mean value of the Ca²⁺ spike probability/min per slice was calculated at basal conditions (mean of the three basal recordings) and after DA neuronal burst firing (mean of 4 consecutive recordings after the burst firing, the first at a time point of 4.5 min after the burst firing and the last at a time point of 24 min after the burst firing). When indicated, Ca²⁺ imaging experiments were performed in slices expressing the genetically encoded Ca²⁺ indicator GCaMP6f and the Gq protein-coupled designer receptor exclusively activated by designer drugs (DREADDs) hM3D in astrocytes (for AAVs delivery details see below). In these experiments, Ca²⁺ elevations were evoked by bath perfusion of the hM3D agonist clozapine N-oxide CNO (10 μ M) 2.5 min after the start of the burst firing. A mean time course of the Ca²⁺ response to CNO was calculated

522 by plotting the $\Delta F/F_0$ of a ROI drawn around the entire recording field. Then, plots were
523 aligned for the Ca^{2+} peak to calculate the mean time course of the Ca^{2+} response to CNO.
524

525 **AAVs delivery.** We bilaterally injected in the VTA of C57BL6J wild-type male mice, at
526 postnatal days 0-2 or 28-30, the viral vector ssAAV-9/2-hGFAP-hM3D(Gq)_mCherry-
527 WPRE-hGHp(A) (VVF-UZH, Zurich, Switzerland, 4.6×10^{12} viral genomes (vg)/ml), that
528 contains the astrocytic promoter to selectively express in astrocytes the mCherry-tagged,
529 Gq-coupled Designer Receptors Activated Only by Designer Drugs (DREADD) hM3D. Two-
530 three weeks after injection, we performed electrophysiological experiments in brain slices
531 from juvenile mice (P14-17) and in vivo single unit recordings in adult mice (P45-50). As
532 control for single unit recording experiments, in the VTA of wild-type male mice we bilaterally
533 injected the viral vector AAV5.GfaABC1D.cyto-tdTomato.SV40 (Addgene, USA, 2.4×10^{13}
534 vg/ml), that carries the astrocytic promoter GfaABC1D to express selectively in astrocytes
535 the tdTomato marker. In a group of P0-P2 mice, both ssAAV-9/2-hGFAP-
536 hM3D(Gq)_mCherry-WPRE-hGHp(A) and AAV5.GfaABC1D.cytoGCaMP6f.SV40
537 (Addgene, USA, 1.81×10^{13} genome copies/ml) viral vectors were injected together to
538 assess the Ca^{2+} responses evoked in astrocytes after activation of hM3D, using the Ca^{2+}
539 indicator GCaMP6f. To selectively express the Cre recombinase in astrocytes, injections of
540 the viral vector ssAAV9/2-hGFAP-mCherry_iCre-WPRE-hGHp(A) (VVF-UZH, Zurich,
541 Switzerland, 5.2×10^{12} vector genomes/ml) carrying the astrocytic promoter GfaABC1D
542 were bilaterally performed in the VTA of P 28-30 male mice containing the D2R or the CB1R
543 floxed gene. As control mice, the same viral vector was injected in age-matched C57BL6J
544 wild-type mice. Brain slice electrophysiological experiments were performed 4 weeks after
545 injection. AAV5.GfaABC1D.mCherry.hPMCA2w/b.SV40 (Penn Vector Core, USA, $1.14 \times$
546 10^{13} genome copies/ml) was bilaterally injected in the VTA of both female and male $\text{IP}_3\text{R}2^-$
547 $^-$ mice, at postnatal days 28-30, to selectively express in astrocytes the Ca^{2+} pump

hPMCA2w/b³¹. Brain slice electrophysiological experiments were performed 4 weeks after injection. AAV5.GfaABC1D.mCherry.hPMCA2w/b.SV40 was injected without previous dilution. The other viral vectors were diluted to 50% in ACSF before injection. When two viral vectors were injected, both vectors were present at a 1:1 ratio. The coordinates for viral injections were (in mm): AP 0.1, ML $\pm$ 0.15, DV -3.8 from lambda for P0-2 mice; AP -3.0, ML $\pm$ 0.5, DV -4.4 from Bregma for P28-30 mice. For injections in P0 mice, animals were anesthetized by hypothermia for three minutes and fixed into a modeled platform. Using a manually graduated pull glassed pipette, connected to a costume-made pressure injection system, we punch the skull bilaterally and injected a total volume of 350 nl containing the viral vector. After injection, the skin was sutured and mice were revitalized under a heat lamp before returning to their cage. For injections in P28-30 mice, animals were anesthetized with isoflurane (induction 4-5%, maintenance 1-2%). Depth of anesthesia was assured by monitoring respiration rate, eyelid reflex, vibrissae movements, and reactions to pinching the tail and toe. After drilling two holes into the skull over the VTA, we injected a total volume of 500 nl by using a pulled glass pipette connected to a peristaltic pump, at a rate of 100 nl/min. Pipette was kept for 10 minutes in the tissue before slow withdrawal. After injections, the skin was sutured and mice were revitalized under a heat lamp and returned to their cage.

566

Immunohistochemistry and cell counting. For the evaluation of the number of mCherry expressing astrocytes and neurons we prepared 70 μ m thick brain slices from young and adult animals injected with ssAAV-9/2-hGFAP-hM3D(Gq)_mCherry-WPRE-hGHp(A), ssAAV9/2-hGFAP-mCherry_iCre-WPRE-hGHp(A) or AAV5.GfaABC1D.mCherry.hPMCA2w/b.SV40. Mice were euthanized with 5% isoflurane and transcardially perfused with PBS followed by ice-cold 4% PFA in PBS. Brains were removed and postfixed overnight at 4°C in the same fixative solution. Horizontal brain slices were obtained with a VT1000S

574 vibratome (Leica), collected as floating sections and blocked for 1 hr in the Blocking Serum
575 (BS: 1% BSA, 2% goat serum and 1% horse serum in PBS) and 0,2% TritonX-100. After
576 blocking, sections were incubated (overnight at 4°C) with the following primary antibodies in
577 BS plus 0,02% Tx-100: anti-NeuN (RRID:AB_2298772, 1:200 mouse,
578 Thermofisher_Millipore MAB377), anti-glial fibrillary acidic protein (GFAP,
579 RRID:AB_10013382, 1:400 rabbit, Dako Agilent , Z0334), anti-S100B (RRID:AB_2315306,
580 1:400 rabbit, Dako Agilent, Z031129), anti-glutamate transporter 1 (GLT1, RRID:AB_90949,
581 1:400 guinea pig, Abcam, AB1783); prefixed in 50% methanol in PBS per 15 minutes). After
582 washing with PBS, slices were incubated for 2 hr at room temperature with secondary
583 antibodies conjugated with AlexaFluor-488 (1:500; A21202 donkey anti-mouse; A21206
584 donkey anti-rabbit; A11073 goat anti-guinea pig, Invitrogen Thermo-Scientific). To evaluate
585 mCherry positive cells, we directly evaluate the red fluorescence of infected slices. Only for
586 IP₃R2^{-/-} mice injected with AAV5.GfaABC1D.mCherry.hPMCA2w/b.SV40, we performed
587 double immunofluorescence of NeuN (or GLT1) together with anti-RFP (RRID:AB_2209751,
588 1:1000 rabbit, Rockland , 600-401-379). In this case, secondary antibodies were anti-mouse
589 (or anti-guinea pig) AlexaFluor-488 conjugated together the anti-rabbit AlexaFluor-555
590 conjugated (A21430 donkey anti-rabbit, Invitrogen Thermo-Scientific, 1:500). Slices were
591 then washed and nuclei were stained with Top-Ro3 (Invitrogen Thermo-Scientific, 1:1000).
592 Negative controls were performed in the absence of the primary antibodies. We used a TCS-
593 SP5-RS laser scanning microscope (Leica, Germany; 20x NA1x/W objective) to acquire
594 sequential three channels, confocal image z-stacks (1 µm z-step, 456,33x456,33 µm) and
595 ImageJ for double-labeled cell counting. We counted mCherry-positive cells and then we
596 evaluated the percentage of mCherry-positive cells that were neurons (mCherry+/NeuN+)
597 or glial cells. VTA from both hemispheres of injected animals was evaluated in at least two
598 mice for each group.

599

600 **Pre- and post-embedding electron microscopy.** Animals. Seven P16 and four P50
601 C57BL/6 mice (3 females, 4 males for P16; 2 females and 2 males for P50) were used. Mice
602 were anesthetized with an intraperitoneal injection of chloral hydrate (300 mg/kg) and
603 perfused transcardially with a flush of saline solution followed by 4% freshly depolymerized
604 paraformaldehyde and 0.2% glutaraldehyde in 0.1 M phosphate buffer (PB; pH 7.4). Brains
605 were removed, post-fixed in the same fixative (for 48 hrs), and cut on a Vibratome in 50 μ m
606 serial horizontal sections from the midbrain which were collected in PB until processing⁵⁶.
607 Horizontal sections were through the dorso-ventral extent of ventral tegmental area (VTA)
608 resulting in 7-8 sections per series. In order to verify the dorso-ventral extension of VTA, a
609 pilot series of sections from a male mouse, were immuno-processed for tyrosine-
610 hydroxylase (primary antibodies from Millipore, AB1542, RRID:AB_90755⁵⁷; 1:500) and for
611 Nissl staining. In immunoreacted sections (see data collection and analysis) lateral VTA was
612 identified as the region medial to the medial terminal nucleus of the accessory optical tract^{5,54}
613 (MT). Information on antibodies used for electron microscopy are reported in Table 1.

614 Immunoperoxidase and pre-embedding procedures. Sections were treated with H₂O₂
615 (1% in PB; 30 min) to remove endogenous peroxidase activity, rinsed in PB and pre-
616 incubated in 10% normal goat serum (NGS, 1 hr; for mGLUR1 α , mGluR1 β , D2, D3, D4) or
617 in 10 % normal donkey serum (NDS, 1 hr; for CB1). Sections were then incubated in a
618 solution containing primary antibodies (see Table 1 for dilutions; 2 hrs at room temperature
619 [RT] and overnight at 4°C). The following day, sections were rinsed 3 times in PB and
620 incubated first in 10% NGS or 10% NDS (15 min) and then in a solution containing
621 secondary biotinylated secondary antibodies (see Table 1 for dilutions; 1.5 hr at RT).
622 Sections were subsequently rinsed in PB, incubated in avidin-biotin peroxidase complex
623 (ABC Elite PK6100, Vector), washed several times in PB, and incubated in
624 3,3'-diaminobenzidine tetrahydrochloride (DAB; 0.05% in 0.05 M Tris buffer, pH 7.6 with
625 0.03% H₂O₂). Method specificity was verified by substituting primary antibodies with PB or

626 NGS. As previously described⁵⁶ after completion of immunoperoxidase procedures,
627 sections were post-fixed in 1% osmium tetroxide in PB for 45 m and contrasted with 1%
628 uranyl acetate in maleate buffer (pH 6.0; 1 h). After dehydration in ethanol and propylene
629 oxide, sections were embedded in Epon/Spurr resin (Electron Microscopy Sciences,
630 Hatfield, PA, USA), flattened between Aclar sheets (Electron Microscopy Sciences) and
631 polymerized at 60°C for (48 hrs). Chips including lateral VTA were selected by light-
632 microscopic inspection, glued to blank epoxy and sectioned with an ultramicrotome (MTX;
633 Research and Manufacturing Company Inc., Tucson, AZ, USA). The most superficial
634 ultrathin sections (~60 nm) were collected and mounted on 300 mesh nickel grids, stained
635 with Sato's lead and examined with a Philips EM 208 and CM10 electron microscopes
636 coupled to a MegaView-II high resolution CCD camera (Soft Imaging System). To minimize
637 the effects of procedural variables, all material from P16 and P50 females and males was
638 processed in parallel.

639 Post-embedding procedures. Sections were processed for an osmium-free
640 embedding method⁵⁸. Dehydrated sections were immersed in propylene oxide, infiltrated
641 with a mixture of Epon/Spurr resins, sandwiched between Aclar films, and polymerized at
642 60 °C for 48 h. After polymerization, chips were cut from the wafers, glued to blank resin
643 blocks and sectioned with an ultramicrotome. Thin sections (60–80 nm) were cut and
644 mounted on 300 mesh nickel grids and processed for immunogold labeling^{58,59}. In brief, after
645 treatment with 4 % para-phenylenediamine in Tris-buffered saline [0.1 M Tris, pH 7.6, with
646 0.005 %Tergitol NP-10 (TBST)], grids were washed in TBST (pH 7.6), transferred for 15 min
647 in 0.25% NDS in TBST (pH 7.6) and then incubated overnight (26°C) in a solution of TBST
648 (pH 7.6) containing a mixture of anti-D2 and anti-CB1 primary antibodies (see Table 1 for
649 dilutions). Grids were subsequently washed in TBST (pH 8.2), transferred for 10 min in 0.5%
650 NDS in TBST (pH 8.2), incubated for 2 hrs (26°C) in TBST (pH 8.2) containing secondary
651 antibodies conjugated to 18 and 12 nm gold particles, washed in distilled water, and then

652 stained with uranyl acetate and Sato's lead. The optimal concentration of antibodies to D2
653 and CB1Rs was sought by testing several dilutions; the concentration yielding the lowest
654 level of background labeling and still immunopositive elements was used to perform the final
655 studies. Gold particles were not detected when primary antiserum was omitted. When
656 normal serum was substituted for immune serum, sparse and scattered gold particles were
657 observed, but they did not show any specific relationship to subcellular compartments.

658 Data collection and analysis. All data were obtained from lateral VTA of
659 immunoreacted sections^{5,54}.

660 For pre-embedding electron microscopy, mGluR1 α , mGluR1 β , CB1, D2, D3, D4R
661 immunoreactive profiles were studied in ultrathin sections from the surface of the embedded
662 blocks. Quantitative data derived from the analysis of microscopic fields of lateral VTA (10–
663 12 ultrathin sections/animal) selected and captured at original magnifications of 12,000x–
664 30,000x. Microscopical fields from females and males containing positive processes were
665 randomly selected. Acquisition of microscopical fields and analysis of female and male mice
666 were performed under blinded conditions.

667 For the analysis of the distribution of mGluR1 α , mGluR1 β , CB1, D2, D3, D4R positive
668 profiles, subcellular compartments were identified according to well-established criteria⁶⁰
669 (Extended Data Figures and Tables 1 and 2). For quantifying mGluR1 α or mGluR1 β in P16
670 VTA, and mGluR1 β in P50 VTA at axon terminals, synapses exclusively characterized by a
671 presynaptic terminal with clear and round vesicles nearby the presynaptic density, a synaptic
672 cleft displaying electron dense material, pre- and postsynaptic membranes defining the
673 active zone and the post-synaptic specialization, and finally by a prominent postsynaptic
674 density the asymmetric synapses^{60,61} were sampled (axon terminals making asymmetric
675 synaptic contacts containing one or more dense core vesicles more likely representative of
676 co-release of glutamate and others neurotransmitters^{62–64} were not included in this group;
677 Tables 2 and 4).

678 For quantifying CB1, D2, D3, D4R at astrocytic processes in P16 VTA, and CB1 and
679 D2R in P50 VTA, astrocytic profiles were identified based on their typical irregular outlines
680 and the paucity of cytoplasmic components (with the exception of ribosomes, glycogen
681 granules and various fibrils⁶⁰. For post-embedding electron microscopy, ultrathin sections
682 (20 ultrathin sections/animal) were examined at 50,000–85,000x and fields that included at
683 least 1 double immunolabeled astrocytic profile were selected. For determining the relative
684 density of D2 and CB1Rs at the membranes of double labeled astrocytic profiles, pyramidal
685 cell nuclei were also identified: gold particles within labeled structures were counted and
686 areas were calculated using ImageJ (NIH, Bethesda, MD, USA). Background was calculated
687 by estimating labeling density over pyramidal cell nuclei^{59,65}. Particle densities were counted
688 and compared with background labeling. Gold particles were considered associated with
689 plasma membrane if they were within 20 nm of the extracellular side of the membrane. To
690 determine the degree of nearness of D2 and CB1R at the membrane of double-labeled
691 profiles the edge-to-edge distances between immunogold labeled D2 and CB1R were
692 measured along the membrane using ImageJ and the distribution of the separation
693 distances between D2 and CB1R was determined^{47,59,66–69}. In the cases in which multiple
694 paths connecting particles gave different inter-distance values, the shortest inter-distance
695 was selected and used for distribution analysis. Given that gold particles with edge-to-edge
696 separation distance ≤ 50 nm are highly suggestive of physical interactions of two detected
697 proteins (i.e., a physical coupling complex^{47,59,66–69}) distribution analysis of the interdistance
698 between particles was based on bin of 50 nm.

699 For all microscopy data, normality tests and statistical analyses were performed using
700 GraphPad Prism Software v.7.0a (San Diego, CA, USA).

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706 Table 1. Primary and Secondary antibodies

Protein	Source	Primary antibody	Host	Characterized in	Dilution	Secondary antibody	Source	Dilution
mGLUR1 α	Frontier Institute	Rb-Af811 (RRID:AB_257179)	Rb	Ohtani et al. 2014 ^a	1:200	Biotinylated	Jackson	1:500
mGLUR1 β	Frontier Institute	Rb-Af250 (RRID: AB_2616586)	Rb	Ohtani et al. 2014 ^b	1:100	Biotinylated	Jackson	1:500
CB1	Frontier Institute	GP-Af530 (RRID: AB_2571593)	GP	Garcia-Ovejero et al. 2013 ^c	1:200 IP 1:25 IG	Biotinylated 18 nm gold	Jackson Jackson	1:500 1:20
D2	Millipore	AB5084P (RRID:AB_2094980)	Rb	Stojanovic et al. 2017 ^d	1:100 IP 1:10 IG	Biotinylated 12 nm gold	Jackson Jackson	1:500 1:20
D3	Alomone	ADR-003 (RRID:AB_2039830)	Rb	Solís et al. 2017 Castro-Hernandez et al. 2015 ^e	1:100	Biotinylated	Jackson	1:500
D4	Millipore	AB324405 (RRID:AB_564550)	Rb	Barili et al. 2000 ^f	1:1000	Biotinylated	Jackson	1:500

707
708
709 Abbreviations: IP, immunoperoxidase for pre-embedding electron microscopy; IG,
710 immunogold for post-embedding electron microscopy.

711
712 ^a Raised against a synthetic peptide corresponding to AA 945-1127 of mouse mGLUR1 α .
713 Immunoreactivity verified in mGluR1 knock-out mice⁷⁰.

714 ^b Raised against a synthetic peptide corresponding to AA 899-906 of mouse mGLUR1 β .
715 Immunoreactivity verified in mGluR1 knock-out mice⁷⁰.

716 ^c Raised against the C-terminal 31 AA of mouse CB1. Lack of immunoreactivity in CB1
717 knock-out mice⁷¹.

718 ^d Raised against a peptide corresponding to 28 AA within the third cytoplasmic loop of
719 human D2 receptor. Lack of immunoreactivity in D2 knock-out mice⁷².

720 ^e Raised against a synthetic peptide corresponding to AA 15-29 within the extracellular N-
721 terminus sequence of rat D3 receptor. Lack of immunoreactivity in D3 knock-out mice^{73,74}.

722 ^f Raised against a synthetic peptide corresponding to AA 176-185 of human D4 receptor.
723 Immunoreactivity totally abolished by preadsorption of D4 antibodies with immunogen
724 peptide⁷⁵.

726 ***In vivo* single unit recordings.** C57BL/6J wild-type male mice, injected 2-3 weeks before
727 with ssAAV-9/2-hGFAP-hM3D(Gq)_mCherry-WPRE-hGHp(A) or AAV5.GfaABC1D.cyto-
728 tdTomato.SV40, were anesthetized using chloral hydrate (400 mg/kg i.p.), supplemented as
729 required to maintain optimal anesthesia throughout the experiment, and placed in the
730 stereotaxic apparatus (Kopf, Germany). Their body temperature was maintained at 36 ± 1
731 °C using a feedback-controlled heating pad. For the placement of a recording electrode, the
732 scalp was retracted and one burr hole was drilled above the parabrachial pigmented nuclei
733 of the posterior VTA (AP: - 3.0-3.5 mm from bregma; L: 0.4-0.6 mm from midline; V: 4-5 mm
734 from the cortical surface) according to the Paxinos and Franklin atlas (2004). Extracellular
735 identification of putative DA neurons was based on their location as well as on the set of
736 electrophysiological features that characterize these cells *in vivo*: (1) a typical triphasic
737 action potential with a marked negative deflection; (2) an action potential width from start to
738 end > 2.5 ms; (3) a slow firing rate (< 10 Hz). VTA putative DA neurons were selected only
739 when all the already published criteria were fulfilled^{76–79}. Single unit activity of putative DA
740 neurons was recorded extracellularly using glass micropipettes filled with 2% Chicago sky
741 blue dissolved in 0.5 M sodium acetate (impedance 3-7 MΩ). An injection pipette (20-40 μm
742 in diameter attached ~150-100 μm above the recording tip) was used for simultaneous local
743 microinjection of CNO (1 mM). Signal was pre-amplified, amplified (Neurolog System,
744 Digitimer, UK), filtered (band-pass 500–5000 Hz), and displayed on a digital storage
745 oscilloscope. Experiments were sampled on- and off-line by a computer connected to CED
746 Power 1401 laboratory interface (Cambridge Electronic Design, Cambridge, UK) running
747 the Spike2 software (Cambridge Electronic Design, Cambridge).

748 Single units were isolated and the spontaneous activity recorded for a minimum of 3
749 min before local application of CNO (1 mM). A total volume of 30-100 nl was infused using
750 brief (10-100 ms) pressure pulses (40 psi, Picospritzer, Narishige, Japan). One injection
751 maximum per hemisphere was given. For statistical analysis, we calculated the mean firing

752 rate (number of spikes/s) and the percentage of spikes in burst (SiB), before and after CNO
753 application (in 2-min bins or in the 10 min of recording after CNO application). Bursts were
754 defined as the occurrence of two spikes at an inter-spike interval of < 80 ms, and terminated
755 when the inter-spike interval exceeded 160 ms⁸⁰. At the end of the experiment, negative
756 DC (15 mA for 5 minutes) was passed through the recording electrode to eject Pontamine
757 sky blue, which allowed the anatomical location of the recorded neuron. Mice were then
758 euthanized and brains were rapidly removed and fixed in 4% paraformaldehyde solution.
759 The position of the electrodes was identified with a microscope in coronal sections (100 μ m).
760 Only recordings in the correct area were considered for analysis.

761

762 **Behavioral test.** Viral Injection. C57BL/6J mice were naïve and two month-old at the time
763 of surgery. All mice were anesthetized with a mix of isoflurane/oxygen 2%/1% by inhalation
764 and mounted into a stereotaxic frame (Kopf). Brain coordinates of viral injections in the VTA
765 were chosen in accordance with the mouse brain atlas: AP: -3 mm; ML: $\pm$ 0.50 mm; DV: -
766 4.7 mm. The volume of AAV injection (AAV9-GFAP-hM3D(Gq)-mcherry or AAV8-GFAP-
767 GFP) was 100 nl per hemisphere. We infused virus through a glass micropipette connected
768 to a 10- μ L Hamilton syringe. After infusion, the pipette was kept in place for 6 min and then
769 slowly withdrawn.

770 Locomotor Activity. Mice were tested during first two hours of the dark phase in an
771 experimental apparatus consisting of 4 grey, opaque open field boxes (40x40x40 cm) evenly
772 illuminated by overhead lighting (5 ± 1 lux). Each session was video recorded with ANY-
773 maze tracking software (Stoelting Co.) for one hour. In the first day of locomotor activity all
774 animal received an injection of CNO (3mg/kg) 30 minutes before the beginning of the test;
775 48 hours later, the animals have been tested for a second time in the same apparatus with
776 a saline injection.

777

778 **Drugs.** Picrotoxin 50 μ M (SIGMA, Aldrich, Milano, IT); AM251 2-4 μ M (Abcam, Cambridge,
779 UK); eticlopride hydrochloride 1 μ M (Abcam, Cambridge, UK); SCH-23390 hydrochloride 10
780 μ M (Abcam, Cambridge, UK); D-AP5 50 μ M (Abcam, Cambridge, UK); LY-367385 100 μ M
781 (Abcam, Cambridge, UK); clozapine-N-oxide (CNO) 10 μ M (MedChemExpress, USA) and
782 L-741,626 10 μ M (Tocris, Bristol, UK) were bath applied. L-NAME 100 μ M (SIGMA, Aldrich,
783 Milano, IT) was included in the patch pipette.

784

785 **Data analysis.** Data analysis was performed with Clampfit 10.5, Origin 8.0 (Microcal
786 Software), Microsoft Excel 2010, ImageJ (NHI), Sigma Plot 11, ANY-maze tracking software
787 and GraphPad Prism 7.0a.

788

789 **Statistical analysis.** No statistical methods were used to predetermine sample size. Data
790 are expressed as mean $\pm$ standard error of the mean (SEM). Normality test (Shapiro-Wilk
791 test) was applied to the data before running statistical tests. Based on the normality test
792 result, data were analyzed using either parametric (two-tailed Student's t-test) or
793 nonparametric tests (Wilcoxon signed-rank test) as appropriate. For electron microscopy
794 data analysis, Mann-Whitney test and contingency Fisher's test were used. For *in vivo* single
795 unit recordings and behavioral tests, two-way RM ANOVA and Bonferroni's multiple
796 comparison test was used. Statistical differences were established with $p < 0.05$ (*), $p < 0.01$
797 (**), $p < 0.001$ (***) and $p < 0.0001$ (****).

798

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811

812 **AUTHOR CONTRIBUTIONS**

813 LMR, MGG and GC designed the study. LMR and MGG performed the electrophysiological
814 and Ca²⁺ imaging experiments in brain slices. AC performed the immunohistochemistry
815 experiments. FM, GP and FP performed the behavioral experiments. MC and ALM
816 performed the in vivo single unit recordings. MM, AP and FC performed the electron
817 microscopy experiments. GM provided the CB1-floxed mice. All authors discussed the
818 results. MGG and GC wrote the paper with inputs from all the authors.

819

820 **COMPETING INTERESTS**

821 The authors declare no competing interests.

822

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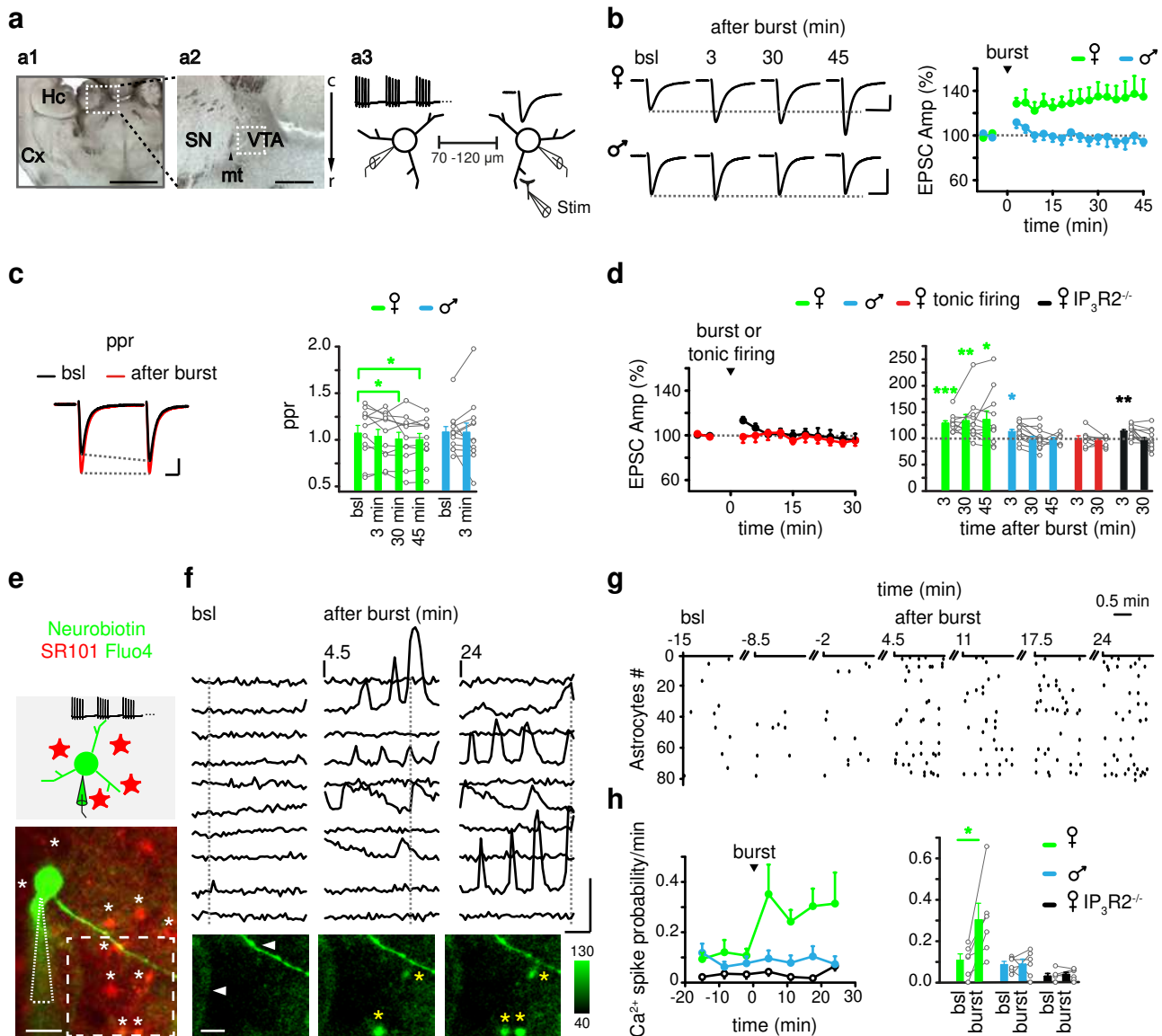


Figure 1. Astrocyte recruitment by DA neuron burst firing induces LTP of excitatory synapses onto adjacent DA neurons in young female mice. a) a1, low-magnification from a horizontal brain slice with a caudal-rostral orientation. Cx, cortex, Hc, hippocampus. Scale bar, 1 mm. a2, high magnification of the area indicated in a1. mt, medial terminal nucleus of the accessory optical tract; SN, substantia nigra (lateral to mt); VTA, ventral tegmental area (medial to mt), c, caudal; r, rostral. Dashed square, lateral part of the VTA where conventional DA neurons were recorded. Scale bar, 200 μ m. a3, schematic of the experimental design showing a DA neuron pair, two recording pipettes, the burst firing imposed to a DA neuron (left) and the EPSC evoked on the other DA neuron (right) by extracellular stimulation of rostral glutamatergic afferents (Stim). b) Left, evoked EPSCs at basal conditions (bsl) and at different time points after bursts, from slices of female (upper traces) and male (lower traces) mice. Stimulus artifacts were removed. Scale bars, 50 pA, 20 ms. Right, EPSC amplitude in female (n = 11) and male (n $\geq$ 6) mice after the burst firing protocol (arrowhead). In this and the other figures reporting EPSC amplitude vs time, t = 0 indicates the end of the burst firing. c) Left, representative pair of evoked EPSCs (time interval, 50 ms) from a DA neuron of a female mouse, before (bsl, black traces) and after bursts (red traces). Scale bars, 20 pA, 10 ms. Right, paired-pulse ratio (ppr) values vs time in female (n = 11) and male (n = 12) mice. d) Left, EPSC amplitude after tonic and burst firing protocol in wt female mice (n = 8) and $IP_3R2^{-/-}$ female mice (n = 12), respectively. Right, EPSC amplitude at time points indicated after burst/tonic firing in the different groups. e) Schematic and fluorescence image of a neurobiotin-488 filled DA neuron (dotted line, patch pipette) and SR101-loaded astrocytes (white asterisks). Scale bar, 30 μ m. **See next page.**

Figure 1. f) Upper panel, time course of Ca^{2+} levels from astrocytes shown in (e) at basal conditions, 4.5 min and 24 min after burst firing. Scale bars, 100 %, 20 s. Lower panel, Fluo-4 fluorescence images of the dashed square shown in e). Arrows, two DA neuron dendrites at different focal planes; yellow asterisks, astrocytes displaying Ca^{2+} transients (upper traces) at the time points indicated (dashed lines). Scale bar, 20 μm . g) Raster plot reporting the onset of Ca^{2+} transients from 84 astrocytes, at basal conditions and after DA neuron burst firing. In this and the other figures reporting the time course of the astrocytic Ca^{2+} spike probability/min, $t = 0$ indicates burst firing onset. h) Left, time course of astrocytic Ca^{2+} spike probability/min in female, male and $\text{IP}_3\text{R}2^{-/-}$ female mice ($n = 6$), before and after burst firing. Right, Ca^{2+} spike probability/min in basal conditions and after burst firing. In this and the other figures, data are represented as mean $\pm$ SEM; *, < 0.05 ; **, < 0.01 ; ***, < 0.001 ; ****, < 0.0001 .

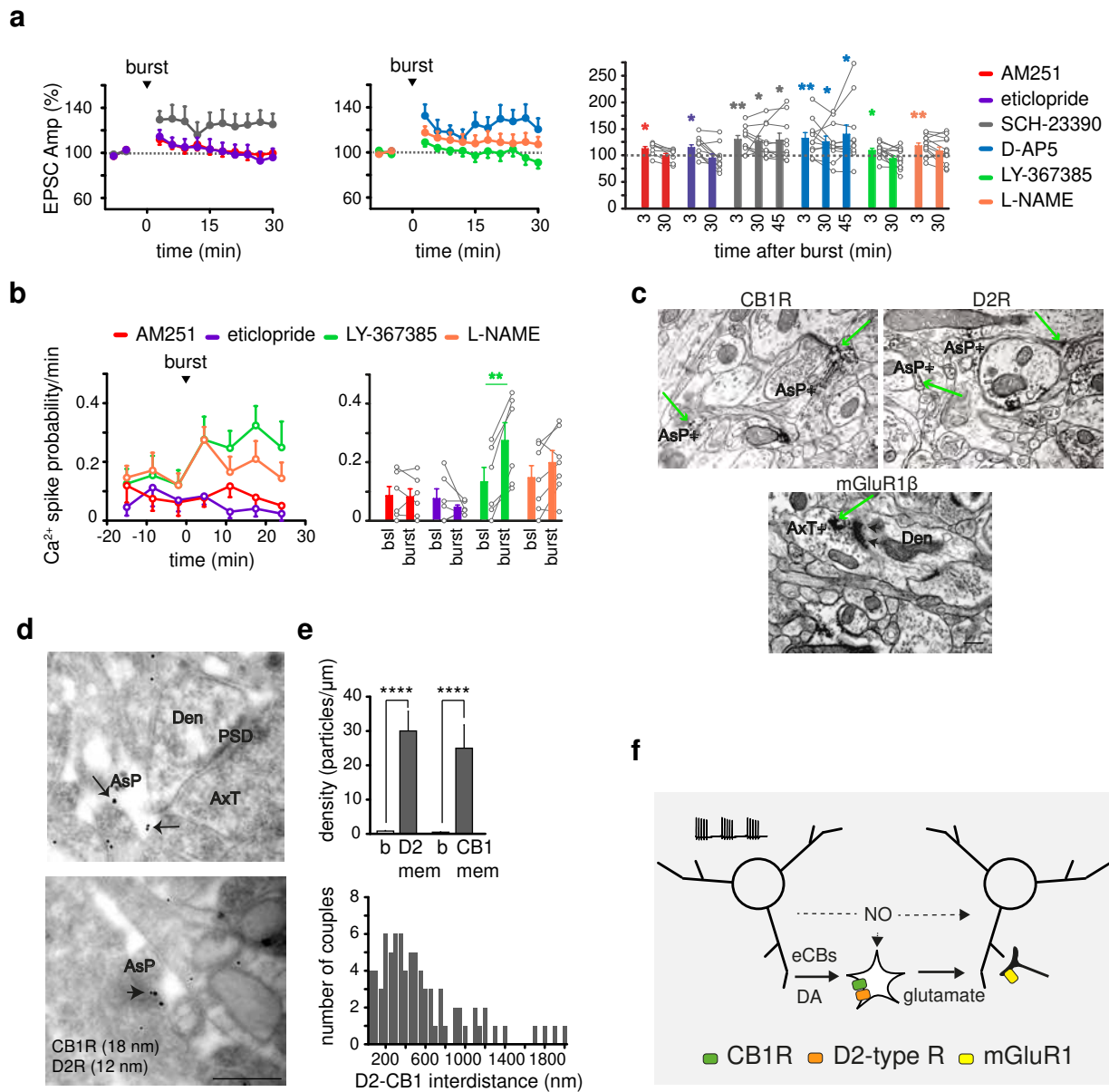


Figure 2. bLTP generation in female mice requires both eCB and DA signaling to astrocytes, mGluR1 activation and NO signaling. a) Time course and bar chart of EPSC amplitude in the presence of different antagonists (AM251 (CB1R), $n = 7$; eticlopride (D2-type R), $n = 10$; SCH-23390 (D1-type R), $n = 10$; D-AP5 (NMDAR), $n = 11$; LY-367385 (mGluR1), $n = 12$; L-NAME (NO synthase), $n = 12$). b) Time course and bar chart of astrocytic Ca²⁺ spike probability/min in the presence of antagonists that impaired bLTP generation ($n \geq 6$). c) Pre-embedding electron microscopy images from lateral VTA of a young female mouse of CB1, D2 and mGluR1 β receptors. Green arrows, immunopositive products in AsP (AsP+) and AxT (AxT+) forming asymmetric synaptic contacts (arrowheads) with a dendrite (Den). Scale bar, 300 nm. d) Post-embedding electron microscopy images of CB1/D2R immunogold double-labelled astrocytic processes (AsP) in lateral VTA (CB1R, 18 nm gold particles; D2R, 12 nm gold particles). Upper panel, a double-labelled AsP expressing CB1 and D2R (arrows) in close apposition to an asymmetric synapse (AxT, axon terminal; Den, dendrite; PSD, post-synaptic density). Lower panel, an edge-to-edge separation distance between these receptors ≤ 50 nm (arrowhead). Scale bar, 300 nm. e) Upper panel, CB1 and D2R immunogold densities at the membranes of astrocytic processes (AsP, $n = 79$ from two P16 females; 30.1 ± 2.6 gold particles/ μm^2 for D2Rs (D2 mem), and 24.9 ± 2.0 particles/ μm^2 for CB1Rs (CB1 mem)) and at neuronal nuclei (b, background values, $n = 10$; 0.82 ± 0.12 for 12 nm gold particles; 0.49 ± 0.07 for 18 nm gold particles, Mann-Whitney test). Lower panel, distribution of the edge-to-edge interdistances (bin, 50 nm) between membrane D2 and CB1R immunogold couples. f) Schematic of the proposed cellular and molecular mechanism for bLTP generation (see text for details).

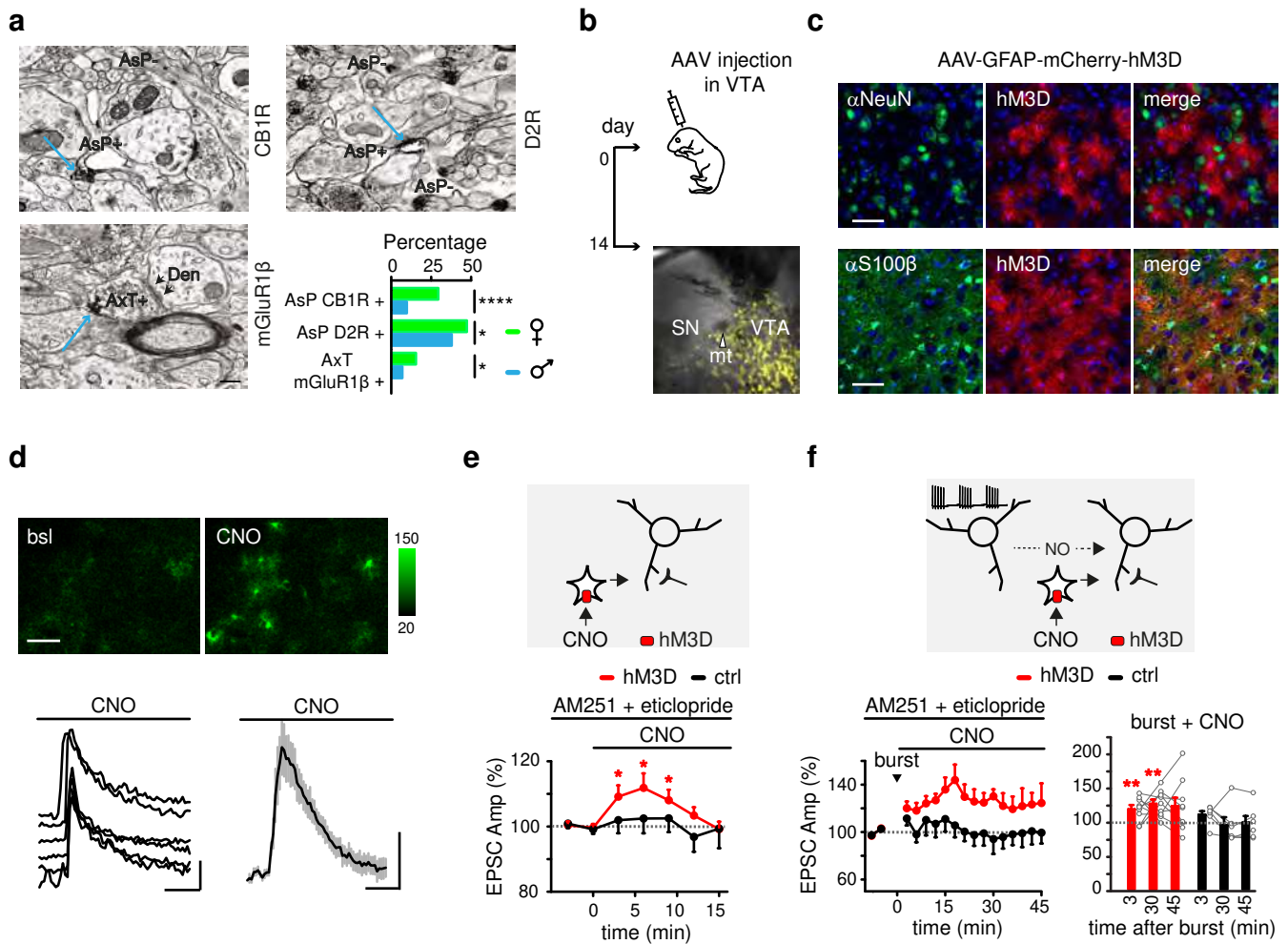


Figure 3. Chemogenetic selective activation of astrocytes rescues bLTP in young male mice. a) Representative pre-embedding electron microscopy in the lateral VTA from a young male mouse of CB1 and D2R expression at AsP and mGluR1β expression at AxT forming asymmetric synaptic contact (arrowheads) with dendrite (Den). Blue arrows, immunopositive products in AsP (AsP+) and AxT. AsP-, AsP without immunoreactivity. Scale bar, 300 nm. Lower panel, quantification and comparison (contingency Fisher's test) of CB1, D2 and mGlu1βR expression in female and male mice. b) Schematic of the AAV-9/2-hGFAP-hM3D(Gq)_mCherry-WPRE-hGHp(A) injection in the VTA of a neonatal male mouse and fluorescence image of a brain slice two weeks after injection (yellow, mCherry-hM3D expression). c) Confocal images of the VTA from a mouse injected with AAV-9/2-hGFAP-hM3D(Gq)_mCherry-WPRE-hGHp(A), showing the fluorescence of mCherry-hM3D (red), the nuclear Top-Ro3 (blue) and the specific green fluorescence for either neurons (α-NeuN staining) or astrocytes (α-S100β staining). Scale bars, 50 μm. d) GCaMP6f fluorescence images of astrocytes in basal conditions and after CNO perfusion. Scale bar, 50 μm. Lower panels, time course of Ca²⁺ elevations evoked by CNO in these astrocytes (scale bars, 100%, 30 s) and mean change of total Ca²⁺ levels in slices (n = 9) expressing GCaMP6f and hM3D in astrocytes in response to CNO (scale bars, 2%, 30 s). e) Upper panel, schematic of the experimental design. Lower panel, CNO transiently increases EPSC amplitude of DA neurons in male mice expressing hM3D in astrocytes (n = 9), but not in non-injected mice (n = 8). These experiments were performed in the presence of AM251 and eticlopride. f) Upper panel, schematic of the experimental design. Lower panel, pairing of burst firing protocol and CNO application partially rescues bLTP in male mice expressing hM3D in astrocytes (n = 9), but not in non-injected mice (n = 6). These experiments were performed in the presence of AM251 and eticlopride.

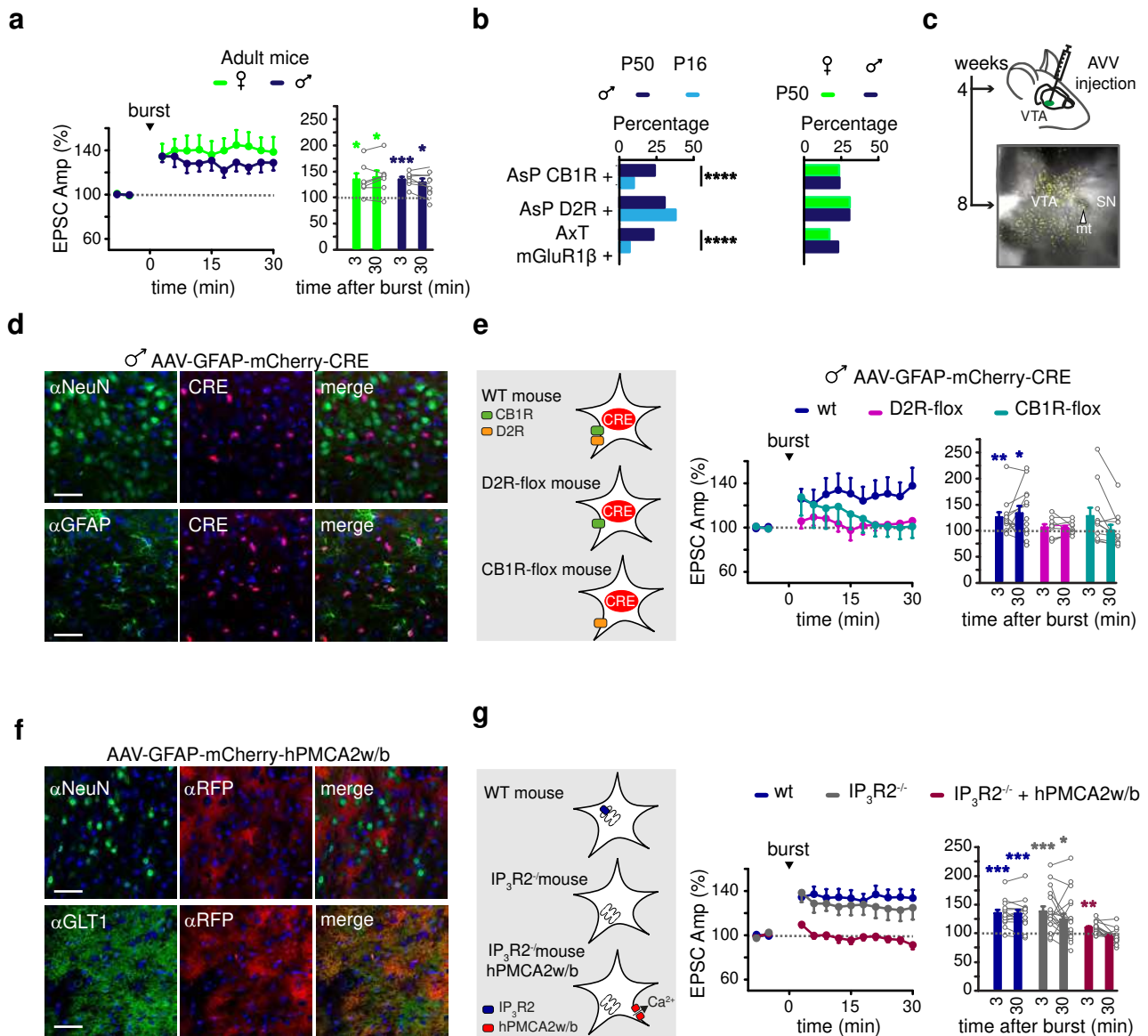


Figure 4. Astrocytes induce bLTP in both female and male adult mice and bLTP is abolished in adult IP₃R2^{-/-} mice expressing a plasmatic Ca²⁺ pump in astrocytes. a) The bLTP is present in both female (n = 7) and male (n = 7) adult mice. b) Quantification and comparison (contingency Fisher's test) of CB1, D2 and mGluR1β receptors expression in young and adult male mice and in adult female and male mice. c) Schematic of the AAV9-hGFAP-mCherry_iCre-WPRE-hGHp(A) injection in adult male mouse VTA and fluorescence image of a brain slice four weeks after viral injection (yellow, mCherry-Cre expression). d) Confocal images of the VTA from an adult male mouse injected with AAV9-hGFAP-mCherry_iCre-WPRE-hGHp(A), showing the fluorescence of mCherry-Cre (red), the nuclear Top-Ro3 (blue) and the specific green fluorescence for either neurons (α-NeuN staining) or astrocytes (α-GFAP staining). Scale bars, 50 μm. e) When male mice express the Cre recombinase in VTA astrocytes, bLTP is preserved in wt mice (n = 13), but abolished in D2R- (n = 9) or CB1R-Floxed (n = 11) mice. f) Confocal images of the VTA from an adult IP₃R2^{-/-} mouse injected with AAV5-GfaABC1D-mCherry-hPMCA2w/b.SV40, showing the expression of the Ca²⁺ pump hPMCA2w/b (α-RFP red staining), the blue nuclear Top-Ro3 staining and specific green staining for either neurons (α-NeuN staining) or astrocytes (α-GLT1 staining). Scale bars, 50 μm. g) In adult mice the bLTP is abolished in IP₃R2^{-/-} mice expressing the Ca²⁺ pump hPMCA2w/b in astrocytes (n = 14).

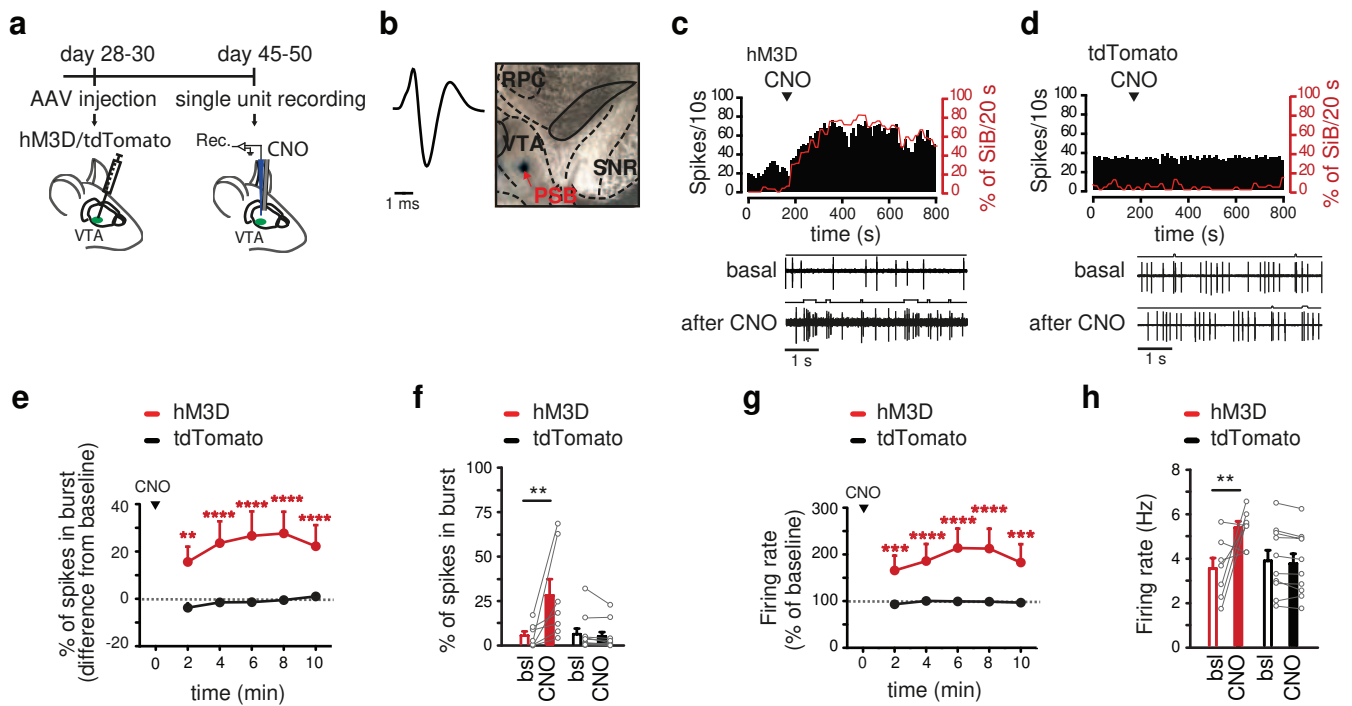


Figure 5. Chemogenetic activation of astrocytes *in vivo* favors bursting and overall firing activity in VTA DA neurons. a) Schematic of the experimental design showing the pipettes for recording (blue) and CNO application (gray). b) Left, representative action potential (start to end > 2.5 ms) from a putative VTA DA neuron. Right, location of a recorded putative VTA DA neuron. PSB; pontamine sky blue; SNR, substantia nigra pars reticulata; RPC, red nucleus. c, d) Upper panels, representative firing rate (Spikes/10 s) histograms and percentage of spikes in burst (SiB/20 s) trends over time of VTA DA neurons from mice expressing hM3D (c) or tdTomato (d) in VTA astrocytes. Lower panel, examples of raw spike traces of the same neurons before and after local CNO applications. e) Time course of the bursting activity (after local CNO applications (hM3D, n = 7; tdTomato n = 10). Two-way RM ANOVA and Bonferroni's multiple comparison test: main effects are not indicated; hM3D 2 minutes vs. hM3D basal **p < 0.01; hM3D 4, 6, 8, 10 minutes vs. hM3D basal **** p < 0.0001. f) Percentage of spikes in bursts before and after local CNO applications (n hM3D = 7; n tdTomato = 10). Two-way RM ANOVA and Bonferroni's multiple comparison test: main effects are not indicated; hM3D after CNO vs. hM3D basal ** p < 0.01. g) Time course of the firing rate after local CNO applications (hM3D, n = 7; tdTomato, n = 10). Two-way RM ANOVA and Bonferroni's multiple comparison test: main effects are not indicated; hM3D 2, 10 minutes vs. hM3D basal ***p < 0.001; hM3D 4, 6, 8 minutes vs. hM3D basal **** p < 0.0001. h) Firing rate before and after local CNO applications. Two-way RM ANOVA and Bonferroni's multiple comparison test: main effects are not indicated; hM3D after CNO vs. hM3D basal ** p < 0.01.

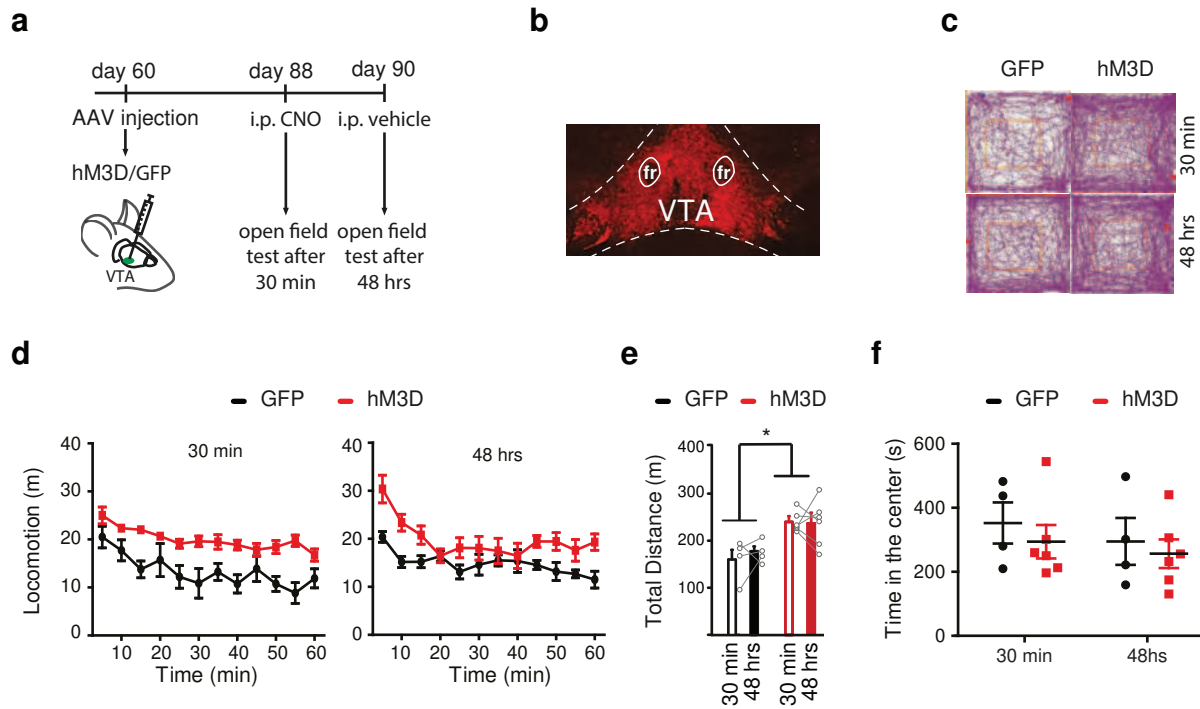


Figure 6. Chemogenetic activation of astrocytes *in vivo* induces a long-lasting motor hyperactivity. a) Schematic of the experimental design for testing locomotion in mice expressing in VTA astrocytes either GFP or hM3D, 30 min and 48 hrs after the intraperitoneal CNO application (3mg/kg). b) Fluorescence image of hM3D expression in VTA; fr, fasciculus retroflexus. c) Representative examples of locomotion in the open field test. d, e) Locomotor activity (d) and total distance (e) traveled by GFP- and hM3D-injected mice 30 min or 48 hrs after the intraperitoneal CNO injection (3 mg/kg, n = 4-6 mice/group). f) Time spent in the open field center of GFP- and hM3D-injected mice, at different time points after the intraperitoneal CNO injection.

Figures

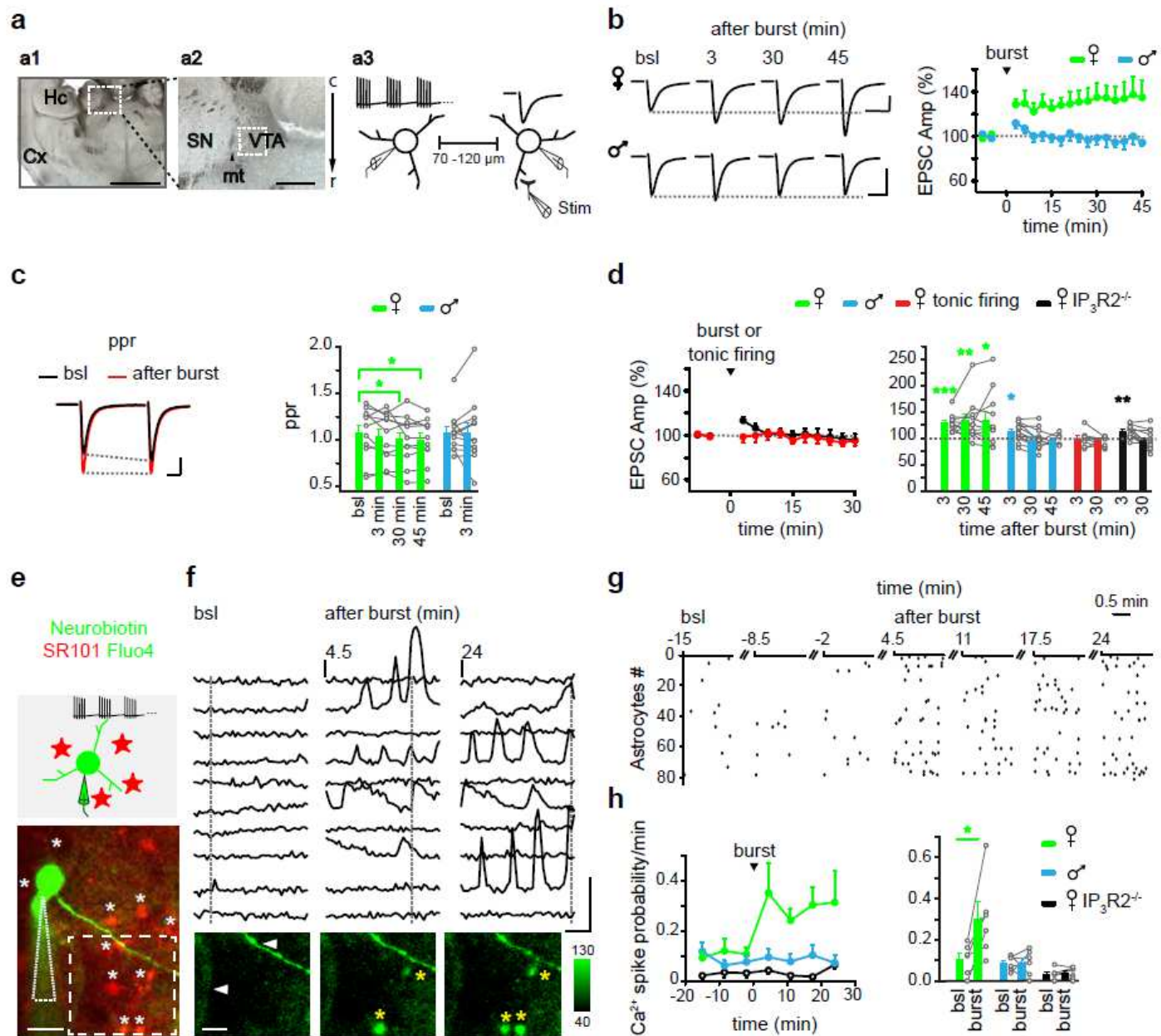


Figure 1

Astrocyte recruitment by DA neuron burst firing induces LTP of excitatory synapses onto adjacent DA neurons in young female mice. **a)** **a1**, low-magnification from a horizontal brain slice with a caudal-rostral orientation. Cx, cortex, Hc, hippocampus. Scale bar, 1 mm. **a2**, high magnification of the area indicated in **a1**. mt, medial terminal nucleus of the accessory optical tract; SN, substantia nigra (lateral to mt); VTA, ventral tegmental area (medial to mt), c, caudal; r, rostral. Dashed square, lateral part of the VTA where conventional DA neurons were recorded. Scale bar, 200 μ m. **a3**, schematic of the experimental design showing a DA neuron pair, two recording pipettes, the burst firing imposed to a DA neuron (left) and the EPSC evoked on the other DA neuron (right) by extracellular stimulation of rostral glutamatergic afferents

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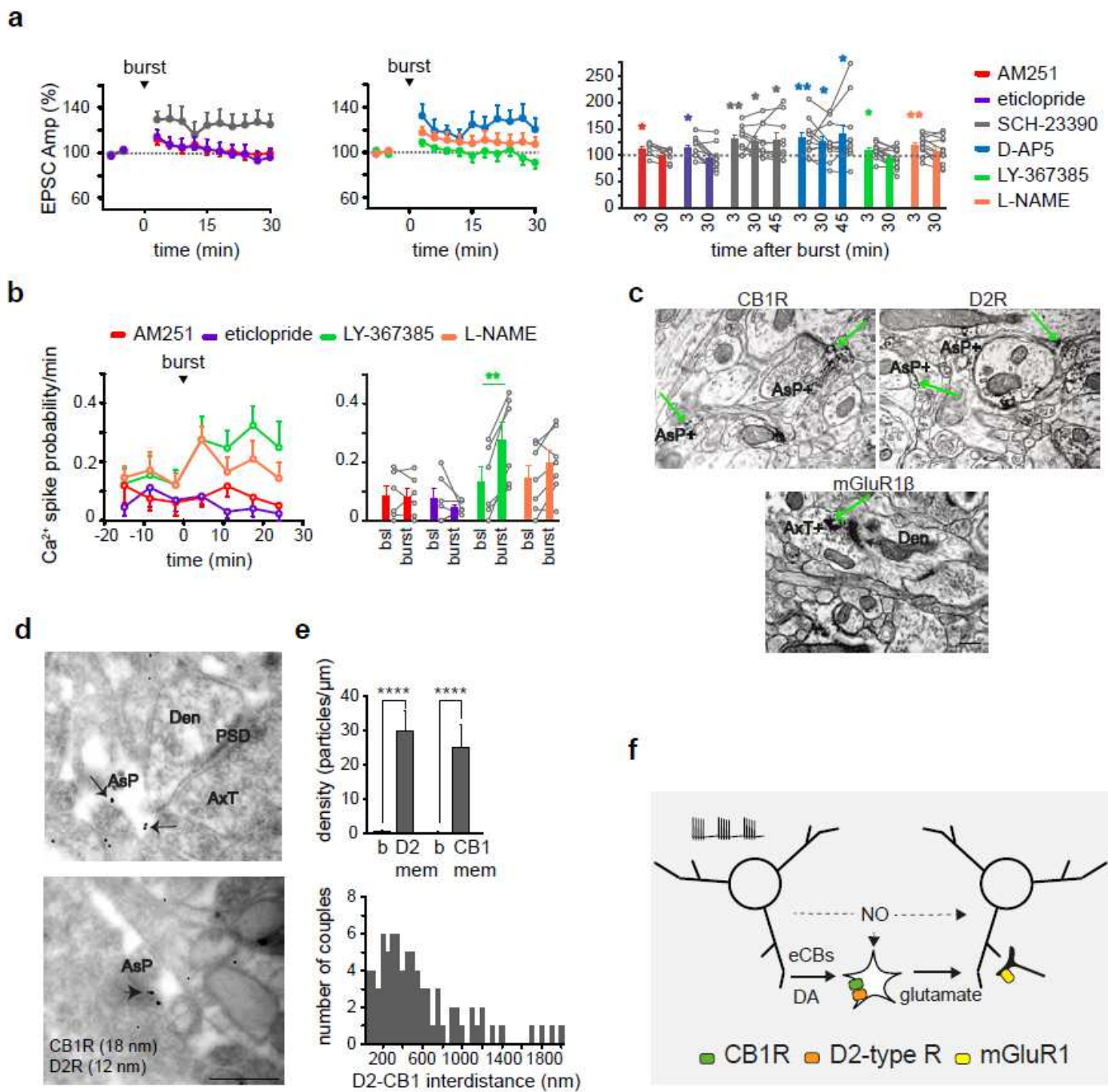


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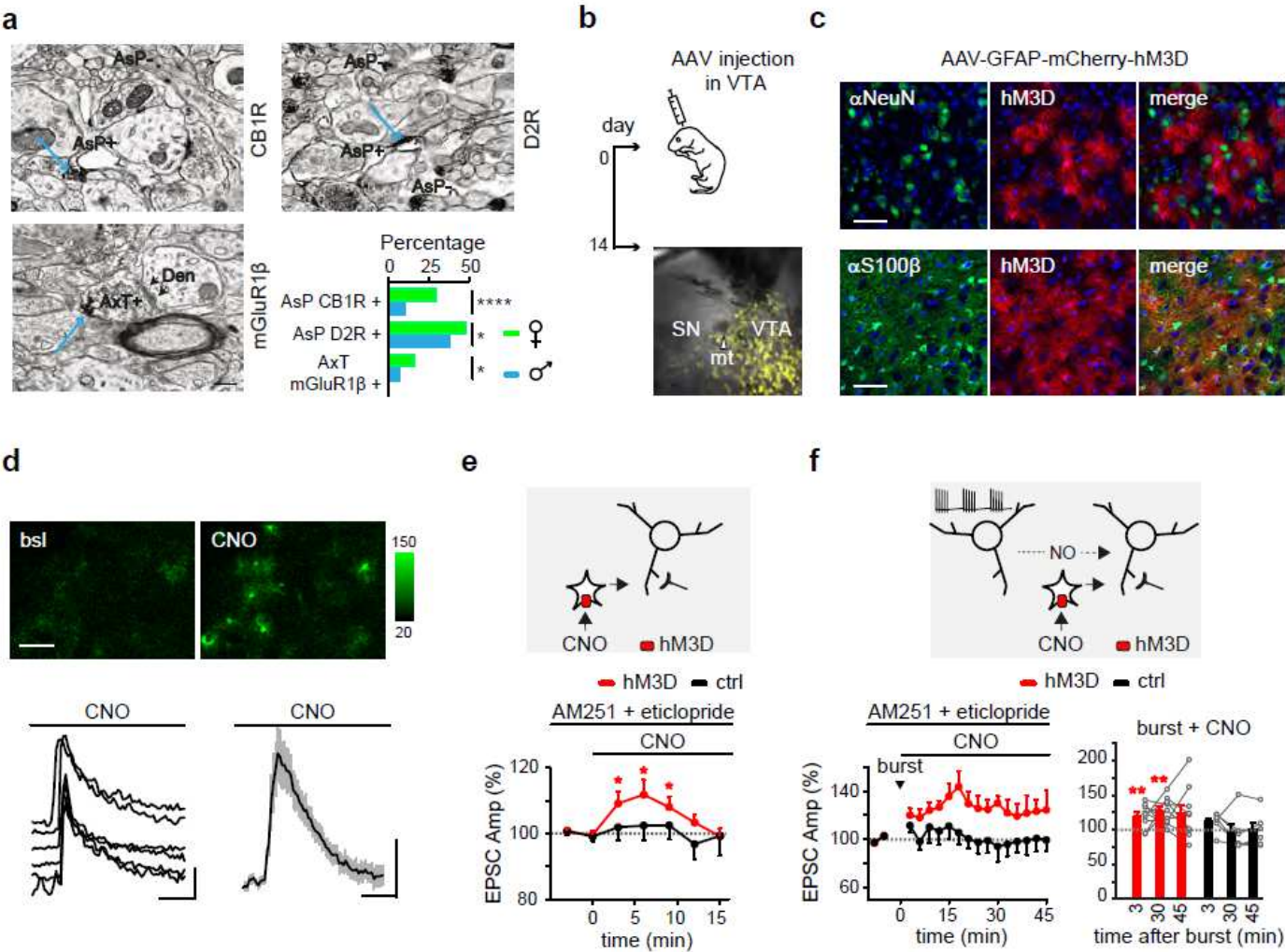


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Chemogenetic selective activation of astrocytes rescues bLTP in young male mice. a) Representative pre-embedding electron microscopy in the lateral VTA from a young male mouse of CB1 and D2R expression at AsP and mGluR1β expression at AxT forming asymmetric synaptic contact (arrowheads) with dendrite (Den). Blue arrows, immunopositive products in AsP (AsP+) and AxT. AsP-, AsP without immunoreactivity. Scale bar, 300 nm. Lower panel, quantification and comparison (contingency Fisher's

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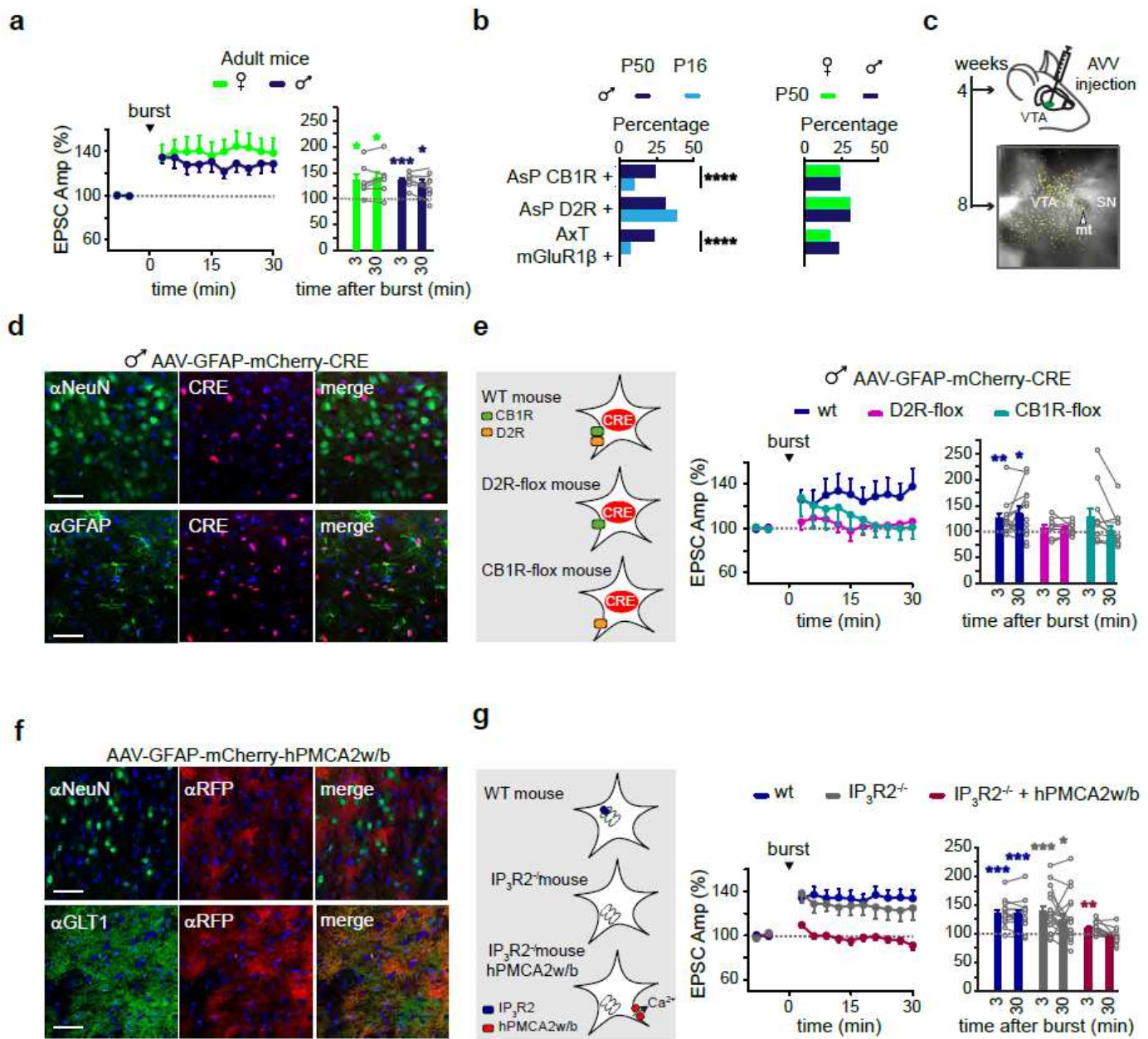


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Astrocytes induce bLTP in both female and male adult mice and bLTP is abolished in adult IP₃R2^{-/-} mice expressing a plasmatic Ca²⁺ pump in astrocytes. a) The bLTP is present in both female (n = 7) and male (n = 7) adult mice. b) Quantification and comparison (contingency Fisher's test) of CB1, D2 and mGluR1 β receptors expression in young and adult male mice and in adult female and male mice. c) Schematic of the AAV9- hGFAP-mCherry_iCre-WPRE-hGHp(A) injection in adult male mouse VTA and fluorescence image of a brain slice four weeks after viral injection (yellow, mCherry-Cre expression). d) Confocal images of the VTA from an adult male mouse injected with AAV9- hGFAP-mCherry_iCre-WPRE-hGHp(A), showing the fluorescence of mCherry-Cre (red), the nuclear Top-Ro3 (blue) and the specific green fluorescence for either neurons (α -NeuN staining) or astrocytes (α -GFAP staining). Scale bars, 50 μ m. e) When male mice express the Cre recombinase in VTA astrocytes, bLTP is preserved in wt mice (n = 13), but abolished in D2R- (n = 9) or CB1R-Floxed (n = 11) mice. f) Confocal images of the VTA from an adult

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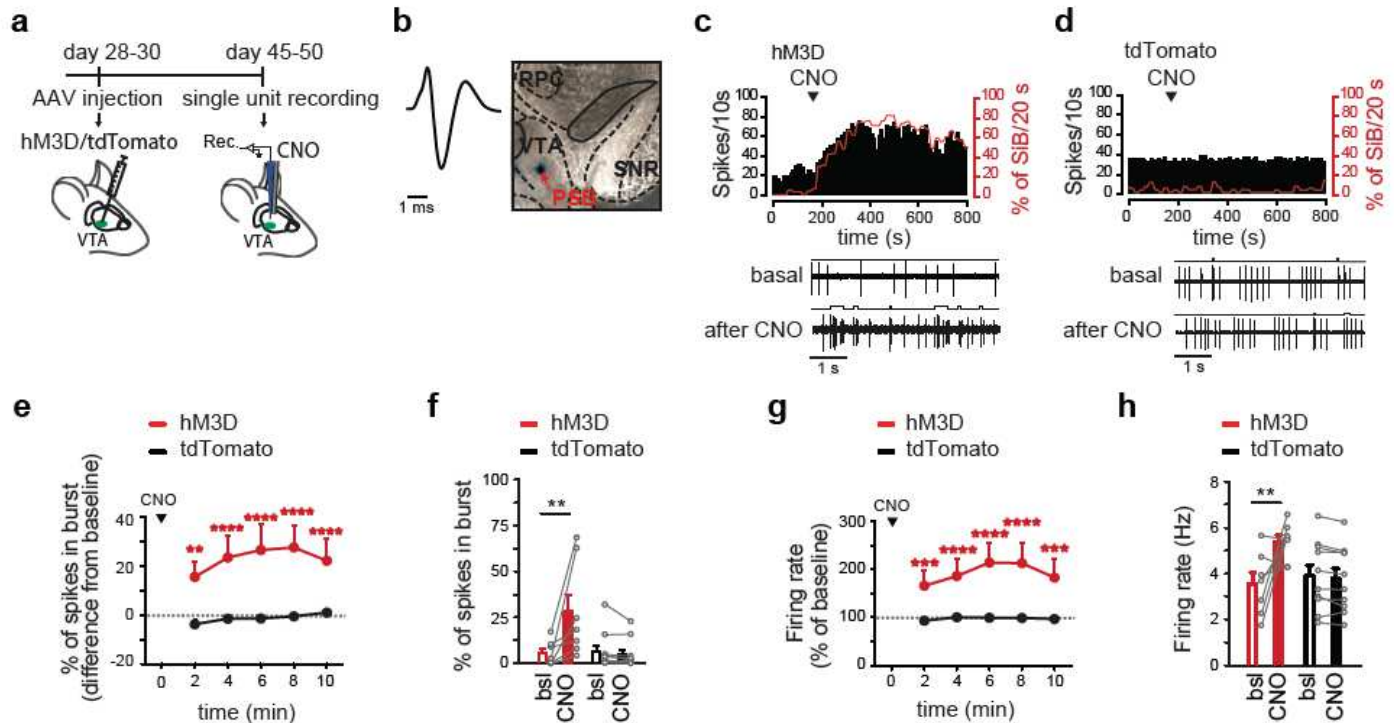


Figure 5

Chemogenetic activation of astrocytes in vivo favors bursting and overall firing activity in VTA DA neurons. a) Schematic of the experimental design showing the pipettes for recording (blue) and CNO application (gray). b) Left, representative action potential (start to end > 2.5 ms) from a putative VTA DA neuron. Right, location of a recorded putative VTA DA neuron. PSB; pontamine sky blue; SNR, substantia nigra pars reticulata; RPC, red nucleus. c, d) Upper panels, representative firing rate (Spikes/10 s) histograms and percentage of spikes in burst (SiB/20 s) trends over time of VTA DA neurons from mice expressing hM3D (c) or tdTomato (d) in VTA astrocytes. Lower panel, examples of raw spike traces of the same neurons before and after local CNO applications. e) Time course of the bursting activity (after local CNO applications) (hM3D, n = 7; tdTomato n = 10). Two-way RM ANOVA and Bonferroni's multiple comparison test: main effects are not indicated; hM3D 2 minutes vs. hM3D basal **p < 0.01; hM3D 4, 6, 8, 10 minutes vs. hM3D basal **** p < 0.0001. f) Percentage of spikes in bursts before and after local CNO applications (n hM3D = 7; n tdTomato = 10). Two-way RM ANOVA and Bonferroni's multiple comparison test: main effects are not indicated; hM3D after CNO vs. hM3D basal ** p < 0.01. g) Time course of the firing rate after local CNO applications (hM3D, n = 7; tdTomato, n = 10). Two-way RM ANOVA and Bonferroni's multiple comparison test: main effects are not indicated; hM3D 2, 10 minutes vs.

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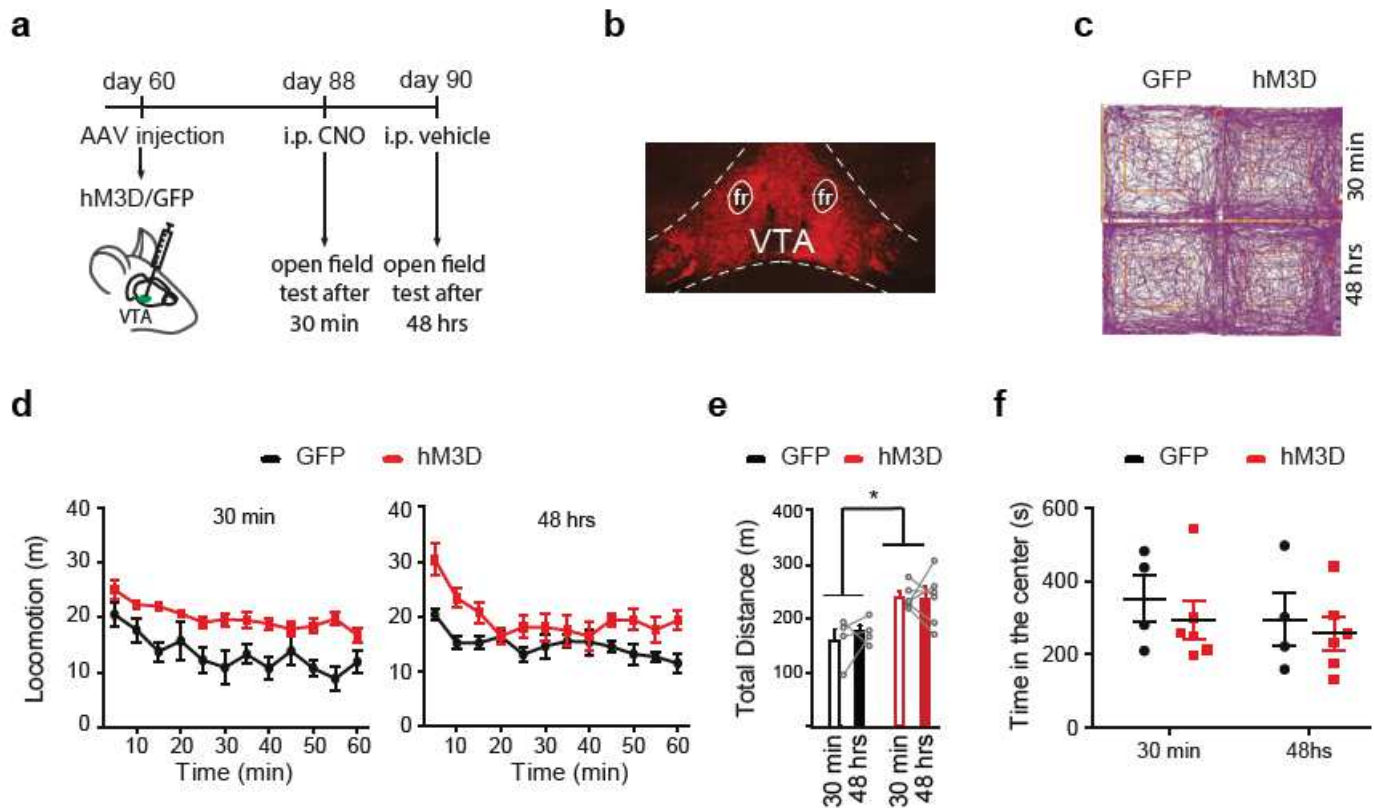


Figure 6

Chemogenetic activation of astrocytes in vivo induces a long-lasting motor hyperactivity. a) Schematic of the experimental design for testing locomotion in mice expressing in VTA astrocytes either GFP or hM3D, 30 min and 48 hrs after the intraperitoneal CNO application (3mg/kg). b) Fluorescence image of hM3D expression in VTA; fr, fasciculus retroflexus. c) Representative examples of locomotion in the open field test. d, e) Locomotor activity (d) and total distance (e) traveled by GFP- and hM3D-injected mice 30 min or 48 hrs after the intraperitoneal CNO injection (3 mg/kg, $n = 4-6$ mice/group). f) Time spent in the open field center of GFP- and hM3D-injected mice, at different time points after the intraperitoneal CNO injection.

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