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Clinical significance of BAP1 in Malignant Pleural Mesothelioma

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ABSTRACT

Introduction: Malignant pleural mesothelioma (MPM) is an aggressive disease, with few available treatment options. Identification of novel prognostic and predictive biomarkers is a priority. In MPM patients, BRCA-associated protein 1 (BAP1) alterations are detected in about 60% of cases and its prognostic role has been amply investigated in the last 10 years. However, the clinic relevance of BAP1 in MPM is still a matter of debate. In this study we aimed to clarify the prognostic role of BAP1 in MPM. Moreover, since miR-31 seems to be involved in BAP1 regulation at post-transcriptional level, we combined BAP1 status with the tissue expression levels of miR-31 in order to improve its prognostic performance.

Methods: A systematic literature search was conducted. The inclusion criteria were: immunohistochemical (nuclear positivity) investigation of BAP1 expression on tumor tissue; hazard ratio (HR) values for the overall survival (OS) obtained through multivariate analysis (or adjusted for histotype).

The expression of BAP1 and miR-31 was analyzed in tissues of 60 MPM patients treated with first-line chemotherapy. OS and progression-free Survival (PFS) were assessed by Kaplan-Meier method and Log-rank test was used to investigate differences among subgroups. Multivariate Cox regression analysis was used to evaluate independent predictors of survival.

Results: In our cohort, BAP1 was positive/retained (≥1%) in 23 samples (38%) and negative/loss in 37 samples (62%). BAP1 loss was significantly associated with epithelioid histotype (p=0.01). At univariate analysis, there were no significant difference in terms of OS between BAP1 retained group (median OS=18.1 months, 95% CI 11.2-

25.8) and BAP1 loss group (median OS=14.8 months, 95% CI: 10.7-29.3, p=0.17). Multivariate analysis showed that non-epithelioid histology was the only independent prognostic factor (HR 3.58, 95 % CI: 1.58–8.14, p=0.002). Even, from meta-analysis consisting of 698 patients, no differences were observed in term of OS according to BAP1 status (HR 1.11; 95% CI, 0.76-1.61; p=0.60).

Lower miR-31 levels were detected in epithelioid MPM (e-MPM) compared to the non-epithelioid subtypes, which was associated with BAP1 loss. By looking at the e-MPM subgroup, loss of BAP1 was not able to predict clinical outcome. Conversely, miR-31 levels were significantly associated with PFS (p=0.028), but not with OS (p=0.059). By combining the two biomarkers, e-MPM patients with BAP1 loss/low miR-31 levels showed a better prognosis compared to the ones with BAP1 retained/high miR-31 levels (median OS 22.6 months, 95% CI: 12.0-33.2 vs 17.0 months, 95% CI: 11.5-22.5, p=0.017 and median PFS 8.7 months, 95% CI: 3.3-14.1 vs 5.1 months, 95% CI: 2.5-7.6, p=0.020). The BAP1 and miR-31 combination was confirmed at multivariate analysis as an independent prognostic factor for e-MPM patients.

Conclusion: BAP1 alone was unable to stratify MPM patients based on its status when histotype was considered. However, in e-MPM patients, prognostic stratification may be improved by simultaneously assessing BAP1 status and miR-31 levels. The two-biomarker score is useful to identify a subgroup of e-MPM tumors characterized by BAP1 retained and high miR-31 levels with worse clinical outcome.

ABBREVIATIONS

AP-1 activator protein 1

AST atypical Spitz tumor

ASXL putative polycomb group protein

BAP1 BRCA-associated protein 1

BARD1 BRCA1-associated RING domain protein 1

Bcl-2 B-cell lymphoma 2

BRCA2 breast cancer type 2 susceptibility protein

CD cluster of differentiation

CDKN2 cyclin-dependent kinase inhibitor 2A

CM cutaneous melanoma

COX7C cytochrome c oxidase subunit 7C

CT computed tomography

CTD C-terminal domain

CTLA-4 cytotoxic T-lymphocyte—associated protein 4

DAMP damage associated molecular pattern

DUB deubiquitinating enzyme

EPD extended pleurectomy/decortication

EPP extrapleural pneumonectomy

ER endoplasmic reticulum

FFPE Formalin-Fixed Paraffin-Embedded

FISH fluorescence *in situ* hybridization

FoxK Forkhead Box K

HBM host cell factor 1 binding domain

HCF1 host cell factor 1

HMGB1 high mobility group box 1

HGF hepatocyte growth factor

HR hazard ratio

IGF insulin-like growth factor

IHC immunohistochemistry

IL interleukin

IP3R3 inositol 1,4,5-trisphosphate receptor type 3

LATS2 large tumor suppressor kinase 2

LDH lactate dehydrogenase

MAM mitochondrial-associated membranes

MAPK mitogen activated protein kinases

MBAIT melanocytic BAP1-mutated Atypical Intradermal Tumor

MCP-1 monocyte chemoattractant protein-1

MCU mitochondrial uniporter channel

miRNA microRNA

MLH1 mutL homolog 1

MLH3 mutL homolog 3

MM malignant mesothelioma

MPM malignant pleural mesothelioma

MRI magnetic resonance imaging

mTOR mammalian target of rapamycin

NF2 neurofibromin 2

NF-kB nuclear factor kappa-light-chain-enhancer of activated B cells

NGS next-generation sequencing

NLS nuclear localization signal

OGT O-linked N-acetylglucosamine transferase

OS overall survival

PDGF platelet-derived growth factor

PD-L1 programmed death-ligand 1

PET positron emission tomography

PFS progression-free survival

PMT pemetrexed maintenance therapy

PRC polycomb-repressive complexes

PR-DUB polycomb group repressive deubiquitinase complex

qRT-PCR real-time PCR

RAGE receptor for advanced glycation endproducts

RCC renal cell carcinoma

RNS reactive nitrogen species

ROS reactive oxygen species

SETD2 SET domain containing 2

SMRP soluble mesothelin-related peptides

TGF- β transforming growth factor β

TNF- α tumor necrosis factor α

TP53 tumor protein 53

UCH ubiquitin carboxyl hydrolase domain

ULK Unc-51 like autophagy activating kinase

UM uveal melanoma

VDAC voltage-dependent anion channel

VEGF vascular endothelial growth factor

WT1 wilms' tumor protein

YY1 Ying Yang 1

1. INTRODUCTION

1.1 Malignant Pleural Mesothelioma

Malignant pleural mesothelioma (MPM) is an aggressive treatment resistant tumor that arises from the neoplastic transformation of the pleural mesothelium, a thin membrane that covers and protects the lung (1). MPM represents up to 80% of all cases of malignant mesothelioma (MM). The other form of mesothelioma originates from other serous membranes coated with mesothelium and are peritoneal mesotheliuma (other 20% of mesothelioma), pericardial mesothelioma and mesothelioma of vaginal tunic that are both very rare (2). Seventy per cent of MPM cases are associated with documented exposure to asbestos and develop after a latency period of 30-40 years (4). MPM develops insidiously in patients and is difficult to be diagnosed in the early stages because it does not show specific symptoms until advanced stages. Even diagnostic tools are not helpful for early detections of MPM and there is a lack of serum biomarkers that have not yet been determined. The body cavities where MPM initially develops, present anatomical location and characteristics that also causes malignant cells to easily spread and invade the adjacent cavities. Due to late diagnosis, only a few patients undergo surgery and current therapy is not very effective so life expectancy ranges from 9 to 18 months from diagnosis (5).

1.1.1 Epidemiology

Until the second half of the 20th century MPM was extremely rare, but its incidence and mortality rates began to rise in the 1960s after the massive use of asbestos, a mineral fiber,

during World War II and thereafter (6). The first studies that confirmed the link between asbestos and MPM were two epidemiological studies conducted by Doll et al and Wagner et al in 1955 and 1960, respectively (7, 8).

There are approximately 400 different types of asbestos fibers present in nature, but those used commercially are six (amphiboles fibers [crocidolite, actinolite, tremolite, anthophyllite, and amosite] and serpentine fibers [chrysotile]) and were collectively called "asbestos" (**Figure 1**) (9).

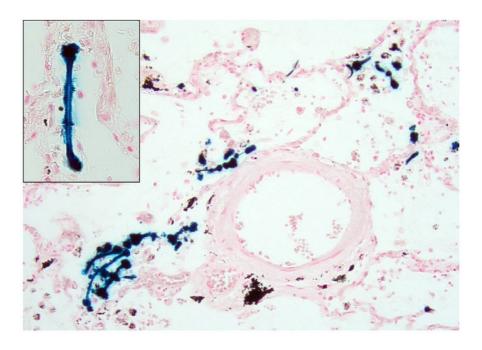


Figure 1. Presence of asbestos fiber in a lung alveoli's biopsy from a patient with mesothelioma who worked for "Eternit,"

The use of these six fibers were regulated in high-resource countries between 1970 and late 1980, but not the remaining approximately 400 mineral fibers, although many of them are carcinogenic and have been associated with mesothelioma (9, 10).

Asbestos is characterized by high resistance to heat, as well as chemical and biological agents, abrasion, and wear; therefore, it was widely used in the shipbuilding and construction industries, especially between the 1940s and 1979 in the United States and

Europe (11). MPM is a disease characterized by a long latency interval between the onset of exposure and the appearance of the disease (from 20 to 40 years and over) (12). Thus, despite controls on the use of asbestos introduced over the years, the incidence and the annual number of deaths from mesothelioma has continued to rise worldwide with a deaths peak expected between 2015-2025 and, according to some experts, even in 2040 (13–18).

According to WHO estimates, more than 100,000 deaths are due to the consequences of occupational exposure to asbestos (mesothelioma, lung cancer, and asbestosis) and it has been estimated that, between 1994 and 2008, age-adjusted mesothelioma mortality rates increased by 5.37% per year worldwide (19). In Western Europe, due to asbestos exposure, are expected 500,000 deaths in the first thirty years of 2000 (20-22) and the underlying level of the mortality rate from mesothelioma is about 1–2 per million/year (23-25). Currently there are about 2,000 to 3,000 cases per year in the United States, but the incidence rate varies between states with or without asbestos industry (from less than one case to 2-3 cases per 100.000 inhabitants). In the U.S. there are 3000 deaths per year and projections estimates 100.000 deaths over the next 40 years (9,10,19, 26-28). According to the Surveillance, Epidemiology, and End Results (SEER) database, the MPM in the USA reached the peak in the 1980s to 1990s with an incidence in men of 2.5 cases per 100.000 and now is stable to 1.8 cases per 100.000. In women the rate has been 0.4 cases per 100.000 and has not changed over time. Women have also threefold better survival than men (29). In the US the mean age of death for MPM was 72.8 years, with a male-to-female (M:F) mortality ratio of 4.2:1, since men were traditionally more likely to be employed in works involving asbestos exposure. But, with equivalent asbestos exposure, men and women have a similar incidence of MPM (30).

The highest age-standardized incidence rates in 2018 were observed in the US, Western Europe, Australia, Russia, Turkey, South Africa, and Argentina (31). The incidence varies between 7 per million in Japan and 40 per million in Australia (32, 33). In the last twenty years, the incidence has steadily increased in Europe in the industrialized countries, with an average incidence of 20 per million inhabitants/year and its peak is expected to be around 2020–2025 (15, 34). In addition, a temporal decline in mesothelioma mortality rates in males has been observed in Sweden and in the United Kingdom, probably thanks to the introduction of regulatory laws during 1970s (35). Instead, rates in those countries among women are still rising as of 2018 (19, 36, 37). Europe incidence data for the year 2000-2007 are available on RARECARE, which estimates the frequency of rare cancers in Europe. (38) (Table 1).

Tumor	Crude Incidence Rate Per 100.000		onfidence erval	Number of Cases Collected in the RARECAREnet Database from 2000- 2007	Estimated New Cases EU 2013
Malignant Mesothelioma	2.14	2.12	2.16	33.552	12.526
Mesothelioma of pleura and pericardium	1.83	1.80	1.85	28.627	10.703
Mesothelioma of peritoneum and tunica vaginalis	0.13	0.12	0.13	2.065	746

Table 1. RARECARE incidence of malignant mesothelioma (MM) 2000-2007. Source: http://www.rarecarenet.eu/analysys.php, Updated 10 June 2019.

Russia, China, Thailand, Brazil, India, Kazakhstan, Iran, and Ukraine were the highest worldwide consumption of asbestos from 1995 to 2003, but the World Health Organization include mesothelioma incidence and mortality data only for Kazakhstan,

where incidence rates increased from approximately 0 to 0.26 cases per 100,000 persons in the past 10 years (31).

Unfortunately, each country banned and regulated the use of asbestos in different times, so, in the next decades, it is expected that mesothelioma rates will follow dissimilar patterns. By 1990, in the most industrialized countries, the use of asbestos had been reduced by at least 75% from the peak asbestos consumption. In 1999, Iran, Korea, Chile, and Egypt reached the same level of reduction, as did Nigeria, Zimbabwe, the United Arab Emirates, Ukraine, and Kazakhstan between 2000 and 2005. But, where the asbestos is still used, such as the Russian Federation, India, and China, an increase in age-adjusted mesothelioma incidence and mortality rates is expected in the coming years (39). In the European Union, the directive 2003/18/CE of the European Parliament and of the Council of 27 March 2003 provides the obligation to the completely stop of the asbestos use by 15 April 2006. In Italy, the law n. 257 of 27 March 1992 declared the "cessation of the use of asbestos", and the ban on the extraction, import, export, marketing, and production of asbestos products and products containing asbestos, but the law did not prohibit the indirect use. Therefore, the Italian territory is disseminated of several million tons of compact materials containing asbestos and many tons of friable asbestos are still present in many contaminated sites, industrial and nonindustrial, public, and private. In Italy, the number of exposed worker is very high because the raw asbestos produced or imported has been used in a wide range of industrial activities such as sectors of industrial production of asbestos-cement manufactured articles, textile manufactures containing asbestos, shipbuilding, repair and demolition of railway rolling stock, construction, and in many other sectors of economic activity (2).

Moreover, given the presence of numerous asbestos mines, in Italy the production of raw asbestos was very high, with 3,748,550 tons of raw asbestos produced up to 1992, and with a peak of more than 160,000 tons/year between 1976 and 1980. For these reasons, Italy is one of the most involved and sensitive country in asbestos related diseases monitoring and control (2). In particular, Piedmont was one of the most affected Italian regions, since it hosted the main European chrysotile quarry (Balangero, operating from 1917 through 1990) and the largest facility for the manufacture of asbestos-cement products (Casale Monferrato, operating from 1907 through 1985) (40-43). The asbestos cement factory, owned by Eternit, in Casale Monferrato, produced plain and corrugated sheets, tubes, and high-pressure pipes. The asbestos cement factory had an average workforce of over 1000 workers and was located close to the town center of Casale Monferrato, causing also airborne asbestos contamination in the town. Therefore, Casale Monferrato is one of the Italian towns with the highest incidence and mortality of malignant mesothelioma [44, 45]. The Registry of Malignant Mesothelioma – ReNaM – of Piedmont reported that, between 2010-2014, the incidence rates in Casale Monferrato was of 90.2/100.000 person years in men and 45.4 in women [Registry of Malignant Mesothelioma – ReNaM – of Piedmont, www.cpo.it/it/articles/show/incidenza-esopravvivenza-dei-mesoteliomi-1990-2014/]. The case of Casale Monferrato is even an example that mesothelioma does not occur only in people who had work in direct contact with asbestos, but even to people that had an environmental exposure to asbestos. Between 2001-2006, an analysis of exposure histories of 847 cases of mesothelioma demonstrated that only 475 cases (56%) was considered occupationally exposed, whereas, 357 (42%) was classified as non-occupationally exposed, mostly due both to living in proximity to the asbestos-cement factory ("environmental" exposures: 200

cases) or to have a factory workers in family ('familial' exposure: 144 cases). These evidences make known the how persisting is asbestos contamination in the workplace and in the general environment from asbestos-cement production (46). Unlike MPM caused by occupational exposure, those caused by the environment tend to occur at a younger age (<55 years) with an M:F ratio close to 1:1.6, probably why environmental exposure often begins at birth and occurs randomly among sexes (19).

In Italy, the epidemiological surveillance of mesothelioma cases is assigned by the Decree of the President of the Council of Ministers n. 308/2002 to the ReNaM established at the National Institute for Insurance against Accidents at Work (INAIL) [47]. For the incidence period between 1993 and 2015, ReNaM has collected 27,356 MM and the modalities of exposure to asbestos have been investigated for 21,387 (78%) cases (**Table 2**).

Madalita of ann arms	Incidence period (1993-2015)				
Modality of exposure -	Male (%)	Female (%)	Total (%)		
Occupational, define	9.300 (59.3)	987 (17.3)	10.287 (48.1)		
Occupational, probable	1.358 (8.7)	191 (3.3)	1.549 (7.2)		
Occupational, possible	2.246 (14.3)	736 (12.9)	2.982 (13.9)		
Familial	152 (1.0)	895 (15.7)	1.047 (4.9)		
Enviromental	409 (2.6)	530 (9.3)	939 (4.4)		
Other non-occupational	128 (0.8)	194 (3.4)	322 (1.5)		
Unlikely	268 (1.7)	308 (5.4)	576 (2.7)		
Uknown	1.824 (11.6)	1.861 (32.6)	3.685 (17.2)		
Total defined	15.685 (100.0)	5.702 (100.0)	21.387 (100.0)		
Total	19.633 (100.0)	7.723 (100.0)	27.356 (100.0)		
Total defined	15.685 (79.9)	5.702 (73.8)	21.387 (78.2)		
Total undefined	3.948 (20.1)	2.021 (26.2)	5.969 (21.8)		

Table 2 Italian National Mesothelioma Register (ReNaM) archives. Collected malignant mesothelioma cases by modality of asbestos exposure and gender. ReNaM archives updated at December 2016, diagnosis period 1993–2015*, Italy

Among them, an occupational exposure has been defined for 14,818 (around 70%) of defined cases. Non-occupational exposure has been defined for 9.3% of cases of which 4.9% due to family exposure and 5% due to environmental exposure. The exposure median period was 1959 [1951-1966]. Average age at diagnosis of malignant mesothelioma (MM) was 70 years, with no appreciable gender differences (70.8 years in woman and 69.5 in men). MM cases younger than 45 years at diagnosis are very rare (less than 2%). More than 90% of collected cases are pleural mesotheliomas (93%). Peritoneal MM cases are 6.5% (5.3% and 9.4% in men and women respectively) and pericardial and tunica vaginalis testis MM cases are very rare (58 and 79 collected cases respectively among the entire ReNaM archives). Epithelioid histology represented the 55% of cases. Gender ratio is, constantly in time, equal to 2.54 (M/F) and to 2.64, if restricted to pleural cases (48). From this analysis it was possible an examination of the geographic distribution of mesothelioma cases in Italy which enabled the identification of clusters in the municipalities with the highest incidence rates, such as Casale Monferrato, Broni, Genoa, La Spezia, Grugliasco-Collegno, Monfalcone, Trieste, Castellamare di Stabia, Bari, Taranto, Biancavilla, and Augusta (49-52) (**Figure 2**).

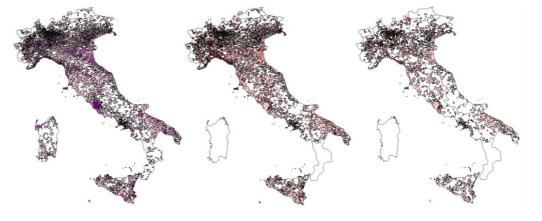


Figure 2. Italian National Mesothelioma Register (ReNaM) archives. Number of MPM cases in men and women (a) and incidence rates of MPM in men (b) and woman (c), by municipality of residence at the time of diagnosis, Italy. ReNaM archives updated to December 2016, diagnosis period 1993–2015. Sardinia, Molise and Calabria are excluded due to lack of data.

1.1.2 Etiopathogenesis

MPM has been associated with asbestos exposure and over 80% of MPM patients have a history of asbestos exposure (53). Although asbestos, in particular crocidolite, is considered to be the carcinogenic agent mostly involved in MPM etiology, only the 5% of asbestos-exposed subjects develop the disease (54). As MPM is a relatively rare malignancy, it is still not clear how much exposure is needed to cause MPM, and what mechanisms are triggered during the long latency from the time of exposure to the time of tumor development. In vivo experiments have shown that there is a clear dose-response, thus, MPM does not develop below a certain dose of exposure (about 1 mg) (55). However, in human, there is no defined threshold limit for mesothelioma risk, which may arise even after short asbestos exposure (24, 56, 57).

When long and thin asbestos fibers are inhaled deeply into the lung and penetrate the pleural space, the interaction between asbestos fibers, mesothelial cells and inflammatory cells occur, causing chronic inflammation and atypical mesothelial hyperplasia. The single flat layer of mesothelial cells that form the pleura and peritoneum round up and forming multicellular layers from which over time mesothelioma may arise (58). Chronic inflammation is the primary mechanism of asbestos-related carcinogenesis. The continuous generation of highly reactive oxygen species (ROS) promotes DNA mutation, and trigger transformation (59). The presence of iron (II) ions (Fe2+) in asbestos fibers can also induce hemolysis by sequestering Fe(II) from hemoglobin (60); this lead to a particular reaction (Fenton reaction), where free Fe(II) disproportionate H₂O₂ into hydroxyl radicals that oxidize DNA, free nucleic acids, proteins, and lipids (61). Also, Macrophage are unable to complete digest asbestos fiber after phagocytosis, and, over time, this generates abundant ROS and even reactive nitrogen species (RNS). Asbestos

fibers are also engulfed by mesothelial cells where can physically interfere with the mitotic process of the cell cycle by disrupting mitotic spindles. This may result in chromosomal structural abnormalities and aneuploidy of mesothelial cells. Asbestos fibers absorb a variety of proteins and chemicals on their broad surface and may result in the accumulation of hazardous molecules including carcinogens. The fibers can also bind important cellular proteins necessary for the normal function of mesothelial cells (62, 63). All these processes lead to DNA damage in the forms of single-strands breaks, crosslinks, and double strand-breaks. Particularly, the oxidation of the 8th carbon on the DNA base guanine (8-oxo-2'deoxyguanosine) changes normal 2'deoxyguanosine Watson-Crick base pairing preference from 2'deoxycitosine to 2'deoxyadenosine, resulting in G to T and C to A transversions (64).

Paradoxically, human mesothelial cells are very susceptible to this asbestos cytotoxicity, which raises an issue of how asbestos causes MPM if human mesothelial cells exposed to asbestos die (65). This paradox was addressed by recent findings which demonstrated a critical role for TNF- α and NF-kB signaling in mediating human mesothelial cell responses to asbestos. TNF- α signaling through NF-kB-dependent mechanisms increases the percent of human mesothelial cells that survive to asbestos exposure, and which can therefore undergo to malignant transformation. These findings suggest a pathogenic model where asbestos causes expression of several cytokines, including MCP-1, which in turn cause the accumulation of macrophages in the pleura and lung where they can encounter asbestos and release TNF- α . At the same time, in mesothelial cells, asbestos directly induces the expression of the TNF- α receptor and the secretion of TNF- α . When TNF- α , released by macrophages and by human mesothelial cells, binds its own receptor this activates NF-kB signaling which promote survival of human mesothelial cells

following asbestos exposure (66). This process allows human mesothelial cells with accumulated asbestos-induced DNA damage to survive, divide and propagate genetic aberrations in pre-malignant cells. Finally, if sufficient genetic damage accumulates occur; these cells could eventually develop into a MPM (**Figure 3**).

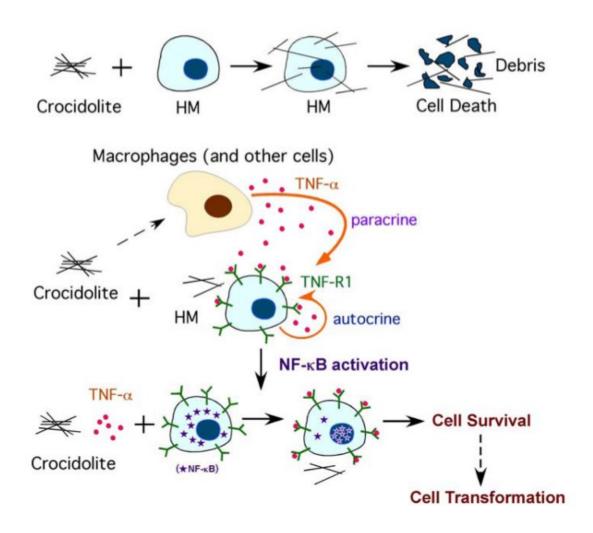


Figure 3. Mechanisms of asbestos-induced carcinogenesis

Even autophagy has a role in asbestos-induced carcinogenesis via the pro inflammatory protein high mobility group box 1 (HMGB1). There are two type of autophagy: constitutive (background) autophagy, that recycles cellular components from aged/damaged organelles and induced (reactive) autophagy, that occurs in response to environmental challenges and protect cells from apoptosis and necrosis (66-69). Yang et

al demonstrated that asbestos induced programmed cell necrosis in human mesothelial cells and induced necrotic cell to release HMGB1 into extracellular space (70). In addition to stimulate inflammation (71-73), HMGB1 triggers the inflammasome pathway (74) and sustain the chronic inflammatory process induced by asbestos; HMGB1 modulates even autophagy in cells under pro-autophagic stressed (75-77). Finally, in a recent study, Xue et al demonstrated the link between autophagy and asbestos-induced HMGB1 translocation from the nucleus to the cytoplasm and to the extracellular space. They found that both cytoplasmic and extracellular HMGB1 mediated asbestos-induced autophagy through the RAGE-mTOR-ULK and Beclin 1 pathways (78). Cytoplasmic HMGB1 can activate p-Beclin 1 displacing Bcl-2 (77) and extracellular HMGB1, thus contributing to p-Beclin 1 activation and autophagy via binding to the cell-surface receptor RAGE, which, in turn, initiated a downstream pathways that culminated in increased levels of p-Beclin 1 (Figure 4) (78). This mechanism allows asbestos-exposed mesothelial cells that have sustained DNA damaged to survive. Otherwise, in human mesothelial cells with HMGB1 silenced, autophagy is inhibited and cells undergoing to apoptosis or necrosis (78).

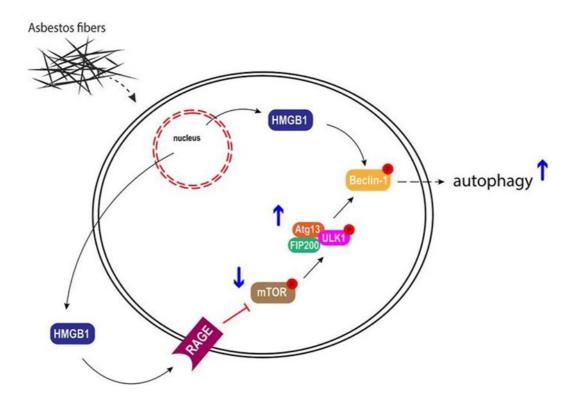


Figure 4. Schematic representations of the asbestos-HMGB1-autophagy pathway.

In addition to TNF- α , other growth factors and cytokines are involved in the carcinogenesis process caused by asbestos: transforming growth factor β (TGF- β), which may play a role in stimulating the growth of cancer cells; platelet-derived growth factor (PDGF), an important mitotic agent, which may act as a regulatory factor in the proliferation of MPM cells; the insulin-like growth factor (IGF), which it would promote tumor proliferation and cell migration (79). Furthermore, interleukins, such as IL-6 and IL-8, could promote tumor cell growth and neo-angiogenesis with the development of newly formed capillaries. In particular, IL-8 plays the role of an autocrine factor for cell growth (80). Other growth factors involved in the pathogenetic mechanism of mesothelioma are the vascular endothelial growth factor (VEGF), that promotes neo-angiogenesis (81) and the hepatocyte growth factor (HGF) which stimulates cell proliferation, as well migration and invasiveness in tissues by the tumor (82).

In addition to the production of growth factors, asbestos stimulates and interacts with several signaling pathways, such as the mitogen activated protein kinases (MAPK) signaling that lead to the activation of the transcription factor AP-1 that stimulates the mitosis of mesothelial cells (62).

Because of the carcinogenic "field effect" caused by asbestos, mesotheliomas are often polyclonal (83, 84). Recently, the Cancer Genome Atlas program, published a study aimed to investigate the genetic alterations of 74 mesotheliomas using next-generation sequencing (NGS). As in the previous comprehensive NGS study by Bueno et al (85), Hmeljak et al (86) reported frequent somatic mutations and/or copy-number alterations of CDKN2A, NF2, TP53, LATS2, and SETD2 (**Figure 5**) (85, 86). In addition, the authors (85) reported a 57% prevalence of BAP1 mutations, confirming the previous comprehensive analysis reporting a 60% prevalence of BAP1 mutations (87).

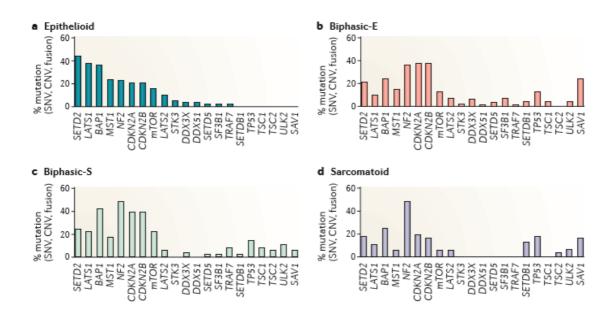


Figure 5. Mutational hierarchy in mesothelioma subtypes.

As previously reported only 5% of asbestos-exposed subjects developed MPM. It has been seen that, in addition to the mechanisms described above, gene predisposition is also involved in the development of mesothelioma (88). Inherited mutations of DNA repair genes and of other genes could accelerate the accumulation of DNA damage and/or the percentage of cells carrying DNA damage (89). In addition, inherited mutations may also increase susceptibility to environmental carcinogens (GxE interaction) (89). Several tumor suppressor genes have recently been found to cause a hereditary predisposition to mesothelioma and, overall, at least 12% of mesotheliomas occur in carriers of genetic mutations (90-92). Most of these heterozygous germline mutations occur in genes that regulate DNA repair, such as *MLH1*, *MLH3*, *TP53*, BRCA2, but the first and the most studied gene associated with mesothelioma predispositions is BAP1 (90-92).

1.1.3 Clinical presentation and diagnosis

MPM symptoms are non-specific and may mimic other respiratory diseases (93,94). Most patients with MPM present dyspnea which is associated with breathlessness, chest pain, weight loss and fatigue (95, 96) and is predominantly related to the development of a pleural effusion. Thoracic pain is common and multifactorial in MPM and is due by tumor invasion of chest wall. When tumor invades neural intercostal, paravertebral or brachial plexus structures may cause bone pain and neuropathic pain. At advanced stages, MPM symptoms include weight loss, fatigue, fevers, cachexia and nights sweats and are often detected hypoalbuminemia, thrombocytosis, elevated erythrocyte sedimentation rate and anaemia (32, 97)

MPM develops initially unilaterally and local invasion of neighboring structures, including lymph node involvement, can occur, causing superior vena cava syndrome,

pericardial effusion and subsequent cardiac tamponade, spinal cord compression as well as a subcutaneous involvement. The affected site becomes fixed and cannot expand. Other MPM progression may involve invasion of contralateral pleural cavity and peritoneum. Unlike lung cancer, distant metastases rarely occur in MPM since patients die before metastases occur (98, 99).

Standard diagnostic work-up in patients with MPM starts with Chest X-ray and/or computed tomography (CT) scan of chest and upper abdomen in order to shows pleural effusion at disease site, pleural thickening and involvement of the interlobar fissure and invasion of the chest wall (**Figure 6**).

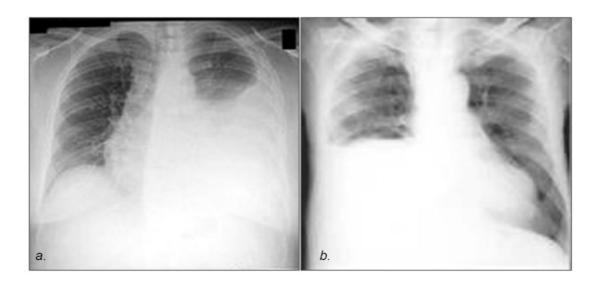


Figure 6. Chest radiographs in P-A projection of the chest with left (a) and right (b) pleural effusion, respectively.

Then, by thoracentesis or by fine-needle aspiration biopsy the pleural effusion is drained, and the fluid is examined cytologically. Pleural biopsy is often required for a definitive diagnosis histological and bio-molecular diagnosis, and pleurodesis with talc poudrage is often performed during the same surgical setting. Recognition and rapid investigations of the pleural or peritoneal effusion are key for early diagnosis. Delayed diagnosis will

inevitably lead to tumor progression, limiting the therapeutic options. Also, by PET and MRI, it is possible define the extension of the disease. Additional investigations include blood markers and pulmonary function tests (100, 101).

1.1.4 Histology and staging

As previously said, a definitive diagnosis of mesothelioma requires histological evaluation. Biopsies are mostly obtained by thoracoscopy or laparoscopy.

According to the 2015 World Health Organization Classification of Tumors of the Pleura, malignant mesothelioma is broadly classified as epithelioid, sarcomatoid, or biphasic types (102) In the NCDB (*National Cancer Database, USA*) from 2003 to 2014, MPM subtypes were 38.4% epithelioid, 12.3% sarcomatoid, 11% biphasic, and 44.7% not otherwise specified (103)

Epithelioid MPM (**Figure 7a**) is the most common histotype (60%–70%) of MPM and is associated with less severe prognosis than non-epithelioid MPM. (104, 105). The median overall survival for the epithelioid mesothelioma is 24.9 month (106). Histologically, the tumor cells have an epithelioid morphology with an ample cytoplasm with well-defined cell borders. The nuclei are frequently bland and can look like reactive mesothelial cells with moderate cytologic atypia. Growth patterns include tubulo-papillary, microglandular (adenomatoid), acinar, and solid. Morphologic variants of epithelioid mesothelioma include clear cell, small cell, deciduoid, adenoid cystic, lymphangiomatoid, and signet ring cell morphology (106).

Sarcomatoid MPM (**Figure 7b**) is the most aggressive subtype with a median survival of 7 months (106) It is characterized by a proliferation of spindle cells infiltrating dense fibrous stroma and exhibit a disorganized growth pattern. The spindle cells may show

marked nuclear atypia and hyperchromasia. Areas of osseous and cartilaginous differentiation have been reported (107). This subtype is non resectable by surgical intervention.

Biphasic MPM is characterized by the simultaneous presence of both epitheliod and sarcomatoid cells. This suggest that survival may correlate with the amount of sarcomatous component present within the tumor (**Figure 7c**) (102). This diagnosis is of clinical importance, because selecting patients for surgical intervention is dependent on the absence of a sarcomatoid component and tumor volume and resectability.

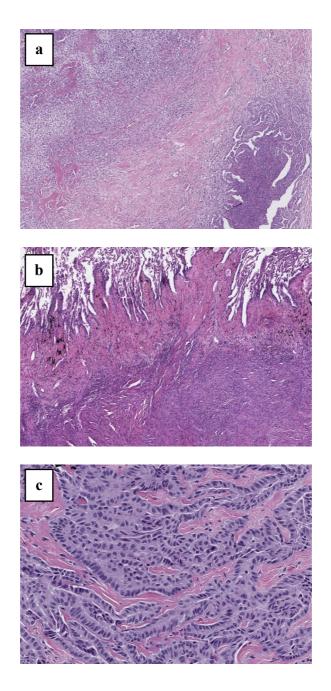


Figure 7. Immunohistochemistry imagines of mesothelioma; (a) epitheliod, (b) sarcomatoid and (c) biphasic subtype.

The staging classifications of MPM is based on the AJCC (The American Joint Committee on Cancer)/UICC (International Union Against Cancer) criteria (**Figure 8**) (108, 109). According to this, T classification is determined based on the extent of tumor invasion within the pleura and into adjacent thoracic structures so, T1 tumors, are those that remain confined to unilateral pleural surfaces. The T2 classification includes

extended tumors that involve ipsilateral parietal or visceral pleura with invasion of lung parenchyma or diaphragm muscle. T3 tumors are locally advanced and are extended to at least one of the following structures: endothoracic fascia, mediastinal adipose tissue, nontransmural invasion of the pericardium, or focal resectable soft tissue of chest wall. Finally, T4 are unresectable tumors with diffuse or multifocal chest wall soft tissue involvement, invasion of brachial plexus, bony components of chest wall or spine, mediastinal organs, contralateral pleura, or extension through diaphragm or pericardium. Unlike TNM staging of most solid tumors, due to the impracticality of measuring tumors with irregular and highly variable morphology, criteria for T classification of MPM do not include consideration of tumor size. N classification of MPM follows the lung cancer map (110). MPM are classified as N1 when invade pulmonary lymphatics that drain predictably and progressively through intraparenchymal and ipsilateral hilar lymph nodes; MPM N2 present metastasis in ipsilateral and midline mediastinal nodes; MPM N3 present metastasis in contralateral and extrathoracic stations. The lung map does not account for some nuances of MPM nodal invasion, however, MPM can even invade pulmonary parenchyma from visceral pleura following this metastatic pattern (111) TNM grouping criteria do not distinguish N1 from N2, although studies have demonstrated worse prognosis for N2 than N1 (112, 113). Nevertheless, evidence-based proposals have been made to refine N classification considering combined N1 and N2 involvement versus N1-only or N2-only disease (114), the number of involved nodes (112) or nodal stations (115), or the specific mediastinal stations involved (113). M classification of MPM indicates the absence (M0) or the presence (M1) of distant blood-borne metastasis. Usually, the presence in brain, bone, kidney and adrenal glands metastases is a rare condition, although it has been documented (116). Probably this is due to the

comparatively rapid and fatal progression of local T4 disease involving vital intrathoracic organs. (117).

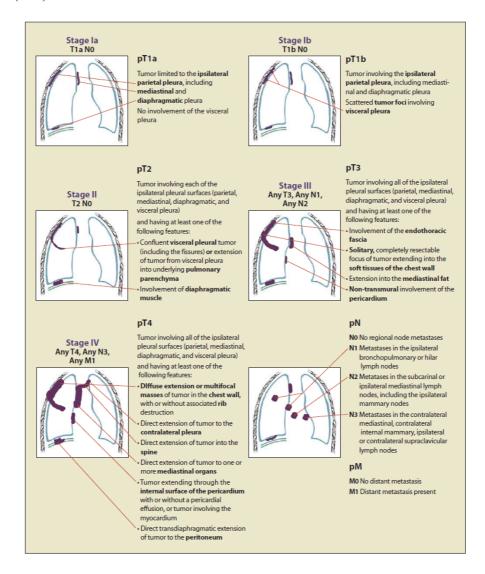


Figure 8. The proposed International TNM Staging System for MPM

1.1.5 Therapeutic approach

Only a minority of MPM patients is fit enough to be a surgical candidate and the indication for surgery has become stricter in the last years. Complete microscopic surgical resection is highly unlikely and MPM almost always recurs after surgery alone (118). Therefore, surgery is usually always part of a multimodal treatment strategy and is

considered only in patients with stage I/II MPM and only with epithelioid histology. Non-epithelioid subtypes are "unresectable".

The ideal surgical intervention to achieve maximal cytoreduction is still under debated (119). There are two types of surgical intervention for MPM, extrapleural pneumonectomy (EPP), and lung-sparing extended pleurectomy/decortication (EPD), but there is no difference in terms of survival between the two methods (120). Otherwise, many experts still prefer EPD, in part because of the preservation of lung parenchyma and theoretical postoperative functional improvement and capacity to tolerate further pulmonary insults (119 (**Figure 9**).

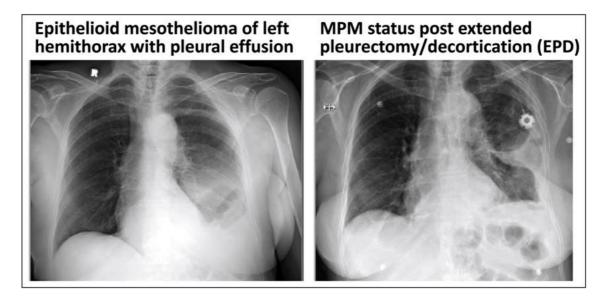


Figure 9. Demonstration of preservation of the lung parenchyma in epitheliod MPM before and after EPD

Most patients with mesothelioma are not offered surgery because of the extent of disease, advanced age, comorbidities, or poor performance status, and are considered for palliative chemotherapy instead. For more than fifteen years, the standard first line treatment, both for resectable and unresectable MPM, has been cisplatin-pemetrexed and it is currently the only regime approved by the Food and Drug Administration (FDA) for MPM. Cisplatin is a genotoxic chemoterapy that induces intra-strand DNA cross-linking,

causing DNA damage and interfering with DNA replication (121). Pemetrexed is a folate antimetabolite that inhibits three enzymes used in purine and pyrimidine synthesis. By inhibiting the formation of precursor purine and pyrimidine nucleotides, pemetrexed prevents the formation of DNA and RNA, both required for the growth and survival of normal and cancer cells. However, MPM is a highly heterogeneous cancer and chemotherapy treatment can cause additional selective pressure by eradicating only sensitive proliferative cells. Resistant cells remain and can still grow or regrow after treatment (122)

In a randomized study was observed that the median OS in the cisplatin-pemetrexed arm was 12.1 vs. 9.3 months in the control arm that received cisplatin mono therapy (P=0.020, two-sided log- rank test). The most common non-hematological toxicities, in both groups, were nausea, vomiting and fatigue within around 90% of patients experiencing grade-3 toxicity (123).

Although platinum-pemetrexed is an active agent in the first line treatment, chemotherapy may only alleviate symptoms but is not curative. Several studies have been conducted to improve the survival of MPM patients by combining the two classic drugs with drugs used for other tumors. VEGF signaling is an important concept in mesothelioma cell pathophysiology (124). The addition of anti-angiogenesis agents to chemotherapy has been tested in several clinical studies. In the phase III MAPS trial, addition of maintenance bevacizumab, an anti-VEGF antibody, to cisplatin-pemetrexed therapy in unresectable treatment naïve patients, prolonged median PFS from 7.3 to 9.2 months and median OS from 16.1 to 18.8 months. Unfortunately, in the bevacizumab combination arm more grade-3 toxicity occurred, like hypertension (23% vs. 0%) and thrombotic events (6% vs. 1%) (125).

In MPM there is no current evidence for maintenance chemotherapy and there is a lack of studies showing a better PFS or survival benefit. In a study with a cohort of 13 patients (out of 30 patients who started with platinum-pemetrexed), treatment with pemetrexed maintenance therapy (PMT) was associated to a better survival with a median OS of 8.5 vs. 3.4 months in the cohort without maintenance therapy. Grade-3 toxicity consisted of neutropenia, leucopenia and anemia. The only non-hematological grade-3 toxicity during PMT was fatigue (15%). The reason to stop PMT was disease progression (69%), toxicity (23%) and in patient's best interest (8%) (126).

Up today, there is no standard second line treatment in MPM, but if there is a good sustaining response at the time of initial chemotherapy interruption, the NCCN guidelines recommend the re-challenge of pemetrexed (if not administrated in the first-line). Other options like vinorelbine, gemcitabine, and immunotherapy (pembrolizumab and nivolumab-ipilimumab) could also be considered (1).

In MPM, prophylactic radiation therapy to prevent procedure-tract metastasis is not recommended on a routine basis (127). Radiotherapy could be considered in a trimodality treatment. In the NCDB, improved survival was associated to trimodality treatment including radiation therapy compared with surgical intervention and chemotherapy alone (103).

In conclusion, conventional treatments for MPM are ineffective and the prognosis is still very poor, with median survival for resectable pleural mesothelioma between 17 to 25 months and, for unresectable mesothelioma, between 9 to 12 months (128). Therefore, it is crucial to identify novel, well defined targets which can be used alone or in combinations with classic treatments.

In a study, which performed a semi-quantitative assessment of the inflammatory response in the tumor and in the stroma of 175 MPM patients, was found that patients who had a high-grade chronic inflammatory response in the stroma (n=59) presented improved survival compared with those who had a low-grade chronic inflammatory response (n=116; median OS, 19.4 months vs 15.0 months; p=0.01) (129).

Several studies even proposed the prognostic role of lymphocytes and macrophages and the presence of immuno-suppression in MPM through analysis of T-cell-inhibitory receptors and chemokines (130). In a study by Bueno et al (85), were identified, in 212 MPM patients, 4 different phenotypic clusters of molecular expression with divergent associated survival and mutational characteristics. Programmed death-ligand 1 (PD-L1) was expressed in 39% of patients and was associated with a worse survival. PD-L1 expression was higher in non-epithelial MPMs (85).

This has led to considering immunotherapy as a possible treatment strategy for MPM. Clinical trials using cytotoxic T-lymphocyte–associated protein 4 (CTLA-4) inhibitors failed to improve survival in MPM (131), while subsequent trials suggested that PD-L1 inhibitors may benefit some patients (132).

Systemic dendritic cell immunotherapy is another immunotherapy treatment that has shown promising result. This therapy lead to a significant antitumor immune responses in MPM as demonstrated in a pilot study of 10 advanced non-sarcomatoid MPM patients. These patients underwent to an 8-10-week course of adjuvant cyclophosphamide and autologous dendritic cell immunotherapy which resulted associated with impressively prolonged survival, with seven patients survived for at least 24 months and with two patients still alive after 50 and 66 months (133).

1.1.6 Prognosis

As discussed above, for most patients, treatment for MPM is palliative. for this reason, every intervention must take into account improved life expectancy with quality of life. A CT scan, in patients who receive chemotherapy, is often performed mid-cycle to assess response. However, MPM is difficult to monitor using conventional CT scanning, so a biomarker that could measure response to chemotherapy or predict recurrence would offer considerable advantages. Currently, the clinical prognostic scoring systems used for MPM patients are established by European Organization for Research and Treatment of Cancer (EORTC) and Cancer and Leukemia Group B (CALGB) (134, 135). This scoring system define that non-epithelioid histology, male gender, anemia, thrombocytosis, leukocytosis, and elevated LDH are associated with poor prognosis. Otherwise, overall survival remains dismal and there is still a need for better prognostic biomarkers. The research in this field has focused both on circulating blood-based markers and on genetic alterations identifiable on tumor tissue that could predict patient's outcome. The soluble mesothelin-related peptides (SMRPs) are found in normal mesothelial cells and are overexpressed in various cancers. SMRP has been analyzed in serum and was studied as a diagnostic marker, with promising results (136), and even as a prognostic factor, but for the latter the results are inconclusive. Indeed, no correlation has been shown between serum SMRP level and progression-free or overall survival in several study (137-138). However, some study found that the SMRP level was inversely associated with overall survival, but, in multivariate analysis limited to epithelial MPM, the prognostic value of SMRP on overall survival was lost (139, 140). Thus, suggesting that histology remains a critical determinant of prognosis.

Osteopontin is a cell adhesion protein that mediates, via interaction with integrin and CD44 receptors, cell-matrix interaction and cell-signaling (141). The prognostic value of osteopontin has been analyzed in several studies which have shown that low baseline plasma levels of the protein were independently associated with favorable progression-free and overall survival (137).

HMGB1 is a damage associated molecular pattern (DAMP) and a key mediator of inflammation. Studies showed that HMGB1 serum levels were higher in MPM patients than in control group with asbestos-exposure (142). Furthermore, another study demonstrated a possible prognostic role of HMGB1 given that, at a cutoff value of 9 ng/mL, there was a significant negative correlation between serum HMGB1 level and survival (143).

In conclusion, SMRP, osteopontin and HMGB1 have been associated with poor prognosis and show potential as prognostic markers in MPM. However, none of them are currently used routinely in clinic for this purpose, as majority of them have not been validated prospectively and their superiority used alone has not been proven over the conventional prognostic EORTC or CALGB models (144).

MicroRNAs (miRNAs) are another group of markers being investigated as diagnostic and prognostic biomarkers due to their tissue specificity, to their ability to classify several types of tumors and to their stability in circulation (145). Even in the MPM, deregulated miRNAs have shown potential as diagnostic and prognostic factors, like miR-29 which if it is up regulated predicts longer survival (146). MiRNAs can be combinated each other in order to improve their prognostic value. For example, in a subsequent study involving MPM patients undergoing surgery, a six-miRNA signature including miR-21-5p, miR-23a-3p, miR-30e-5p, miR-221-3p, miR-222-3p, and miR-31-5p was found to predict an

overall survival of more than 20 months (147). However, the use of miRNAs as a biomarker is not yet applicable to clinical practice and larger studies are required to validate their diagnostic and prognostic values.

Taken together, there has been a strong interest in the research of circulating biomarkers due to their non-invasiveness, but many of them have varying levels making them unsuitable for diagnostic or prognostic use. For this reason, a strong interest has also been placed in the search for tumor markers on biopsy sections. Genetic aberrations, identifiable by immunoistochemestry (IHC) or FISH analysis on tumor tissue, were studied as prognostic factor.

One of the most common events found in MPM is the 9p21 deletion. The *CDKN2A* gene contained in this region, encodes for p16 protein, a well-established tumor suppressor in a variety of tumor types, including MPM. This genetic aberration has been confirmed in several studies and has been correlated with poor patient prognosis (148, 149).

Cedres *et al.* have been demonstrated that the expression of WT1 gene in tumor section of MPM is associated with a better survival, but they don't find a correlation between calretinin expression, another promising prognostic factor in MPM, and survival (150) The expression of the Programmed death-ligand 1 (PD-L1) in MPM has been correlated with shorter survival, compared to MPMs without this condition (151-153). Tumor cells that express PD-L1 on their extracellular surface downregulate the anti-tumor activity of infiltrating lymphocytes. Positive PD-L1 expression, determined by IHC (>1% tumor cell staining), is reported in 11–72% of MPMs (131-138) and allows patients to be subjected to immunotherapy treatments such as Pembrolizumab (an anti-PD-1 monoclonal antibody). Instead, nivolumab (another anti-PD-1 agent) is accepted as salvage therapy regardless of PD-L1 IHC findings. (154).

Finally, interest was placed on the BAP1 gene. BRCA-1-associated protein 1 (BAP1) is a deubiquitinase enzyme, involved in the regulation of various cellular pathways (155). The prognostic role of BAP1 expression in MPM has been studied, but the results are still confusing.

1.2 BAP1

1.2.1 Structure and function

BRCA1-associated protein 1 (BAP1) was identified in 1998 and, from the first studies, it was reported that it had a growth suppression activity in breast cancer cells. Moreover, it was also observed that this anti-tumor activity was carried out in cooperation with BRCA1 in cultured cells.

BAP1 is a deubiquinating enzyme, with the gene is located on the short arm of chromosome 3 (3p21.1) and consists of 17 exons (157). BAP1 protein is a 90 kDA, nuclear localized deubiquitinating enzyme (DUB) consisting of 729 amino acids (2). BAP1 protein has three main domain: an N-terminal ubiquitin carboxyl hydrolase domain (UCH); a host cell factor 1 (HCF1) binding domain (HBM) in the middle portion; a C-terminal domain (CTD) containing a coiled-coil motif for interaction with ASXL1/2 and a nuclear localization signal (NLS). There are also various binding regions to other protein interaction partners (**Figure 10**) (158-160).

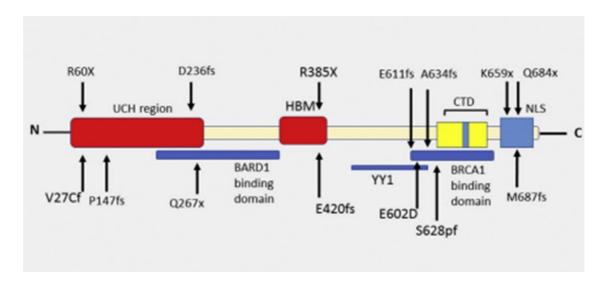


Figure 10. Schematic structure of BRCA1-associated protein-1 (BAP1) domains and locations of reported germline mutations.

Many cellular processes are regulated by protein ubiquitination and deubiquitination, a reversible post-translational modification. BAP1 works as a tumor suppressor by its deubiquitinase activity, regulating target genes involved in transcription, cell cycle control, DNA repair and cellular differentiation (**Figure 11**) (161).

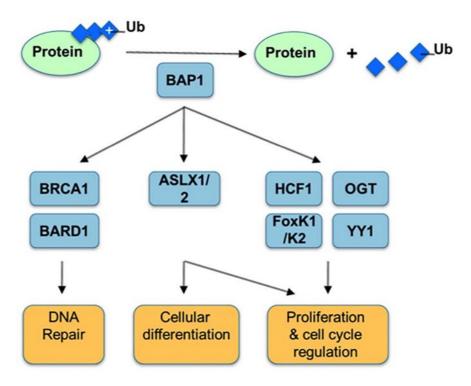


Figure 11. Schematic rapresentations of BRCA1-associated protein 1 activity.

BAP1 interacts with numerous proteins and carries out its anti-tumor activity in various ways. BAP1 can activate the transcriptional regulator HCF1 (162). Once activated, HCF1 modulates chromatin architecture by recruiting histone-modifying complexes and activating transcription factors such as the E2F family, which controls G1/S phase progression in the cell cycle (158). BAP1 also deubiquitinates O-linked N-acetylglucosamine transferase (OGT), which in turn modifies and activates HCF1 (163). BAP1 has been shown to form a ternary complex with HCF1 and transcription factor Ying Yang 1 (YY1), that controls cellular proliferation (164). This complex is recruited to the promoter of COX7C which encodes a component of the mitochondrial respiratory chain (164). Furthermore, BAP1 is involved in cell proliferation and cell cycle control even by forming a ternary complex with HCF1 and the forkhead transcription factors FoxK1/K2 (159)

BAP1 form the polycomb group repressive deubiquitinase complex (PR-DUB) by interacting with ASXL1/2 and regulate physiological processes such as stem cell pluripotency, embryonic development, self-renewal and differentiation. This group of protein contain even a polycomb-repressive complexes (PRCs) that ubiquitinate histones and lead to gene silencing (158). There is a transcriptional balance and control due by ubiquitination by PRCs and deubiquitination by PR-DUB (158).

BAP1 is involved even in the DNA damage repair process. BAP1 form a complex with several recombination proteins including Breast Cancer type 1 (BRCA1) and BRCA1-associated RING domain protein 1 (BARD1), which promotes E3 ubiquitin ligase activity to regulate DNA damage response (165).

Initially, it was thought that BAP1, given its NLS domain, was present only in the nucleus, where it is involved in several anti-tumor processes such as DNA repair.

However, in a recent study conducted by Bonomi et al, the presence of BAP1 was also found in the cytoplasm. Here, BAP1 is found in the endoplasmic reticulum (ER) fraction where it is involved in the apoptotic process (166, 167).

In detail, cytoplasmic BAP1 regulates Ca²⁺ transfer from the ER, where Ca²⁺ is normally stored in the cell, to the cytoplasm by deubiquitinating the IP3R3 receptor channel. Ca²⁺ is released in areas of the ER, called MAMs (mitochondrial-associated membranes), that are in close contact with the mitochondrial outer membrane. Here, Ca²⁺ flows inside the mitochondria first through the voltage-dependent anion channel (VDAC) channel localized on the outer mitochondrial membrane and then by the mitochondrial uniporter channel (MCU) located on the inner mitochondrial membrane.

This process is finely regulated, because Ca²⁺ in the mitochondria is necessary for the Krebs cycle, but if DNA damage occurred and cannot be repaired, higher amounts of Ca²⁺ is released from the ER through the IP3R3 leading to high mitochondrial Ca²⁺ concentrations. Finally, mitochondria release cytochrome c into the cytosol, thus inducing the apoptotic process (167) (**Figure 12**). In case of BAP1 inactivating mutation, cells cannot release enough amounts of Ca²⁺ to induce the apoptotic process and with low level of Ca²⁺ the Krebs cycle is impaired. This cause the cells to switch into anaerobic glycolysis (Warburg effect), a metabolic shift that favors malignant growth (166).

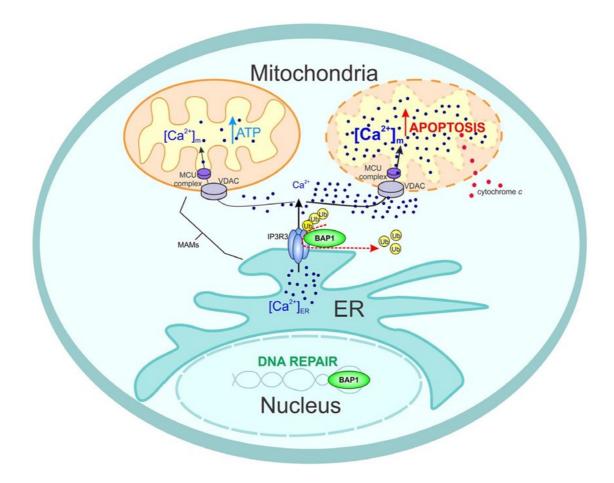


Figure 12. Representations of BAP1 activity in the cytoplasm.

Subsequently, Zhang et al showed that cells with reduced BAP1 activity also have impaired ferroptosis (168) providing an additional mechanism by which BAP1-mutated cells escape cell death (155). Thus, cells with reduced or absent BAP1 activity could accumulate more DNA damage (166) since they cannot properly repair the DNA (167, 169) and, at the same time, they cannot execute apoptosis, which normally eliminates cells that contain genetic mutations. This leads to a possible transformation in cancer cells

1.2.2 BAP1 and MPM

In a study by Carbone et al it was shown that the high incidence of MPM in some Cappadocian and American families was due to the presence of germline mutations of the BAP1 gene. The study demonstrated the existence of a predisposition to MPM which was transmitted in an autosomal dominant fashion (88, 170-172). Like other tumors suppressors, affected individuals inherited a non-functional BAP1 allele. The remaining functional allele was inactivated later in life.

Since 2011, over 600 articles have confirmed and expanded the pathogenic role of BAP1 mutations in MPM and in other cancers (10, 172-175).

This condition was named the "BAP1 cancer syndrome" (BAP1-TPDS) because affected family members present an high risk of developing tumors, predominantly MPMs, uveal melanomas (UM), cutaneous melanoma (CM) and renal cell carcinomas (RCC). While, basal cell carcinoma, breast carcinomas, cholangiocarcinomas, sarcomas, and various types of brain tumors were less frequently (10, 164, 173-175). In addition, Wiesner et al. recognized that a benign atypical skin melanocytic tumor was associated with germline BAP1 mutation (176). This melanocytic lesion was histologically between benign Spitz nevus and malignant melanoma, so was named Melanocytic BAP1-mutated Atypical Intradermal Tumor (MBAIT), or atypical Spitz tumor (AST) (176). The detection of MBAIT, which develop between 20-30 years, allows dermatologists to suspect the diagnosis of BAP1-TPDS, which is then verified and confirmed by DNA sequencing (173-175, 177).

The incidence of these tumors in people with BAP1 syndrome is very high and, in most cases, they have a positive family history of at least two of the main cancers (including UM, MPM, MBAIT, CM, and RCC) among their first- or second-degree relatives (178). Moreover, affected individuals may have more than one type of primary cancer (172, 173, 178).

Furthermore, somatically mutated (acquired mutations occurring during tumor cell growth) BAP1 has been found in approximately 60% of MPMs, underscoring the critical role that BAP1 has in preventing MPM growth (85, 87, 179-181).

Several different alterations in the BAP1 gene have been described, including large deletions of exons leading to loss of the N-terminal region, focal deletions, frameshift mutations due to insertions or deletions, splice site mutations, and base substitutions leading to non-sense and missense mutations (158, 167). In MPM, the BAP1 gene is commonly lost by chromosomal deletion, and more than 70% of reported germline BAP1 mutations are truncation (167, 178).

From these studies it was found that BAP1 plays a key role in MPM, therefore it was also analyzed as a possible diagnostic and prognosis factor.

BAP1 status determinations by IHC has improved the ability to diagnose MPM. BAP1 wild-type (BAP1^{WT}) is found in the nucleus and the cytoplasm, with strong nuclear staining and less intense cytoplasmic staining (87), while MPM BAP1 loss are determined by the complete absence of staining or by cytoplasmic staining without nuclear staining. BAP1 positivity nuclear staining is a specific and reliable marker to distinguish benign atypical mesothelial hyperplasia at its earliest stages of development from mesothelioma because benign cells always express BAP1 in the nucleus (87, 182-184). Overall, approximately 70% of epithelial and 50% of sarcomatoid MPMs contain somatic BAP1 gene mutations, resulting in an absence of BAP1 nuclear staining (87, 182-186).

BAP1 loss is highly specific for MPM and this is helpful in the differential diagnosis with carcinoma of the lung, breast, and stomach. Only one study showing BAP1 loss in just 1% of lung cancers (187-189). Furthermore, multiple studies have indicated that BAP1 loss is highly specific for MPM in the differential diagnosis with ovarian carcinoma (188,

190, 191). However, multiple other malignancies (including renal cell carcinoma and melanoma) also frequently show BAP1 loss (158, 187, 192). These tumors may occasionally metastasize to the serosal surfaces, thus careful consideration of the clinical and radiographic data, tumor morphology, and lineage-specific markers is critical when interpreting BAP1 IHC (154).

In addition to its diagnostic role, multiple studies indicate that BAP1 even show a prognostic value and the preponderance of them supports a better outcome in MPM with BAP1 loss (185, 193-199). Instead, one study found that BAP1 loss was correlated to a poorer prognosis and suggests that the improvement in prognosis identified in the other studies was likely due to confounding factors (200). Importantly, these studies have not generally discriminated between germline and sporadic BAP1 loss.

Specific studies showed that germline *BAP1* mutations were associated to better prognosis compared to non-germline-mutated MPM, irrespective of tumor site, with 5-year overall survival of 47%, and 7%, respectively (90, 201).

In addition to germline and somatic gene mutations (202), post-transcriptional mechanisms are involved in the regulation of BAP1 expression. For instance, 25% of MPM with negative nuclear IHC staining of BAP1 are also negative for BAP1 gene mutations (158). MiRNA are small noncoding RNAs that control gene expression at post-transcriptional level binding by 7-8 nucleotides the complementary ones in the 3'-untranslated regions of their targets. MiRNA may function as either oncogene or tumor suppressors depending on target genes and cancer type (203). It has been demonstrated in pre-clinical studies that miR-31 is a post-transcriptional regulator of BAP1 (204-206).

2. AIM OF THE THESIS

BAP1 loss has been detected in about 60% of MPM and its prognostic role has been amply investigated in order to find a potential link between BAP1 status and patient outcome. However, the clinic relevance of BAP1 loss-of-function in MPM still remain under debates, with conflicting reports that it may be associated with epithelioid subtype. In order to clarify the prognostic role of BAP1 expression in MPM, we conducted a systematic review and meta-analysis for summarize the available evidence in this setting. To improve the performance of BAP1 in distinguishing patients with a worse outcome, BAP1 status has been combined with the tissue expression of miR-31, a miRNA involved in the BAP1 regulation at post-transcriptional level.

3. MATERIAL AND METHODS

3.1. Cohort study

3.1.2. Study population

Between 2003 and 2019, clinical records of 85 patients with a histological diagnosis of MPM were retrospectively collected at the Clinical Oncology, Polytechnic University of Marche, AOU Ospedali Riuniti.

Formalin-Fixed Paraffin-Embedded (FFPE) tissues with BAP1 IHC staining (n=60) were collected at the Clinic of Pathological Anatomy before any treatment including the first-line chemotherapy.

The collected patients characteristics and clinical-pathological features were: BAP1 expression (BAP1 retained vs BAP1 loss, as assessed by IHC), age, gender, smoking status (current/former smokers vs never smokers), asbestos exposure, histotype (epithelial, biphasic and sarcomatoid), side of disease, clinical TNM stage (stage IV versus others [VIII edition]) (207), treatment modality (chemotherapy plus surgery versus only chemotherapy), type of chemotherapy, response to chemotherapy according to modified Response Evaluation Criteria in Solid Tumors for assessment of response in malignant pleural mesothelioma (mRECIST criteria) (208).

3.1.3. Immunohistochemical analysis

All selected MPM histological samples and relative diagnoses were reviewed by an experienced mesothelioma pathologist and divided into epithelioid (e-MPM), sarcomatoid (including desmoplastic, s-MPM) and biphasic (b-MPM) MPM according to the 2015 World Health Organization (WHO) classification (209). For the inclusion of MPM sample in the b-MPM subgroup was required at least the presence of both sarcomatoid and epithelioid components in 10% of the tumor.

All samples were FFPE and, for each patient, a single 4-mm-thick paraffin section was cut from the sample with the greatest amount of tumor tissue. All sections were deparaffinized and rehydrated in graded concentrations of xylene and ethanol. Sections were coated with 1:50 mouse monoclonal BAP1 antibody (clone C4:sc-28383; Santa Cruz Biotechnology, USA) and incubated at 4°C overnight. Then, automated IHC was performed on Omnis platform (Agilent, USA).

BAP1 IHC status was considered as "positive/retained" if there was an unambiguous positive nuclear staining in any number of tumor cells, and "negative/loss" if the nuclear staining was absent in neoplastic cells. Tumor cells with cytoplasmic reactivity without a clear nuclear staining were considered negative. Non-neoplastic cells, such as vascular endothelium, fibroblasts or inflammatory cells, were considered as internal positive control.

3.1.4. Mir-31 assay

Total RNA was extracted from FFPE tissue samples (10–100 µg) using the RecoverAllTM Total Nucleic Acid Isolation Kit for FFPE (Thermo Fisher Scientific) according to the manufacturer's protocol. The RNA concentration and purity were determined in the Nanodrop 1000 spectrophotometer (Thermo Fisher Scientific). The retro-transcription reaction of miR-31 was performed using the High-Capacity cDNA Reverse Transcription Kit (Applied Biosystems). The reaction mix was prepared using 20 ng/ μ l of total RNA and incubated in the Biometra thermal cycler. The thermal protocol was the following: 16°C for 30 minutes, 42°C for 30 minutes and 5 minutes at 85°C.

Quantitative RT-PCR (qPCR) was performed using the TaqMan Fast Advanced Master Mix (Applied Biosystems) with U6 as the housekeeping gene. The qPCR assays were carried out using the Mastercycler EP Realplex instruments (Eppendorf) using the following thermal protocol: 95°C for 10 minutes for the polymerase activation, then 40 cycles at 95 °C for 15 seconds and at 60°C for 1 minute. MiR-31 levels were considered as percentiles and then divided into quartiles, in order to quantify the miR-31 expression for each sample. Because the 25th and 50th percentile showed similar miR-31 expression, the respective inter-quartiles were used as reference and results of remaining samples were expressed as relative level using the Δ CT method (2^{- Δ CT}). MiR-31 expression was considered "high" if miR-31 levels were over the 75th percentile and "low" if miR-31 levels were below the 75th percentile.

3.1.5. Statistical analysis

Discrete data were expressed either as mean, standard deviation, minimal and maximal values (if normally distributed), or as median, quartile and range (if not-normally distributed). Categorical variables were reported as either fractions or percentage. Differences between groups were analyzed using means of chi-squared method or Fisher's exact test for categorical variables and unpaired Student t test, the Mann-Whitney U and one-way analysis of variance (ANOVA) with Tukey post-hoc analysis test for continuous variables. OS was defined as the temporal interval between the date of the first cycle of first-line chemotherapy and the date of death or censoring at the date of last follow-up of alive patients. PFS was considered as the time from the first cycle of first-line chemotherapy until clinical or instrumental disease progression or last follow-up. Survival curves were constructed using the Kaplan–Meier method and log-rank method was used to assess difference between subgroups. Patients with missing survival data were excluded from the analysis.

Multivariate Cox regression analysis was used to evaluate independent predictors of survival. Nonsignificant prognostic factors were excluded from the model using backward elimination. A p value < 0.05 was considered statistically significant.

Statistical analysis was performed by using SPSS for Windows version 19.0 (SPSS Inc, Chicago, IL) and GraphPad PRISM (GraphPad Software, Inc., La Jolla, CA),

3.2. Systematic review and meta-analysis

3.2.1. Literature search

We conducted a systematic review and meta-analysis following the procedures of the Preferred Reporting Items for Systematic reviews and Meta-Analyses (PRISMA) statement and the Meta-analysis of Observational Studies in Epidemiology (MOOSE) guidelines.

Relevant literature was identified by systematic search of PubMed (Medline) and Embase datasets until September 28, 2019, considering only English-written publications. Our search strategy included the following mesh terminology: (mesothelioma) AND (BAP1 OR bap1 OR UCHL2 OR hucep-6 OR HUCEP-13 OR "BRCA1 associated protein-1" OR "ubiquitin carboxy-terminal hydrolase") AND (mortality OR mortalities OR fatality OR fatalities OR death OR survival OR prognosis OR "hazard ratio" OR HR OR "relative risk" OR RR OR prognosis OR progression OR recurrence OR PFS OR OS).

3.2.2. Study selection

The inclusion criteria were as follows: (i) both prospective and retrospective full-text observational cohort studies regarding MPM; (ii) immunohistochemical (nuclear positivity) investigation of BAP1 expression on tumor tissue (iii) enough data to calculate the correlation of BAP1 status with OS.

Studies conducted on other cancer than MPM (e.g. peritoneal mesothelioma), with no report data on patient outcomes or that had investigated BAP1 status by other methods than IHC (e.g. DNA sequencing) were excluded. When more than one study from the

same institute/authors was reported, only the most recent or the highest quality study (reporting a higher number of patients) was included in the analysis.

Title and abstract of the primary studies identified in the electronic search were independently screened by two reviewers. Duplicate studies were excluded. Any disagreement or discrepancies was resolved by a third reviewer or contacting the study authors when necessary.

The reference list of all identified documents was scrutinized, with the aim of identifying additional potentially eligible studies.

3.2.3. Data extraction and meta-analysis outcomes

For each study we extracted author name, year of publication, number of patients, mean age, gender, histotype, HR and 95 % CI of BAP1 immunohistochemical expression for OS.

Our primary objective of the meta-analysis was to determine the HR of death in patients whose tumor retains BAP1 expression (BAP1 retained) compared to patients whose tumor does not express BAP1 (BAP1 loss).

In the BAP1 retained MPM patients a HR > 1 indicates higher risk of death and a HR < 1 lower risk. We were considered only HR values obtained through multivariate analysis (adjusted for histotype). Finally, if the survival information was only available in Kaplan–Meier curves, we collected HR only when epithelioid and non-epithelioid subtypes were separately analyzed.

3.2.4. Data analysis

DerSimonian-Laird random-effects model was used for calculating pooled HRs with correspondent 95% CIs of death between BAP1 retained versus BAP1 loss MPM patients. Because of the heterogeneity of the observational studies, this method was chosen a priori. To determine inconsistency across the results of the studies was used the Higgins I^2 index and chi-square statistics. Funnel plots were drawn to evaluate the publication bias of the included literature. All p values were two sided and p < 0.05 was considered significant. Data analyses and generation of forest plots were performed using R 3.6.0 (R Foundation for Statistical Computing) and RevMan 5.2 (Copenhagen, Denmark).

4. RESULTS

4.1. Demographic and clinic-pathological characteristics of MPM patients

Our cohort consisted of 85 patients (81% male). Fifty-one patients had progression of disease and 41 patients died at a median follow-up time of 41.28 months (interquartile range [IQR]: 14.64-not reached). Median age at diagnosis was 71 years (range 52-81 years). Median OS was 14.76 (95% CI: 7.92–26.52) months and median PFS was 4.56 (95% CI: 2.88–9.48) months. Asbestos exposure was documented in 55% of patients, and 57% of patients were current/former smokers. Epithelioid MPM (e-MPM) represented 66% of cases (n=56), while 15% was represented by sarcomatoid MPM (s-MPM) (n=13) and 10% was biphasic MPM (b-MPM) (n=10). Only 18 patients (21%) underwent surgical resection. 74% of patients (n=63) received first-line platinum-based chemotherapy in association with pemetrexed (83%) and 17% (n=20) pemetrexed in monotherapy (17%). Demographic and clinicopathological characteristics are summarized in **Table 3**.

Characteristic	Total patients (n=85)		Patients with BAP1 staining available (n = 60)				
			BAP1 loss (n=37)	В	AP1 retained (n=23)	P valu	
	n	%	n	%	n %		
Age							
\leq 70 years	41	48	13	35	10 43	0.11	
> 70 years	44	52	24	65	13 57	0.11	
Gender							
Male	69	81	28	76	20 87	0.46	
Female	16	19	9	24	3 13	0.46	
Smoking							
Current/former	48	57	21	57	13 57	1.00	
Never	31	36	14	38	9 39	1.00	
Unknown	6	7	2	5	1 4		
Asbestos exposure							
Yes	47	55	19	51	9 39	0.51	
No	38	45	18	49	14 61		
NO	36	43	16	49	14 01		
Histotype							
Epithelioid	56	66	32	87	11 48		
Sarcomatoid	13	15	2	5	5 22	0.01^{a}	
Biphasic	10	12	3	8	5 22		
Unknown	6	7	0	0	2 8		
Clinical stage							
I/II/III	64	21	31	84	19 83	0.25	
IV	11	13	1	3	3 13	0.35	
Unknown	10	12	5	13	1 4		
Surgery							
Yes	18	21	10	27	3 13	0.01	
No	66	78	26	70	20 87	0.31	
Unknown	1	1	1	3	0 0		
First-line							
chemoterapy							
Platinum-based							
chemoterapy	63	74	29	78	18 78	0.22	
Pemetrexed	17	20	6	16	4 18		
Other	1	1	1	3	0 0		
Unknown	4	5	1	3	1 4		
BOR							
PR	10	13	7	19	2 10	0.58	
SD/PD	68	87	29	81	18 90		

Table 3. Patient demographics and baseline clinical characteristics.

^a Comparison between epithelioid subtype and non-epithelioid subtype (sarcomatoid and biphasic).

4.2. Outcomes according to BAP1 status

Immunohistochemical staining for BAP1 expressions was conduct only for patients that had adequate tissue (n=60, 70%). Immunohistochemical BAP1 positive/retained (≥1%) was found in 38% of cases (n=23) and BAP1 negative/loss in 62% of cases (37%). BAP1 loss was mainly associated with the epithelioid subtype (p=0.01; 74% of e-MPM were BAP1 negative/loss and 33% of non-epithelioid MPM were BAP1 negative/loss; **Table**3). Among patients with BAP1 loss and BAP1 retained there was no difference in age, gender, smoking status, asbestos exposure, clinical stage, and surgery. In **table 3** is summarized BAP1 expression status according to clinical data.

At univariate analysis, there were no difference in terms of OS between the two group (BAP1 retained vs BAP1 loss), with a median OS of 18.1 months (95% CI: 11.2-25.8) for positive BAP1 expression and 14.8 months (95% CI: 10.7-29.3 months) for negative BAP1 expression (p=0.17, **Figure 13a**). At multivariate analysis, after adjusting for age and histotype, no significant differences were observed among the two groups for OS (HR 1.09, 95% CI: 0.51-2.31, p=0.81). Conversely, non-epithelioid histology held its independent negative prognostic value for OS (HR 7.03, 95% CI: 2.48-19.86, p=0.0002) Looking at PFS, both groups showed a similar trend, with median PFS of 5.04 months (95% CI: 3.00-8.64) for BAP1 retained tumors and 6.36 months (95% CI: 3.24-11.64) for BAP1 loss tumor (p=0.14, **Figure 13b**). Even for PFS, at multivariate analysis, non-epithelioid histology was the only independent prognostic factor (HR 3.58, 95% CI: 1.58-8.14, p=0.002).

Similarly, no differences in response to treatment (BOR) were noted (10% in BAP1 retained patients versus 19% in BAP1 loss, p 0.58).

In the platinum-based chemotherapy subpopulations (n=47) were no differences according to BAP1 status (18.1 months vs 14.8 months, p=0.3)

Finally, for the 12 patients who underwent surgery (before or after chemotherapy), we found an improved OS for patients with BAP1 loss (median OS of 25.9 months, 95% CI: 11.1-not reached, vs 5.5 months 95% CI: 5.0-not reached for BAP1 retained group, p=0.02). However, after adjusting for histotype, BAP1 status has lost its prognostic value (p 0.14).

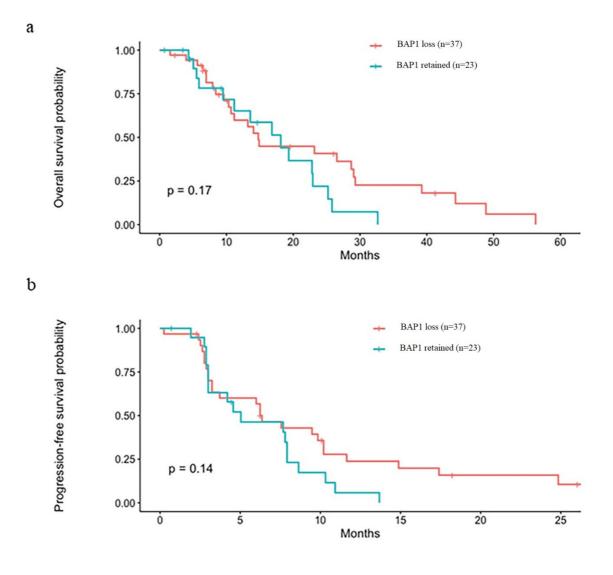


Figure 13. Kaplan-Meier curves of survival according to BRCA1 associated protein-1 (BAP1) expression. (a) Overall survival. (b) Progression-free survival.

4.3. Meta-analysis results

4.3.1. Search results and study population of included studies

From systematic research was extracted 346 potentially relevant articles and, after the exclusion of duplicates, 243 were considered for meta-analysis. After screening of title and abstract, 19 articles were selected for full-text review. Among them, 14 articles were excluded, since they evaluated BAP1 status with techniques other than IHC (n=5) (85, 180, 210-212), took into account only germline BAP1 mutations (n=2) (90, 92), did not report any survival analysis (n=1) (30), confounding factors were not taken into account in a multivariate analysis (n=4) (194, 199, 213, 214) and two because they were redundant from the same institution. Finally, five studies for a total of six cohort, (87, 185, 196, 198, 200) qualified all the selection criteria and were included for the meta-analysis.

The PRISMA flow chart showed the detailed literature search steps (Figure 14).

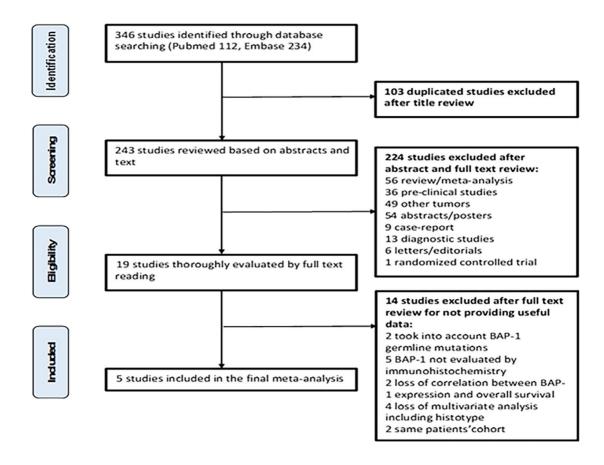


Figure 14. PRISMA flow chart summarizing the process for the identification of eligible studies for the meta-analysis.

All studies were published after 2015 and assessed for BAP1 status by IHC on whole-sections or tissue microarrays (185, 196). The number of adjustments in the multivariate analyses ranged from 1 to 3 and, in particular, histotype was always included.

We also added our cohort to the meta-analysis, thus, the overall populations consisted of 690 MPM patients. Seventy-eight per cent (n=543) were male and the proportion of epithelioid MPM was 62 % (n=428). Detailed descriptions of the 5 included studies (in addition to ours) are summarized in **Table 4**.

	Forest et al.	Pulford et al.	Farzin et al.	McGregor et al.	Nasu et al.
Year	2018	2017	2015	2015	2015
Country	France	Australia	Australia	USA	USA
Patients	117	81	229	111	92
Age ^a Total cohort BAP1 loss BAP1 retained	73 (43-92) NA NA	74 (35-94) 74 (53-92) 74 (35-91)	72 (35-93) 70 (35-93) 75 (44-93)	70 (43-89) 69 (43-89) 69 (45-81)	62 (20-85) 62 (20-82) 62 (23-85)
Gender ^a Male Totale cohort	88 (79 %)	63 (78 %)	188 (82 %)	87 (78 %)	67 (73 %)
BAP1 loss BAP1 retained	NA NA	39 (62 %) 24 (38 %)	83 (44 %) 105 (56 %)	57 (65 %) 30 (35 %)	44 (66 %) 23 (34 %)
Female Total cohort BAP1 loss BAP1 retained	29 (21 %) NA NA	18 (22 %) 8 (44 %) 10 (56 %)	41 (18 %) 23 (56 %) 18 (44 %)	22 (25 %) 11 (50 %) 11 (50 %)	25 (27 %) 17(68 %) 8(32 %)
Histotype ^a e-MPM					
Total cohort BAP1 loss BAP1 retained	87 (74 %) 62 (71 %) 23 (26 %)	57 (70 %) 35 (61 %) 22(39 %)	120 (53 %) 75 (63 %) 45 (37 %)	58 (52 %) 47 (81 %) 11(19 %)	64(70 %) 48(75 %) 16(25 %)
Non e-MPM Total cohort BAP1 loss BAP1 retained	30 (26 %) 6 (20%) 23 (77 %)	24 (30 %) 12 (50 %) 12 (50 %)	109 (47 %) 31 (28 %) 78 (72 %)	53 (48 %) 21 (40 %) 32 (60 %)	28(30 %) 13(46 %) 15(54 %)
Variables included in multivariate analysis	BAP1 loss, histotype, absence of necrosis, nuclear grading	BAP1 status, age, gender, histotype	BAP1 status, age, gender, histotype	BAP1 status, histotype	BAP1 status, histotype
OS (months) ^{a,b} BAP1 loss	17.9 (e-MPM) p = 0.034; 5.3 (non e-MPM) p = 0.60	6 (surgical sections) p = 0.015; 8 (cytology	16.1 (all patients) p < 0.01	NA	NA
BAP1 retained	12.4 (e-MPM) p = 0.034; 14.1 (non e-MPM) p = 0.60	sections) p = 0.205 11 surgical sections) p = 0.01; 9(cytology sections) p = 0.205	6.3 (all patients) p < 0.01	NA	NA

Table 4. Characteristics of studies included in the meta-analysis.

^a Continuous variables are reported as mean, minimal and maximal values; categorical variables are reported as absolute numbers and percentage; OS is reported as median.

b OS was calculated from the date of diagnosis in two studies (Forest et al., McGregor et al.) and from the date of surgery in other two studies (Pulford et al., Farzin et al.). The numeric value of median OS referred to the whole cohort of patients was not reported in the studies by McGregor et al. and Nasu et al.

4.3.2. Association with clinical-pathological characteristics and prognostic role of BAP1

Overall, BAP1 loss was the most represented status (388 out of 690, 56%) and was strongly associated with the e-MPM subtype (p for difference<0.0001). In the total populations, BAP1 loss was found in 70 % of epithelioid MPM and in 34 % of non-epithelioid MPM.

The median OS ranged between 6 months (200) and 17.9 months (198) and no association between BAP1 expression and gender was found (p for difference 0.94).

Pooling data from the 7 cohorts, BAP1 loss was not associated with a lower risk of all-cause mortality. The pooled HR was 1.11 (95 % CI, 0.76–1.61; p 0.60, I²=78 %, p for heterogeneity=0.0001, random effect model) (**Figure 15a**) which indicate that BAP1 expression had no obvious impact on patient survival when histotype is taken into account. The funnel plot showed no evidence of publication bias (**Figure 15b**).

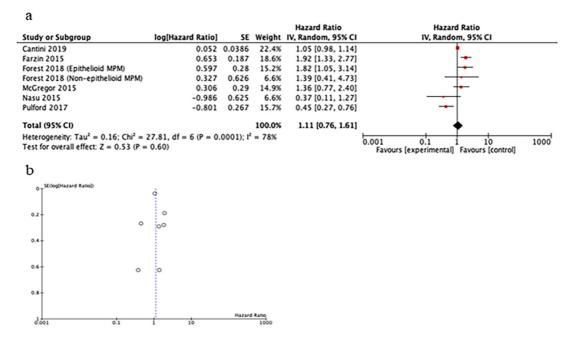


Figure 15. (a) Adjusted risk of death for MPM patients with retained BAP1 expression. (b) Funnel plot for the assessment of potential publication bias among included studies.

4.4. Prognostic value of BAP1 and miR-31 in epithelioid MPM patients

Considering the significant prognostic impact of epithelioid histotype on OS, as reported both in our cohort and from meta-analysis, we analyzed the role of BAP1 status within the e-MPM subgroup (n=40). In this specific subtype, BAP1 was not able to significantly predict OS (median OS of 19.8 months, 95% CI: 6.3-33.3 for BAP1 loss vs 18.4 months, 95% CI: 15.6-21.2 for BAP1 retained, p=0.271) and PFS (median PFS 6.3 months, 95% CI: 0.4-12.2 for BAP1 loss vs 8.1 months, 95% CI: 5.0-11.2 for BAP1 retained, p=0.453), (**Figure 16**).

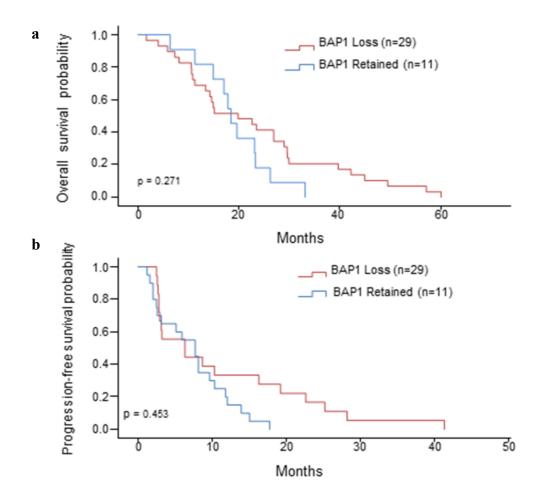


Figure 16. Kaplan-Meier survival curves for MPM epithelioid subtype subdivided by BAP1 expression. (a) Overall survival. (b) Progression-free survival. Comparisons between groups were made using log-rank test and two-sided p values lower than 0.05 were considered statistically significant.

In order to improve the performance of BAP1, we combined BAP1 status with the tissue expression of miR-31. We analyzed the expression level of miR-31 on the whole cohort (n=60). As shown in **Figure 17**, miR-31 was differently expressed between the different histotype of MPM as it was poorly expressed in the e-MPM and increased in the b-MPM until reaching higher levels in the s-MPM. Furthermore, low miR-31 levels were associated with the high percentage of MPM patients with BAP1 loss.

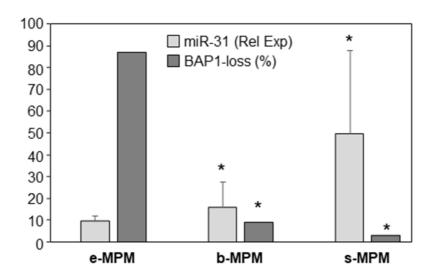


Figure 17. Distribution of miR-31 and BAP1 loss among the MPM histotypes. MiR-31 relative expression and the percentage of BAP1 loss in epithelioid MPM subtype (e-MPM), biphasic MPM subtype (b-MPM) and sarcomatoid MPM subtype (s-MPM). Comparisons among groups were determined by one-way ANOVA with Tukey post-hoc analysis. The symbol "*" indicates significant differences compared with the e-MPM with p<0.05.

We evaluated the prognostic role of miR-31 alone in e-MPM and we observed that Low miR-31 levels were significantly associated with better PFS (median PFS 7.7 months, 95% CI: 2.95-12.4 for low miR-31 levels vs 5.9 months, 95% CI: 0.0-15.5 for high miR-31 levels, p=0.028). A trend, but not significant, toward better OS was also detected (median OS 18.4 months, 95% CI: 8.3-28.5 for low miR-31 levels vs 17.0 months, 95% CI: 10.9-23.1 for high miR-31 levels, p=0.059), (**Figure 18**).

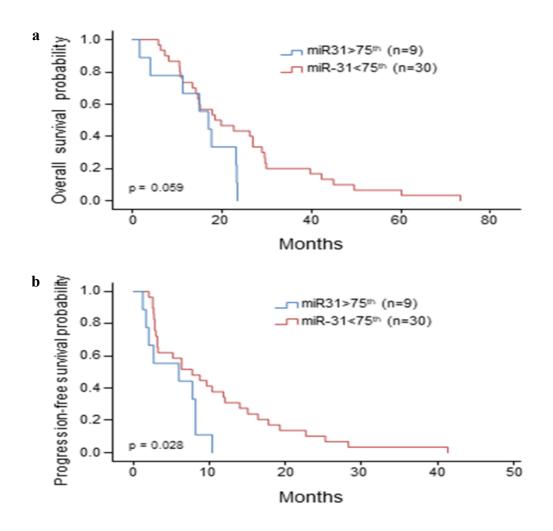
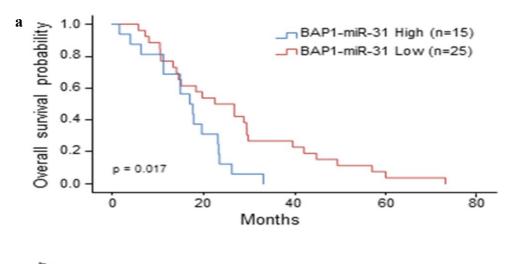


Figure 18. Kaplan-Meier survival curves for MPM epithelioid subtype subdivided by miR-31 level. (a) Overall survival. (b) Progression-free survival. Comparisons between groups were made using log-rank test and two-sided p values lower than 0.05 were considered statistically significant.

Finally, in order to further improve patient risk stratification, we stratified e-MPM patients according to BAP1 status and miR-31 levels taken together, obtaining a combined score (**Figure 19**).



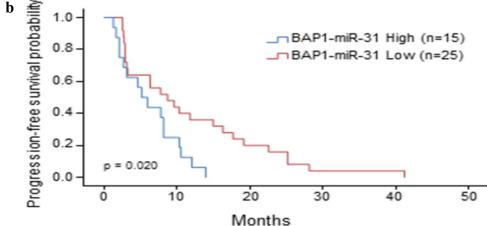


Figure 19. Kaplan-Meier survival curves for MPM epithelioid subtype subdivided by miR-31-BAP1 combination (a) Overall survival. (b) Progression-free survival. Comparisons between groups were made using log-rank test and two-sided p values lower than 0.05 were considered statistically significant.

Patients with BAP1 loss/low miR-31 levels (BAP1-miR-31 Low) had significantly better OS (median OS 22.6 months, 95% CI: 12.0-33.2 vs 17.0 months, 95% CI 11.5-22.5, p=0.017) and PFS (median PFS 8.7 months, 95% CI: 3.3-14.1 vs 5.1, 95% CI: 2.5-7.6, p=0.020), compared to the BAP1 retained/high miR-31 (BAP1-miR-31 High) subgroup. The multivariate analysis, showed in **Table 5**, confirmed BAP1 status/miR-31 level combination as an independent prognostic factor for e-MPM patients (HR of the BAP1 retained and high miR-31 level group 2.207, 95% CI: 1.062-4.587, p=0.034 for OS and HR 2.146, 95% CI: 1.050-4.388, p=0.036 for PFS).

B) MPM epithelioid (n=40)								
os			PFS					
Variable	HR	95% CI (OR)	p-value	Variable	HR	95% CI (OR)	P value	
Age	1.017	0.949-1.089	0.641	Age	0.999	0.931-1.073	0.980	
Gender	0.731	0.198-2.695	0.638	Gender	1.377	0.319-5.954	0.668	
Smoking	0.605	0.246-1.483	0.272	Smoking	0.753	0.311-1.823	0.529	
Asb-exp	2.082	1.082-4.004	0.028	Asb-exp	1.513	0.745-3.072	0.252	
Surgery	0.420	0.173-1.021	0.056	Surgery	0.424	0.156-1.152	0.093	
miR-31-BAP1	2.207	1.062-4.587	0.034	miR-31-BAP1	2.146	1.050-4.388	0.036	

Table 5. Multivariate Cox regression analysis associated with OS and PFS. Regression model with stepwise Wald-backward adjusted for age, gender, smoking, surgery, histotype, and miR-31-BAP1. OS, Overall Survival; PFS, Progression-Free Survival; HR, hazard ratio; CI, confidence interval.

5. DISCUSSION

MPM is an aggressive tumor, mostly unresectable, that responds poorly to current platinum-based chemotherapy. In daily clinical practice, heterogeneity of treatment response, also within the same MPM histotype, remains a hard challenge for clinicians. Therefore, identification of novel prognostic biomarkers may be helpful for patient risk stratification (215). BAP1 is a deubiquinating enzyme with tumor suppressor and apoptotic activity. BAP1 gene alterations, both inherited and somatic, have been detected in about 60% of MPM and, in the last 10 years, its prognostic role has been investigated. At today, this argument is still under debate and the prognostic role of BAP1 remains unclear. The past analyses were conducted in heterogeneous cohorts of patients, in which histology, treatment and other patient characteristics were not deeply analyzed (216-218). Furthermore, germline mutations could have a different prognostic significance than somatic ones, as suggested by Hassan et al. (92), but the analysis of BAP1 through IHC does not allow distinguishing between them.

In our study, conducted on MPM patients who homogenously received first-line chemotherapy, we revealed that there no differences in terms of OS according to BAP1 expression assessed by IHC (BAP1 retained vs BAP1 loss). We also showed that, at multivariate analysis, histological subtype mainly affected the patient survival and we confirmed that BAP1 loss is more frequent in epithelioid histotype, as previously observed (85, 90, 92, 180, 209, 211, 212). We only found a link between BAP1 loss and better outcome in patients who underwent surgery, but, after adjusting for histology at multivariate analysis, BAP1 status lost its prognostic value.

In order to definitely clarify the prognostic role of BAP1, we pooled our data with past studies in a comprehensive meta-analysis of 690 MPM patients. From this analysis, we demonstrated that, when tumor histology is taken into account, BAP1 immunohistochemical expression does not represent an independent prognostic factor. Several reasons might explain the discrepancy of our data with previously meta-analyses that had investigated the relationship between BAP1 expression and prognosis (216-218). In the meta-analysis conduct by Luchini et al (216), only two studies were included, and only one reported the adjusted HR for all-cause mortality (196). This study was also included in our meta-analysis (HR for BAP1 loss versus BAP1 retained 0.52, 95 % CI: 0.36–0.75).

Wang et al (217), from their meta-analysis, associated worse prognosis with BAP1 retained tumor (HR for BAP1 retained versus BAP1 loss 2.03, 95 % CI: 1.67–2.47). However, not all of the seven studies included in the analysis have conducted a multivariate analysis and some potential confounding factors (such as the tumor histotype) might have influenced the data.

The last meta-analysis presents data in agreement with ours but in this case, there was a focus on BAP1 gene mutation instead of protein expression (218).

We demonstrated that BAP1 loss was strongly associated with epithelioid histotype both in our cohort (p=0.01) and in meta-analysis (p for difference < 0.0001). E-MPM is strongly associated with better prognosis than non-e-MPM tumors.

This association between BAP1 loss and epithelioid histotype might have affected the previous meta-analysis, where the prognostic value of BAP1 is instead due to the e-MPM subtype.

By looking only at epithelioid cases in our cohort, no association between BAP1 expression and survival was observed. Despite BAP1 loss has been associated with mutations or chromosomal deletions, BAP1 loss assessed by IHC has been found also in tumors without BAP1 genetic alterations (158).

In this context, a post-transcriptional regulation mediated by miRNAs might be involved. MiR-31 showed altered levels of expression in different tumors and it has been investigated as a potential upstream regulator of BAP1 (204, 219-221). In MPM the role of miR-31 is still under investigation and data are conflicting. Ivanov et al. (222) reported that miR-31 loss in MPM cell lines was associated with cell cycle progression and its restoration inhibited cell proliferation and migration. Conversely, miR-31 over-expression in miR-31-null NCI-H2452 cells significantly increased resistance to cisplatin and carboplatin (223). In the present study, we found that miR-31 was differentially expressed accordingly to the histotype: miR-31 was highly expressed in s-MPM compared to e-MPM. High miR-31 levels were also associated with BAP1 retained, worse PFS and a trend toward a shorter OS. In the cancerous tissue the direct relationship between miR-31 levels and BAP1 expression (high miR-31 corresponded to low BAP1 expression) previously described was lost (204, 205). Most probably, the genetic and epigenetic alterations in the malignant tissues contributed to a genetic rearrangement, thus affecting gene expression.

By focusing on the e-MPM subgroup, high miR-31 levels correlated with worse PFS, and a trend to worse OS was also detected. Our results are consistent with data described by Matsumoto et al. showing a miR-31 upregulation in patients affected by s-MPM, which correlated with worse prognosis (224). Notably, the BAP1-miR-31 combination was strongly associated with OS and PFS, which was further confirmed in the multivariate

model. Accordingly, retained BAP1 and high miR-31 expression was associated with a non-epithelioid and a more aggressive phenotype.

The MPM is a highly heterogeneous cancer characterized by multiple molecular profiles. Molecular diversity has been shown to occur between different histotypes, as well as within specific histological subtypes. Therefore, we can postulate that the combination of BAP1 status with miR-31 levels may help to detect within the e-MPM an aggressive subtype with BAP1 retained and high miR-31 associated with a worse outcome.

Recently the EURACAN/IASLC (pathology) group published a study on "Updating the Histologic Classification of Pleural Mesothelioma", underlining the importance of a multidisciplinary approach based on the integration of both histological and molecular parameters (225). More detailed diagnosis may lead to improved patients risk stratification, which is essential for guiding treatment. Alongside, a better knowledge of miRNAs role among different MPM histotypes may lead to a better understanding of the complex MPM biology, as well as to the development of new miRNA-based targeted therapies. Despite being limited by the small sample size and the retrospective nature, we can postulate from our study that the combination of BAP1 status and miR-31 levels is helpful in the context of e-MPM subtype for identifying a tumor subgroup (BAP1 retained-miR-31 high) with a worse outcome. Prospective studies are needed to better analyze the role of this combined score in predicting outcome in MPM and explore the emerging idea of a molecular model classification complementary to the histological one, where miRNAs might play a key role.

6. CONCLUSION

In this study we demonstrated that BAP1 alone was unable to stratify MPM patients based on its expression when histotype was considered. We also confirmed that the epithelioid histotype was associated with better survival in terms of OS and PFS and that BAP1 loss was more represented in this histotype. However, we observed that within the epithelioid histotype, the prognostic value of BAP1 improved, but did not reach statistical significance. Therefore, in order to improve its prognostic performance, the BAP1 status was combined with miR-31 expression levels. We showed that the combination of BAP1 retained and miR-31-high was significantly associated with a worse prognosis in MPM patients.

In conclusion, BAP1 and miR-31 can be routinely detected in diagnostic biopsies and can help to identify epithelioid MPMs with worse prognosis. The combinations of IHC method (BAP1) and molecular assay (miR-31) allow to detection prognostic/predictive factor in cancer.

6. REFERENCES

- 1. Robinson BW, Creaney j, Lake R, Nowak A, Musk AW, de Klerk N, Winzell P, Hellstrom KE, Hellstrom I. Soluble mesothelin-related protein a blood test for mesothelioma. Lung Cancer. 2005;49(Suppl 1):S109-111.
- 2. Marinaccio A, Binazzi A, Di Marzio D, Scarselli A, Verardo M, Mirabelli D, Gennaro V, Mensi C, Merler E, De Zotti R, Mangone L, Chellini E, Pascucci C, Ascoli V, Menegozzo S, Cavone D, Cauzillo G, Nicita C, Melis M, Iavicoli S. Incidence of extrapleural malignant mesothelioma and asbestos exposure, from the Italian national register. Occup Environ Med. 2010;67(11):760-765
- 3. Bridda A, Padoan I, Mencarelli R, Frego M. Peritoneal mesothelioma: a review. MedGenMed. 2007;9(2):32
- 4. Selikoff IJ, Hammond EC, Seidman H. Latency of asbestos disease among insulation workers in the United States and Canada. Cancer. 1980;46(12):2736-2740.
- 5. Kumar N, Alrifai D, Kolluri KK, Sage EK, Ishii Y, Guppy N, Borg E, Falzon M, Nankivell M, Nicholson A, Janes AS. Retrospective response analysis of BAP1 expression to predict the clinical activity of systemic cytotoxic chemotherapy in mesothelioma. Lung Cancer. 2019;127:164–166.
- 6. Grondin SC, Sugarbaker DJ. Malignant mesothelioma of the pleural space. Oncology. 13;(1999);919–926
- 7. Doll R. Mortality from lung cancer in asbestos workers. Br. J. Ind. Med. 1955;12:81-86.
- 8. Wagner JC, Sleggs CA, Marchand P. Diffuse pleural mesothelioma and asbestos exposure in the North Western Cape Province. Br. J. Ind. Med. 1960;17:260–271.

- 9. Baumann F, Ambrosi JP, Carbone M. Asbestos is not just asbestos: an unrecognized health hazard. Lancet Oncol. 2013;14:576-578.
- 10. Carbone M, Kanodia S, Chao A, Miller A, Wali A, Weissman D, Adjei A, Baumann F, Boffetta P, Buck B, de Perrot M, Dogan AU, Gavett S, Gualtieri A, Hassan R, Hesdorffer M, Hirsch FR, Larson D, Mao W, Masten S, Pass HI, Peto J, Pira E, Steele I, Tsao A, Woodard GA, Yang H, Malik S. Consensus Report of the 2015 Weinman International Conference on Mesothelioma. J Thorac Oncol. 2016;11(8):1246-1262.
- 11. Ismail-Khan R, Robinson LA, Williams CC Jr, Garrett CR, Bepler G, Simon GR. Malignant pleural mesothelioma: a comprehensive review. Cancer Control. 2006;13:255–263.
- 12. Berry G, Reid A, Aboagye-Sarfo P, de Klerk NH, Olsen NJ, Merler E, Franklin P, Musk AW. Malignant mesotheliomas in former miners and millers of crocidolite at Wittenoom (Western Australia) after more than 50 years follow-up. Br J Cancer. 2012;106(5):1016-20
- 13. Ministero della Salute. Stato dell'arte e prospettive in materia di contrasto alle patologie asbesto-correlate. Quad. Del Minist. Della Salut. 2012;15:2038–5293. Available online:
- http://www.salute.gov.it/imgs/C_17_pubblicazioni_2570_allegato.pdf (accessed on 11 June 2019).
- 14. Kazan-Allen L. Asbestos and mesothelioma: Worldwide trends. Lung Cancer. 2005;49(Suppl. 1): 3–8.
- 15. World Health Organization. Elimination of Asbestos-related Diseases. 2006. Available online: http://whqlibdoc.who.int/hq/2006/WHO_SDE_OEH_06.03_eng.pdf.

- 16. World Health Organization. Chrysotile Asbestos. 2014. Available online: https://www.who.int/ipcs/assessment/public health/chrysotile asbestos summary.pdf.
- 17. Boffetta P. Malignant Mesothelioma: Epidemiology. In Occupational Cancers. Anttila S, Boffetta P, Eds. Springer Verlag: London, UK, 2014; pp. 253–264. Springer Verlag: London, UK, 2014; pp. 253–264.
- 18. Kameda T, Takahashi K, Kim R, Jiang Y, Movahed M, Park EK, Rantanen J. Asbestos: Use, bans and disease burden in Europe. Bull. World Health Organ. 2014;92:790–797.
- 19. Baumann F, Carbone M. Environmental risk of mesothelioma in the United States: an emerging concern-epidemiological issues. J Toxicol Environ Health B Crit Rev. 2016;19(5-6):231-249.
- 20. World Health Organization. Fact Sheet n° 343; World Health Organization: Geneva, Switzerland, July 2010.
- 21. Delgermaa V, Takahashi K, Park EK, Le GV, Hara T, Sorahan T. Global mesothelioma deaths reported to the World Health Organization between 1994 and 2008. Bull. World Health Org. 2011;89:716–724.
- 22. WHO. Download the Raw Data Files of the WHO Mortality Database. Available online: http://www.who.int/healthinfo/statistics/mortality_rawdata/en/.
- 23. Robinson BW, Musk AW, Lake RA. Malignant mesothelioma. Lancet. 2005;366:397–408.
- 24. Hillerdal G. Mesothelioma: Cases associated with non-occupational and low dose exposures. Occup. Environ. Med. 1999;56:505–513.

- 25. Park EK, Takahashi K, Hoshuyama T, Cheng TJ, Delgermaa V, Le GV, Sorahan T. Global magnitude of reported and unreported mesothelioma. Environ. Health Perspect. 2011;119:514–518. [CrossRef] [PubMed
- 26. Prüss-Ustün A, Vickers C, Haefliger P, Bertollini R. Knowns and unknowns on burden of disease due to chemicals: A systematic review. Environ. Health 2011;10:9.
- 27. Sun H. North-south gradient of mesothelioma and asbestos consumptionproduction in the United States-Progresses since the 1st asbestos partial ban in 1973. Am J Ind Med. 2019;62:337-346.
- 28. Lemen R.A. Mesothelioma from asbestos exposures: Epidemiologic patterns and impact in the United States. J. Toxicol. Environ. Health B Crit. Rev. 2016;19:250–265.
- 29. Taioli E, Wolf AS, Camacho-Rivera M, Flores RM. Women with malignant pleural mesothelioma have a threefold better survival rate than men. Ann Thorac Surg. 2014;98:1020-1024.
- 30. Vivero M, Bueno R, Chirieac LR. Clinicopathologic and genetic characteristics of young patients with pleural diffuse malignant mesothelioma. Mod Pathol. 2018;31:122-131.
- 31. Ferlay J, Ervik M, Lam F. Global Cancer Observatory: Cancer Today. Lyon, France: International Agency for Research on Cancer; 2018.
- 32. Geltner C, Errhalt P, Baumgartner B, Ambrosch G, Machan B, Eckmayr J, Klikovits T, Hoda MA, Popper H, Klepetko W; Austrian Mesothelioma Interest Group (AMIG). Management of malignant pleuralmesothelioma—part 1: Epidemiology, diagnosis, and staging Consensus of the Austrian Mesothelioma Interest Group (AMIG). Wien. Klin Wochenschr. 2016;128:611-617.

- 33. Järvholm B, Burdorf A. Emerging evidence that the ban on asbestos use is reducing the occurrence of pleural mesothelioma in Sweden. Scand. J. Public Health. 2015;43:875–881.
- 34. Peto, J, Decarli, A, La Vecchia C, Levi, F, Negri E. The European mesothelioma epidemic. Br. J. Cancer 1999;79:666–672.
- 35. Bianchi C, Bianchi T. Global mesothelioma epidemic: Trends and features. Indian J. Occup. Environ. Med. 2014;18:82–88.
- 36. Andujar P, Lacourt A, Brochard P, Pairon JC, Jaurand MC, Jean D. Five years update on relationships between malignant pleural mesothelioma and exposure to asbestos and other elongated mineral particles. J. Toxicol. Environ. Health B Crit. Rev. 2016;19:151–172.
- 37. Liu B, van Gerwen M, Bonassi S, Taioli E. Epidemiology of environmental exposure and malignant mesothelioma. J Thorac Oncol. 2017;12:1031-1045.
- 38. RARECARENET. Available online: http://www.rarecare.eu/
- 39. Allen LP, Baez J, Stern MEC, Takahashi K, George F. Trends and the economic effect of asbestos bans and decline in asbestos consumption and production worldwide. Int J Environ Res Public Health. 2018;15:pii: E531.
- 40. Vigliani EC. A glance at the early Italian studies on the health effects of asbestos. Med Lav. 1991;82(6):489-91.
- 41. Carnevale F, Chellini E. The diffusion of information on the carcinogenicity of asbestos in the Italian scientific community before 1965. [Italian]. Med Lav. 1995;86(4):295-302.

- 42. Scansetti G. L'amianto ieri e oggi. In: Minoia C, Scansetti G, Piolatto G, Massola A. (Eds). L'amianto: dall'ambiente di lavoro all'ambiente di vita. Nuovi indicatori per future effetti. Pavia: Fondazione Salvatore Maugeri, IRCCS; 1997. p. 9-24.
- 43. Donelli G, Marsili D, Comba P. Le problematiche scientifico-sanitarie correlate all'amianto: l'attività dell'Istituto Superiore di Sanità negli anni 1980-2012. Roma: Istituto Superiore di Sanità, 2012. (I beni storico-scientifici dell'istituto Superiore di Sanità, Quaderno 9). Available from: www.iss.it/binary/pres/cont/libro_amianto.pdf.
 44. Corfiati M, Scarselli A, Binazzi A, Di Marzio D, Verardo M, Mirabelli D, Gennaro V, Mensi C, Schallemberg G, Merler E, Negro C, Romanelli A, Chellini E, Silvestri S, Cocchioni M, Pascucci C, Stracci F, Romeo E, Trafficante L, Angelillo I, Menegozzo S, Musti M, Cavone D, Cauzillo G, Tallarigo F, Tumino R, Melis M, Iavicoli S, Marinaccio A; ReNaM Working Group. Epidemiological patterns of asbestos exposure and spatial clusters of incident cases of malignant mesothelioma from the Italian national registry.
- 45. Pasetto R, Fazzo L, Zona A, Bruno C, Pirastu R, Binazzi A, Corfiati M, Silvestri S, Comba P, Marinaccio A. SENTIERI-ReNaM: valutazione globale del carico di mesotelioma. Epidemiol Prev. 2016;40 (5 Suppl 1):99-104.

BMC Cancer. 2015;15:286.

- 46. Ferrante D, Mirabelli D, Tunesi S, Terracini B, Magnani C. Pleural mesothelioma and occupational and nonoccupational asbestos exposure: a case-control study with quantitative risk assessment. Occup Environ Med. 2016;73(3):147-53.
- 47. Decreto del Presidente del Consiglio dei Ministri, n. 308 del 10 Dicembre 2002. Regolamento per la Determinazione del Modello e Delle Modalità di Tenuta del Registro dei casi di Mesotelioma Asbesto Correlati ai Sensi Dell'art. 36 del Decreto Legislativo n.

- 277 del 1991; Gazzetta Uffciale n.31 del 07.02.2003; Decreto del Presidente del Consiglio dei Ministri: Roma, Italy, 2003.
- 48. INAIL (National Workers Compensation Authority). Il registro Nazionale dei Mesoteliomi V Rapporto. Available online: https://www.inail.it/cs/internet/comunicazione/pubblicazioni/catalogogenerale/ilregistro -nazionale-dei-mesoteliomi-v-rapporto.html.
- 49. Marsili D, Terracini B, Santana VS, Ramos-Bonilla JP, Pasetto R, Mazzeo A, Loomis D, Comba P, Algranti E. Prevention of Asbestos-Related Disease in Countries Currently Using Asbestos. Int. J. Environ. Res. Public Health 2016;13:494.
- 50. Martuzzi M, Pasetto R, Martin-Olmedo P. (Eds.) Industrially contaminated sites and health. J. Environ. Public Health 2014. Available online: http://www.hindawi.com/journals/jeph/si/480565/.
- 51. Vimercati L, Cavone D, Lovreglio P, De Maria L, Caputi A, Ferri GM, Serio G. Environmental asbestos exposure and mesothelioma cases in Bari, Apulia region, southern Italy: A national interest site for land reclamation. Environ. Sci. Pollut. Res. Int. 2018;25:15692–15701.
- 52. Marinaccio A, Binazzi A, Bonafede M, Di Marzio D, Scarselli A; Regional Operating Centres. Epidemiology of malignant mesothelioma in Italy: surveillance systems, territorial clusters and occupations involved. J Thorac Dis. 2018;10(Suppl 2):S221-S227. 53. Carbone M, Kratzke R, Testa JR. The pathogenesis of mesothelioma. Semin Oncol. 29:2-17, 2002.
- 54. IARC. Arsenic, metals, fibres, and dust. IARC Monogr Eval Carcinog Risks Hum. 2012;100(Pt C):11-465.

- 55. Saffiotti U. Mesothelioma carcinogenesis: in vivo models, in Pass HI, Vogelzang NJ, Carbone M (eds): Malignant Mesothelioma. New York, NY, Springer, 2005:60-86.
- 56. Magnani C, Agudo A, González CA, Andrion A, Calleja A, Chellini E, Dalmasso P, Escolar A, Hernandez S, Ivaldi C, Mirabelli D, Ramirez J, Turuguet D, Usel M, Terracini B. Multicentric study on malignant pleural mesothelioma and non-occupational exposure to asbestos. Br J Cancer. 2000;83(1):104-11
- 57. Marsh GM, Riordan AS, Keeton KA, Benson SM. Non-occupational exposure to asbestos and risk of pleural mesothelioma: review and meta-analysis. Occup Environ Med. 2017;74(11):838-846.
- 58. Carbone M, Adusumilli PS, Alexander HR Jr, Baas P, Bardelli F, Bononi A, Bueno R, Felley-Bosco E, Galateau-Salle F, Jablons D, Mansfield AS, Minaai M, de Perrot M, Pesavento P, Rusch V, Severson DT, Taioli E, Tsao A, Woodard G, Yang H, Zauderer MG, Pass HI. Mesothelioma: Scientific clues for prevention, diagnosis, and therapy. CA Cancer J Clin. 2019;69(5):402-429.
- 59. Robledo R, Mossman B. Cellular and molecular mechanisms of asbestos-induced fibrosis. J. Cell. Physiol. 1999;180:158–166.
- 60. Chew SH, Toyokuni S. Malignant mesothelioma as an oxidative stress-induced cancer: An update. Free Radic. Biol. Med. 2015;86:166–178.
- 61. Kamp DW, Weitzman SA. The molecular basis of asbestos induced lung injury. Thorax 1999;54:638–652.
- 62. Heintz NH, Janssen-Heininger YM, Mossman BT. Asbestos, lung cancers, and mesotheliomas: from molecular approaches to targeting tumor survival pathways. Am J Respir Cell Mol Biol. 2010;42(2):133-139.

- 63. Toyokuni S. Mechanisms of asbestos-induced carcinogenesis. Nagoya J Med Sci. 2009;71(1-2):1-10.
- 64. Tubbs A, Nussenzweig A. Endogenous DNA damage as a source of genomic instability in cancer. Cell. 2017;168:644–656.
- 65. Liu W, Ernst JD, Broaddus VC. Phagocytosis of crocidolite asbestos induces oxidative stress, DNA damage, and apoptosis in mesothelial cells. Am J Respir Cell Mol Biol. 2000;23(3):371-378.
- 66. Yang H, Bocchetta M, Kroczynska B, Elmishad AG, Chen Y, Liu Z, Bubici C, Mossman BT, Pass HI, Testa JR, Franzoso G, Carbone M. TNF-α inhibits asbestosinduced cytotoxicity via a NF-kB dependent pathway, a possible mechanism for asbestos-induced oncogenesis. Proc Natl Acad Sci. 2006;103:10397-10402.
- 67. Filomeni G, De Zio D, Cecconi F. Oxidative stress and autophagy: the clash between damage and metabolic needs. Cell Death Differ. 2015;22(3):377-388.
- 68. Dikic I, Elazar Z. Mechanism and medical implications of mammalian autophagy. Nat Rev Mol Cell Biol. 2018;19(6):349-364.
- 69. Levy JMM, Towers CG, Thorburn A. Targeting autophagy in cancer. Nat Rev Cancer. 2017;17(9):528-542.
- 70. Yang H, Rivera Z, Jube S, Nasu M, Bertino P, Goparaju C, Franzoso G, Lotze MT, Krausz T, Pass HI, Bianchi ME, Carbone M. Programmed necrosis induced by asbestos in human mesothelial cells causes high-mobility group box 1 protein release and resultant inflammation. Proc Natl Acad Sci U S A. 2010;107(28):12611-6.
- 71. Klune JR, Dhupar R, Cardinal J, Billiar TR, Tsung A. HMGB1: endogenous danger signaling. Mol Med. 2008;14(7-8):476-84.

- 72. Scaffidi P, Misteli T, Bianchi ME. Release of chromatin protein HMGB1 by necrotic cells triggers inflammation. Nature. 2002;418(6894):191-5.
- 73. Bianchi ME, Crippa MP, Manfredi AA, Mezzapelle R, Rovere Querini P, Venereau E. High-mobility group box 1 protein orchestrates responses to tissue damage via inflammation, innate and adaptive immunity, and tissue repair. Immunol Rev. 2017;280(1):74-82.
- 74. Carbone M, Yang H. Molecular pathways: targeting mechanisms of asbestos and erionite carcinogenesis in mesothelioma. Clin Cancer Res. 2012;18(3):598-604.
- 75. Kang R, Livesey KM, Zeh HJ, Loze MT, Tang D. HMGB1: a novel Beclin 1-binding protein active in autophagy. Autophagy. 2010;6(8):1209-1211.
- 76. Kang R, Livesey KM, Zeh HJ 3rd, Lotze MT, Tang D. Metabolic regulation by HMGB1-mediated autophagy and mitophagy. Autophagy. 2011;7(10):1256-8.
- 77. Tang D, Kang R, Livesey KM, Cheh CW, Farkas A, Loughran P, Hoppe G, Bianchi ME, Tracey KJ, Zeh HJ 3rd, Lotze MT. Endogenous HMGB1 regulates autophagy. J Cell Biol. 2010;190(5):881-892.
- 78. Xue J, Patergnani S, Giorgi C, Suarez J, Goto K, Bononi A, Tanji M, Novelli F, Pastorino S, Xu R, Caroccia N, Dogan AU, Pass HI, Tognon M, Pinton P, Gaudino G, Mak TW, Carbone M, Yang H. Asbestos induces mesothelial cell transformation via HMGB1-driven autophagy. Proc Natl Acad Sci U S A. 2020;117(41):25543-25552.
- 79. Liu Z, Klominek J. Chemotaxis and chemokinesis of malignant mesothelioma cells to multiple growth factors. Anticancer Res. 2004;24:1625–1630.
- 80. Galffy G, Mohammed KA, Dowling PA, et al. Interleukin 8: an autocrine growth factor for malignant mesothelioma. Cancer Res. 1999;59:367–371.

- 81. Strizzi L, Catalano A, Vianale G, et al. Vascular endothelial growth factor is an autocrine growth factor in human malignant mesothelioma. J Pathol 2001;193:468–475.
- 82. Cacciotti P, Libener R, Betta P, et al. SV40 replication in human mesothelial cells induces HGF/Met receptor activation: a model for viral-related carcinogenesis of human malignant mesothelioma. Proc Natl Acad Sci U S A 2001;98:12032–12037.
- 83. Comertpay S, Pastorino S, Tanji M, Mezzapelle R, Strianese O, Napolitano A, Baumann F, Weigel T, Friedberg J, Sugarbaker P, Krausz T, Wang E, Powers A, Gaudino G, Kanodia S, Pass HI, Parsons BL, Yang H, Carbone M. Evaluation of clonal origin of malignant mesothelioma. J Transl Med. 2014
- 84. Oey H, Daniels M, Relan V, Chee TM, Davidson MR, Yang IA, Ellis JJ, Fong KM, Krause L, Bowman RV. Whole-genome sequencing of human malignant mesothelioma tumours and cell lines. Carcinogenesis. 2019;40(6):724-734;12:301.
- 85. Bueno R, Stawiski EW, Goldstein LD, Durinck S, De Rienzo A, Modrusan Z, Gnad F, Nguyen TT, Jaiswal BS, Chirieac LR, Sciaranghella D, Dao N, Gustafson CE, Munir KJ, Hackney JA, Chaudhuri A, Gupta R, Guillory J, Toy K, Ha C, Chen YJ, Stinson J, Chaudhuri S, Zhang N, Wu TD, Sugarbaker DJ, de Sauvage FJ, Richards WG, Seshagiri S. Comprehensive genomic analysis of malignant pleural mesothelioma identifies recurrent mutations, gene fusions and splicing alterations. Nat Genet. 2016;48(4):407-16. 86. Hmeljak J, Sanchez-Vega F, Hoadley KA, Shih J, Stewart C, Heiman D, Tarpey P, Danilova L, Drill E, Gibb EA, Bowlby R, Kanchi R, Osmanbeyoglu HU, Sekido Y, Takeshita J, Newton Y, Graim K, Gupta M, Gay CM, Diao L, Gibbs DL, Thorsson V, Iype L, Kantheti H, Severson DT, Ravegnini G, Desmeules P, Jungbluth AA, Travis WD, Dacic S, Chirieac LR, Galateau-Sallé F, Fujimoto J, Husain AN, Silveira HC, Rusch VW, Rintoul RC, Pass H, Kindler H, Zauderer MG, Kwiatkowski DJ, Bueno R, Tsao AS,

- Creaney J, Lichtenberg T, Leraas K, Bowen J; TCGA Research Network, Felau I, Zenklusen JC, Akbani R, Cherniack AD, Byers LA, Noble MS, Fletcher JA, Robertson AG, Shen R, Aburatani H, Robinson BW, Campbell P, Ladanyi M. Integrative Molecular Characterization of Malignant Pleural Mesothelioma. Cancer Discov. 2018;8(12):1548-1565.
- 87. Nasu M, Emi M, Pastorino S, Tanji M, Powers A, Luk H, Baumann F, Zhang YA, Gazdar A, Kanodia S, Tiirikainen M, Flores E, Gaudino G, Becich MJ, Pass HI, Yang H, Carbone M. High Incidence of Somatic BAP1 alterations in sporadic malignant mesothelioma. J Thorac Oncol. 2015;10(4):565-76.
- 88. Emri SA. The Cappadocia mesothelioma epidemic: its influence in Turkey and abroad. *Ann Transl Med.* 2017;5:239.
- 89. Carbone M, Amelio I, Affar EB, Brugarolas J, Cannon-Albright LA, Cantley LC, Cavenee WK, Chen Z, Croce CM, Andrea A, Gandara D, Giorgi C, Jia W, Lan Q, Mak TW, Manley JL, Mikoshiba K, Onuchic JN, Pass HI, Pinton P, Prives C, Rothman N, Sebti SM, Turkson J, Wu X, Yang H, Yu H, Melino G. Consensus report of the 8 and 9th Weinman Symposia on Gene x Environment Interaction in carcinogenesis: novel opportunities for precision medicine. Cell Death Differ. 2018;25(11):1885-1904.
- 90. Pastorino S, Yoshikawa Y, Pass HI, Emi M, Nasu M, Pagano I, Takinishi Y, Yamamoto R, Minaai M, Hashimoto-Tamaoki T, Ohmuraya M, Goto K, Goparaju C, Sarin KY, Tanji M, Bononi A, Napolitano A, Gaudino G, Hesdorffer M, Yang H, Carbone M. A Subset of Mesotheliomas With Improved Survival Occurring in Carriers of BAP1 and Other Germline Mutations. J Clin Oncol. 2018;36(35):JCO2018790352.
- 91. Panou V, Gadiraju M, Wolin A, Weipert CM, Skarda E, Husain AN, Patel JD, Rose B, Zhang SR, Weatherly M, Nelakuditi V, Knight Johnson A, Helgeson M, Fischer D,

- Desai A, Sulai N, Ritterhouse L, Røe OD, Turaga KK, Huo D, Segal J, Kadri S, Li Z, Kindler HL, Churpek JE. Frequency of Germline Mutations in Cancer Susceptibility Genes in Malignant Mesothelioma. J Clin Oncol. 2018;36(28):2863-2871.
- 92. Hassan R, Morrow B, Thomas A, Walsh T, Lee MK, Gulsuner S, Gadiraju M, Panou V, Gao S, Mian I, Khan J, Raffeld M, Patel S, Xi L, Wei JS, Hesdorffer M, Zhang J, Calzone K, Desai A, Padiernos E, Alewine C, Schrump DS, Steinberg SM, Kindler HL, King MC, Churpek JE. Inherited predisposition to malignant mesothelioma and overall survival following platinum chemotherapy. Proc Natl Acad Sci U S A. 2019;116(18):9008-9013.
- 93. Kim KC, Vo HP. Localized malignant pleural sarcomatoid mesothelioma misdiagnosed as benign localized fibrous tumor. J Thorac Dis 2016;8:E379-84.
- 94. Perrotta F, Cerqua FS, Cammarata A, Izzo A, Bergaminelli C, Curcio C, Guarino C, Grella E, Forzano I, Cennamo A, Tafuri D, Rocca A, Bianco A, Mazzarella G. Integrated therapeutic approach to giant solitary fibrous tumor of the pleura: report of a case and review of the literature. Open Med (Wars). 2016;11(1):220-225...
- 95. Scherpereel A, Astoul P, Baas P, Berghmans T, Clayson H, de Vuyst P, Dienemann H, Galateau-Salle F, Hennequin C, Hillerdal G, Le Péchoux C, Mutti L, Pairon JC, Stahel R, van Houtte P, van Meerbeeck J, Waller D, Weder W; European Respiratory Society/European Society of Thoracic Surgeons Task Force. Guidelines of the European Respiratory Society and the European Society of Thoracic Surgeons for the management of malignant pleural mesothelioma. Eur Respir J. 2010;35(3):479-95.
- 96. British Thoracic Society Standards of Care Committee. BTS statement on malignant mesothelioma in the UK, 2007. Thorax 2007;62:ii1-ii19.

- 97. Robinson BW, Lake RA. Advances in malignant mesothelioma. N Engl J Med 2005;353:1591-603.
- 98. Bibby AC, Tsim S, Kanellakis N, Ball H, Talbot DC, Blyth KG, Maskell NA, Psallidas I. Malignant pleural mesothelioma: an update on investigation, diagnosis and treatment. Eur Respir Rev. 2016;25(142):472-486.
- 99. Baas P, Fennell D, Kerr KM, Van Schil PE, Haas RL, Peters S; ESMO Guidelines Committee. Malignant pleural mesothelioma: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. Ann Oncol. 2015;26 Suppl 5:v31-9.
- 100. Labby ZE, Armato SG 3rd, Dignam JJ, Straus C, Kindler HL, Nowak AK. Lung volume measurements as a surrogate marker for patient response in malignant pleural mesothelioma. J Thorac Oncol. 2013;8(4):478-86.
- 101. Maniscalco M, Bianco A, Mazzarella G, Motta A. Recent Advances on Nitric Oxide in the Upper Airways. Curr Med Chem. 2016;23(24):2736-2745.
- 102. Travis WD, Brambilla E, Burke AP, et al. WHO classification of tumours of the lung, pleura, thymus and heart. Lyon (France): International Agency for Research on Cancer; 2015.
- 103. Saddoughi SA, Abdelsattar ZM, Blackmon SH. National trends in the epidemiology of malignant pleural mesothelioma: A National Cancer Data Base Study. Ann Thorac Surg 2018; 105:432–437.
- 104. Rusch VW, Giroux D, Kennedy C, et al. Initial analysis of the International Association for the Study of Lung Cancer mesothelioma database. J Thorac Oncol 2012;7(11):1631–9.

- 105. Meyerhoff RR, Yang CF, Speicher PJ, et al. Impact of mesothelioma histologic subtype on outcomes in the surveillance, epidemiology, and end results database. J Surg Res 2015;196(1):23–32.42.
- 106. Kadota K, Suzuki K, Sima CS, Rusch VW, Adusumilli PS, Travis WD. Pleomorphic epithelioid diffuse malignant pleural mesothelioma: a clinicopathological review and conceptual proposal to reclassify as biphasic or sarcomatoid mesothelioma. J Thorac Oncol. 2011.
- 107. Klebe S, Mahar A, Henderson DW, Roggli VL. Malignant mesothelioma with heterologous elements: clinicopathological correlation of 27 cases and literature review. Mod Pathol. 2008;21(9):1084-94.
- 108. International Union Against Cancer (UICC): TNM Atlas. Berlin: Springer; 1992.109. AJCC Manual for staging of cancer. Philadelphia: J.B. Lippincott; 1992.
- 110. Mountain CF, Dresler CM. Regional lymph node classification for lung cancer staging. Chest 1997;111:1718-23.
- 111. Okiemy G, Foucault C, Avisse C, Hidden G, Riquet M. Lymphatic drainage of the diaphragmatic pleura to the peritracheobronchial lymph nodes. Surg Radiol Anat. 2003 Apr;25(1):32-5.
- 112. Edwards JG, Stewart DJ, Martin-Ucar A, Muller S, Richards C, Waller DA. The pattern of lymph node involvement influences outcome after extrapleural pneumonectomy for malignant mesothelioma. J Thorac Cardiovasc Surg. 2006 May;131(5):981-7.
- 113. Richards WG, Godleski JJ, Yeap BY, Corson JM, Chirieac LR, Zellos L, Mujoomdar A, Jaklitsch MT, Bueno R, Sugarbaker DJ. Proposed adjustments to

- pathologic staging of epithelial malignant pleural mesothelioma based on analysis of 354 cases. Cancer. 2010 Mar 15;116(6):1510-7.
- 114. Sugarbaker DJ, Richards WG, Bueno R. Extrapleural pneumonectomy in the treatment of epithelioid malignant pleural mesothelioma: novel prognostic implications of combined N1 and N2 nodal involvement based on experience in 529 patients. Ann Surg 2014;260:577-80; discussion 580-2.
- 115. Flores RM, Routledge T, Seshan VE, Dycoco J, Zakowski M, Hirth Y, Rusch VW. The impact of lymph node station on survival in 348 patients with surgically resected malignant pleural mesothelioma: implications for revision of the American Joint Committee on Cancer staging system. J Thorac Cardiovasc Surg. 2008;136(3):605-10.
- 116. Finn RS, Brims FJH, Gandhi A, Olsen N, Musk AW, Maskell NA, Lee YCG. Postmortem findings of malignant pleural mesothelioma: a two-center study of 318 patients. Chest. 2012 Nov;142(5):1267-1273
- 117. Nauta RJ, Osteen RT, Antman KH, Koster JK. Clinical staging and the tendency of malignant pleural mesotheliomas to remain localized. Ann Thorac Surg. 1982 Jul;34(1):66-70.
- 118. Kindler HL, Ismaila N, Armato SG 3rd, Bueno R, Hesdorffer M, Jahan T, Jones CM, Miettinen M, Pass H, Rimner A, Rusch V, Sterman D, Thomas A, Hassan R. Treatment of Malignant Pleural Mesothelioma: American Society of Clinical Oncology Clinical Practice Guideline. J Clin Oncol. 2018 May 1;36(13):1343-1373.
- 119. Waller DA, Tenconi S. Surgery as part of radical treatment for malignant pleural mesothelioma. Curr Opin Pulm Med 2017; 23:334–338.
- 120. Verma V, Ahern CA, Berlind CG, Lindsay WD, Sharma S, Shabason J, Culligan MJ, Grover S, Friedberg JS, Simone CB 2nd. National Cancer Database Report on

- Pneumonectomy Versus Lung-Sparing Surgery for Malignant Pleural Mesothelioma. J Thorac Oncol. 2017 Nov;12(11):1704-1714.waller
- 121. Dasari S, Tchounwou PB. Cisplatin in cancer therapy: Molecular mechanisms of action. Eur. J. Pharmacol. **2014**;740:364–378.
- 122. Oehl K, Vrugt B, Opitz I, Meerang M. Heterogeneity in Malignant Pleural Mesothelioma. Int J Mol Sci. 2018 May 30;19(6):1603.
- 123. Vogelzang NJ, Rusthoven JJ, Symanowski J, Denham C, Kaukel E, Ruffie P, Gatzemeier U, Boyer M, Emri S, Manegold C, Niyikiza C, Paoletti P. Phase III study of pemetrexed in combination with cisplatin versus cisplatin alone in patients with malignant pleural mesothelioma. J Clin Oncol. 2003;21(14):2636-44.
- 124. Levin PA, Dowell JE. Spotlight on bevacizumab and its potential in the treatment of malignant pleural mesothelioma: the evidence to date. Onco Targets Ther 2017;10:2057-66.
- 125. Zalcman G, Mazieres J, Margery J, Greillier L, Audigier-Valette C, Moro-Sibilot D, Molinier O, Corre R, Monnet I, Gounant V, Rivière F, Janicot H, Gervais R, Locher C, Milleron B, Tran Q, Lebitasy MP, Morin F, Creveuil C, Parienti JJ, Scherpereel A; French Cooperative Thoracic Intergroup (IFCT). Bevacizumab for newly diagnosed pleural mesothelioma in the Mesothelioma Avastin Cisplatin Pemetrexed Study (MAPS): a randomised, controlled, open-label, phase 3 trial. Lancet. 2016
- 126. van den Bogaert DP, Pouw EM, van Wijhe G, Vernhout RM, Surmont VF, Hoogsteden HC, van Klaveren RJ. Pemetrexed maintenance therapy in patients with malignant pleural mesothelioma. J Thorac Oncol. 2006;1(1):25-30.;387(10026):1405-1414.
- 127. Arnold DT, Clive AO. Prophylactic radiotherapy for procedure tract metastases

in mesothelioma: a review. Curr Opin Pulm Med 2017; 23:357–364.

128. Tsao AS, Lindwasser OW, Adjei AA, Adusumilli PS, Beyers ML, Blumenthal GM, Bueno R, Burt BM, Carbone M, Dahlberg SE, de Perrot M, Fennell DA, Friedberg J, Gill RR, Gomez DR, Harpole DH Jr, Hassan R, Hesdorffer M, Hirsch FR, Hmeljak J, Kindler HL, Korn EL, Liu G, Mansfield AS, Nowak AK, Pass HI, Peikert T, Rimner A, Robinson BWS, Rosenzweig KE, Rusch VW, Salgia R, Sepesi B, Simone CB 2nd, Sridhara R, Szlosarek P, Taioli E, Tsao MS, Yang H, Zauderer MG, Malik SM. Current and Future Management of Malignant Mesothelioma: A Consensus Report from the National Cancer Institute Thoracic Malignancy Steering Committee, International Association for the Study of Lung Cancer, and Mesothelioma Applied Research Foundation. J Thorac Oncol. 2018;13(11):1655-1667.

129. Suzuki K, Kadota K, Sima CS, Sadelain M, Rusch VW, Travis WD, Adusumilli PS. Chronic inflammation in tumor stroma is an independent predictor of prolonged survival in epithelioid malignant pleural mesothelioma patients. Cancer Immunol Immunother. 2011;60(12):1721-8.

130. Ujiie H, Kadota K, Nitadori JI, Aerts JG, Woo KM, Sima CS, Travis WD, Jones DR, Krug LM, Adusumilli PS. The tumoral and stromal immune microenvironment in malignant pleural mesothelioma: A comprehensive analysis reveals prognostic immune markers. Oncoimmunology. 2015;4(6):e1009285.

131. Mutti L, Peikert T, Robinson BWS, Scherpereel A, Tsao AS, de Perrot M, Woodard GA, Jablons DM, Wiens J, Hirsch FR, Yang H, Carbone M, Thomas A, Hassan R. Scientific Advances and New Frontiers in Mesothelioma Therapeutics. J Thorac Oncol. 2018(9):1269-1283.

- 132. Alley EW, Lopez J, Santoro A, Morosky A, Saraf S, Piperdi B, van Brummelen E. Clinical safety and activity of pembrolizumab in patients with malignant pleural mesothelioma (KEYNOTE-028): preliminary results from a non-randomised, open-label, phase 1b trial. Lancet Oncol. 2017;18(5):623-630.
- 133. Cornelissen R, Hegmans JP, Maat AP, Kaijen-Lambers ME, Bezemer K, Hendriks RW, Hoogsteden HC, Aerts JG. Extended Tumor Control after Dendritic Cell Vaccination with Low-Dose Cyclophosphamide as Adjuvant Treatment in Patients with Malignant Pleural Mesothelioma. Am J Respir Crit Care Med. 2016;193(9):1023-31.
- 134. Curran D, Sahmoud T, Therasse P, van Meerbeeck J, Postmus PE, Giaccone G. Prognostic factors in patients with pleural mesothelioma: the European Organization for Research and Treatment of Cancer experience. J Clin Oncol. 1998;16(1):145-52.
- 135. Herndon JE, Green MR, Chahinian AP, Corson JM, Suzuki Y, Vogelzang NJ. Factors predictive of survival among 337 patients with mesothelioma treated between 1984 and 1994 by the Cancer and Leukemia Group B. Chest. 1998;113(3):723-31.
- 136. Hollevoet K, Reitsma JB, Creaney J, Grigoriu BD, Robinson BW, Scherpereel A, Cristaudo A, Pass HI, Nackaerts K, Rodríguez Portal JA, Schneider J, Muley T, Di Serio F, Baas P, Tomasetti M, Rai AJ, van Meerbeeck JP. Serum mesothelin for diagnosing malignant pleural mesothelioma: an individual patient data meta-analysis. J Clin Oncol. 2012;30(13):1541-9.
- 137. Hollevoet K, Nackaerts K, Gosselin R, De Wever W, Bosquée L, De Vuyst P, Germonpré P, Kellen E, Legrand C, Kishi Y, Delanghe JR, van Meerbeeck JP. Soluble mesothelin, megakaryocyte potentiating factor, and osteopontin as markers of patient response and outcome in mesothelioma. J Thorac Oncol. 2011 Nov;6(11):1930-7.

- 138. Hollevoet K, Nackaerts K, Thas O, Thimpont J, Germonpré P, De Vuyst P, Bosquée L, Legrand C, Kellen E, Kishi Y, Delanghe JR, van Meerbeeck JP. The effect of clinical covariates on the diagnostic and prognostic value of soluble mesothelin and megakaryocyte potentiating factor. Chest. 2012;141(2):477-484.
- 139. Grigoriu BD, Scherpereel A, Devos P, Chahine B, Letourneux M, Lebailly P, Grégoire M, Porte H, Copin MC, Lassalle P. Utility of osteopontin and serum mesothelin in malignant pleural mesothelioma diagnosis and prognosis assessment. Clin Cancer Res. 2007;13(10):2928-35.
- 140. Cristaudo A, Foddis R, Vivaldi A, Guglielmi G, Dipalma N, Filiberti R, Neri M, Ceppi M, Paganuzzi M, Ivaldi GP, Mencoboni M, Canessa PA, Ambrosino N, Chella A, Mutti L, Puntoni R. Clinical significance of serum mesothelin in patients with mesothelioma and lung cancer. Clin Cancer Res. 2007;13(17):5076-81.
- 141. Wai PY, Kuo PC. The role of Osteopontin in tumor metastasis. J Surg Res 2004;121:228-41.
- 142. Rai AJ, Flores RM, Mathew A, Gonzalez-Espinoza R, Bott M, Ladanyi M, Rusch V, Fleisher M. Soluble mesothelin related peptides (SMRP) and osteopontin as protein biomarkers for malignant mesothelioma: analytical validation of ELISA based assays and characterization at mRNA and protein levels. Clin Chem Lab Med. 2010 Feb;48(2):271-8.
- 143. Napolitano A, Antoine DJ, Pellegrini L, Baumann F, Pagano I, Pastorino S, Goparaju CM, Prokrym K, Canino C, Pass HI, Carbone M, Yang H. HMGB1 and Its Hyperacetylated Isoform are Sensitive and Specific Serum Biomarkers to Detect Asbestos Exposure and to Identify Mesothelioma Patients. Clin Cancer Res. 2016 Jun 15;22(12):3087-96.

- 144. Sandri A, Guerrera F, Roffinella M, Olivetti S, Costardi L, Oliaro A, Filosso PL, Lausi PO, Ruffini E. Validation of EORTC and CALGB prognostic models in surgical patients submitted to diagnostic, palliative or curative surgery for malignant pleural mesothelioma. J Thorac Dis. 2016 Aug;8(8):2121-7.
- 145. Panou V, Vyberg M, Weinreich UM, Meristoudis C, Falkmer UG, Røe OD. The established and future biomarkers of malignant pleural mesothelioma. Cancer Treat Rev. 2015 Jun;41(6):486-95.
- 146. Pass HI, Levin SM, Harbut MR, Melamed J, Chiriboga L, Donington J, Huflejt M, Carbone M, Chia D, Goodglick L, Goodman GE, Thornquist MD, Liu G, de Perrot M, Tsao MS, Goparaju C. Fibulin-3 as a blood and effusion biomarker for pleural mesothelioma. N Engl J Med. 2012 Oct 11;367(15):1417-27. doi: 10.1056/NEJMoa1115050. Erratum in: N Engl J Med. 2012;367(18):1768.
- 147. Kirschner MB, Cheng YY, Armstrong NJ, Lin RC, Kao SC, Linton A, Klebe S, McCaughan BC, van Zandwijk N, Reid G. MiR-score: a novel 6-microRNA signature that predicts survival outcomes in patients with malignant pleural mesothelioma. Mol Oncol. 2015 Mar;9(3):715-26.
- 148. Krasinskas AM, Bartlett DL, Cieply K, Dacic S. CDKN2A and MTAP deletions in peritoneal mesotheliomas are correlated with loss of p16 protein expression and poor survival. Mod Pathol. 2010 Apr;23(4):531-8.
- 149. Dacic S, Kothmaier H, Land S, Shuai Y, Halbwedl I, Morbini P, Murer B, Comin C, Galateau-Salle F, Demirag F, Zeren H, Attanoos R, Gibbs A, Cagle P, Popper H. Prognostic significance of p16/cdkn2a loss in pleural malignant mesotheliomas. Virchows Arch. 2008 Dec;453(6):627-35.

- 150. Cedrés S, Montero MA, Zamora E, Martínez A, Martínez P, Fariñas L, Navarro A, Torrejon D, Gabaldon A, Ramon Y Cajal S, Felip E. Expression of Wilms' tumor gene (WT1) is associated with survival in malignant pleural mesothelioma. Clin Transl Oncol. 2014 Sep;16(9):776-82.
- 151. Khanna S, Thomas A, Abate-Daga D, Zhang J, Morrow B, Steinberg SM, Orlandi A, Ferroni P, Schlom J, Guadagni F, Hassan R. Malignant Mesothelioma Effusions Are Infiltrated by CD3⁺ T Cells Highly Expressing PD-L1 and the PD-L1⁺ Tumor Cells within These Effusions Are Susceptible to ADCC by the Anti-PD-L1 Antibody Avelumab. J Thorac Oncol. 2016 Nov;11(11):1993-2005.
- 152. Combaz-Lair C, Galateau-Sallé F, McLeer-Florin A, Le Stang N, David-Boudet L, Duruisseaux M, Ferretti GR, Brambilla E, Lebecque S, Lantuejoul S. Immune biomarkers PD-1/PD-L1 and TLR3 in malignant pleural mesotheliomas. Hum Pathol. 2016 Jun;52:9-18.
- 153. Nguyen BH, Montgomery R, Fadia M, Wang J, Ali S. PD-L1 expression associated with worse survival outcome in malignant pleural mesothelioma. Asia Pac J Clin Oncol. 2018 Feb;14(1):69-73.
- 154. Chapel DB, Schulte JJ, Husain AN, Krausz T. Application of immunohistochemistry in diagnosis and management of malignant mesothelioma. Transl Lung Cancer Res. 2020 Feb;9(Suppl 1):S3-S27.
- 155. Affar EB, Carbone M. BAP1 regulates different mechanisms of cell death. Cell Death Dis. 2018 Nov 19;9(12):1151.
- 156. Jensen DE, Proctor M, Marquis ST, Gardner HP, Ha SI, Chodosh LA, Ishov AM, Tommerup N, Vissing H, Sekido Y, Minna J, Borodovsky A, Schultz DC, Wilkinson KD, Maul GG, Barlev N, Berger SL, Prendergast GC, Rauscher FJ 3rd. BAP1: a novel

- ubiquitin hydrolase which binds to the BRCA1 RING finger and enhances BRCA1-mediated cell growth suppression. Oncogene. 1998;16(9):1097-112.
- 157. BAP1 BRCA1 associated protein 1 [Homo sapiens (human)]—Gene—NCBI: Pubs; 2016. http://www.ncbi.nlm.nih.gov/pubmed/.
- 158. Murali R, Wiesner T, Scolyer RA. Tumours associated with BAP1 mutations. Pathology 2013;45:116–26.
- 159. Okino Y, Machida Y, Frankland-Searby S, Machida YJ. BRCA1-associated protein 1 (BAP1) deubiquitinase antagonizes the ubiquitin-mediated activation of FoxK2 target genes. J Biol Chem. 2015;290(3):1580-91.
- 160. Daou S, Hammond-Martel I, Mashtalir N, Barbour H, Gagnon J, Iannantuono NV, Nkwe NS, Motorina A, Pak H, Yu H, Wurtele H, Milot E, Mallette FA, Carbone M, Affar el B. The BAP1/ASXL2 Histone H2A Deubiquitinase Complex Regulates Cell Proliferation and Is Disrupted in Cancer. J Biol Chem. 2015;290(48):28643-63.
- 161. White AE, Harper JW. Emerging anatomy of the BAP1 tumor suppressor system. Science 2012;337:1463–4.
- 162. Misaghi S, Ottosen S, Izrael-Tomasevic A, Arnott D, Lamkanfi M, Lee J, Liu J, O'Rourke K, Dixit VM, Wilson AC. Association of C-terminal ubiquitin hydrolase BRCA1-associated protein 1 with cell cycle regulator host cell factor 1. Mol Cell Biol. 2009;29(8):2181-92.
- 163. Forma E, Jóźwiak P, Bryś M, Krześlak A. The potential role of O-GlcNAc modification in cancer epigenetics. Cell Mol Biol Lett. 2014;19(3):438-60.
- 164. Carbone M, Yang H, Pass HI, Krausz T, Testa JR, Gaudino G. BAP1 and cancer. Nat Rev Cancer. 2013;13(3):153-9.

- 165. Wang A, Papneja A, Hyrcza M, Al-Habeeb A, Ghazarian D. Gene of the month: BAP1. J Clin Pathol. 2016;69(9):750-753.
- 166. Bononi A, Yang H, Giorgi C, Patergnani S, Pellegrini L, Su M, Xie G, Signorato V, Pastorino S, Morris P, Sakamoto G, Kuchay S, Gaudino G, Pass HI, Napolitano A, Pinton P, Jia W, Carbone M. Germline BAP1 mutations induce a Warburg effect. Cell Death Differ. 2017;24(10):1694-1704.
- 167. Bononi A, Giorgi C, Patergnani S, Larson D, Verbruggen K, Tanji M, Pellegrini L, Signorato V, Olivetto F, Pastorino S, Nasu M, Napolitano A, Gaudino G, Morris P, Sakamoto G, Ferris LK, Danese A, Raimondi A, Tacchetti C, Kuchay S, Pass HI, Affar EB, Yang H, Pinton P, Carbone M. BAP1 regulates IP3R3-mediated Ca²⁺ flux to mitochondria suppressing cell transformation. Nature. 2017;546(7659):549-553.
- 168. Zhang Y, Shi J, Liu X, Feng L, Gong Z, Koppula P, Sirohi K, Li X, Wei Y, Lee H, Zhuang L, Chen G, Xiao ZD, Hung MC, Chen J, Huang P, Li W, Gan B. BAP1 links metabolic regulation of ferroptosis to tumour suppression. Nat Cell Biol. 2018 Oct;20(10):1181-1192.
- 169. Yu H, Pak H, Hammond-Martel I, Ghram M, Rodrigue A, Daou S, Barbour H, Corbeil L, Hébert J, Drobetsky E, Masson JY, Di Noia JM, Affar el B. Tumor suppressor and deubiquitinase BAP1 promotes DNA double-strand break repair. Proc Natl Acad Sci U S A. 2014;111(1):285-90.
- 170. Carbone M, Emri S, Dogan AU, Steele I, Tuncer M, Pass HI, Baris YI. A mesothelioma epidemic in Cappadocia: scientific developments and unexpected social outcomes. Nat Rev Cancer. 2007;7(2):147-54.

- 171. Roushdy-Hammady I, Siegel J, Emri S, Testa JR, Carbone M. Genetic-susceptibility factor and malignant mesothelioma in the Cappadocian region of Turkey. Lancet. 2001;357(9254):444-5.
- 172. Testa JR, Cheung M, Pei J, Below JE, Tan Y, Sementino E, Cox NJ, Dogan AU, Pass HI, Trusa S, Hesdorffer M, Nasu M, Powers A, Rivera Z, Comertpay S, Tanji M, Gaudino G, Yang H, Carbone M. Germline BAP1 mutations predispose to malignant mesothelioma. Nat Genet. 2011 Aug 28;43(10):1022-5.
- 173. Carbone M, Ferris LK, Baumann F, Napolitano A, Lum CA, Flores EG, Gaudino G, Powers A, Bryant-Greenwood P, Krausz T, Hyjek E, Tate R, Friedberg J, Weigel T, Pass HI, Yang H. BAP1 cancer syndrome: malignant mesothelioma, uveal and cutaneous melanoma, and MBAITs. J Transl Med. 2012;10:179.
- 174. Carbone M, Flores EG, Emi M, Johnson TA, Tsunoda T, Behner D, Hoffman H, Hesdorffer M, Nasu M, Napolitano A, Powers A, Minaai M, Baumann F, Bryant-Greenwood P, Lauk O, Kirschner MB, Weder W, Opitz I, Pass HI, Gaudino G, Pastorino S, Yang H. Combined Genetic and Genealogic Studies Uncover a Large BAP1 Cancer Syndrome Kindred Tracing Back Nine Generations to a Common Ancestor from the 1700s. PLoS Genet. 2015;11(12):e1005633.
- 175. Haugh AM, Njauw CN, Bubley JA, Verzì AE, Zhang B, Kudalkar E, VandenBoom T, Walton K, Swick BL, Kumar R, Rana HQ, Cochrane S, McCormick SR, Shea CR, Tsao H, Gerami P. Genotypic and Phenotypic Features of BAP1 Cancer Syndrome: A Report of 8 New Families and Review of Cases in the Literature. JAMA Dermatol. 2017;153(10):999-1006.
- 176. Wiesner T, Obenauf AC, Murali R, Fried I, Griewank KG, Ulz P, Windpassinger C, Wackernagel W, Loy S, Wolf I, Viale A, Lash AE, Pirun M, Socci ND, Rütten A,

Palmedo G, Abramson D, Offit K, Ott A, Becker JC, Cerroni L, Kutzner H, Bastian BC, Speicher MR. Germline mutations in BAP1 predispose to melanocytic tumors. Nat Genet. 2011 Aug 28;43(10):1018-21.

177. Piris A, Mihm MC Jr, Hoang MP. BAP1 and BRAFV600E expression in benign and malignant melanocytic proliferations. *Hum Pathol.* 2015;46:239-245.

178. Rai K, Pilarski R, Cebulla CM, Abdel-Rahman MH. Comprehensive review of BAP1 tumor predisposition syndrome with report of two new cases. Clin Genet. 2016;89(3):285e294.

179. Yoshikawa Y, Emi M, Hashimoto-Tamaoki T, Ohmuraya M, Sato A, Tsujimura T, Hasegawa S, Nakano T, Nasu M, Pastorino S, Szymiczek A, Bononi A, Tanji M, Pagano I, Gaudino G, Napolitano A, Goparaju C, Pass HI, Yang H, Carbone M. High-density array-CGH with targeted NGS unmask multiple noncontiguous minute deletions on chromosome 3p21 in mesothelioma. Proc Natl Acad Sci U S A. 2016;113(47):13432-13437.

180. Guo G, Chmielecki J, Goparaju C, Heguy A, Dolgalev I, Carbone M, Seepo S, Meyerson M, Pass HI. Whole-exome sequencing reveals frequent genetic alterations in BAP1, NF2, CDKN2A, and CUL1 in malignant pleural mesothelioma. Cancer Res. 2015;75(2):264-9.

181. Lo Iacono M, Monica V, Righi L, Grosso F, Libener R, Vatrano S, Bironzo P, Novello S, Musmeci L, Volante M, Papotti M, Scagliotti GV. Targeted next-generation sequencing of cancer genes in advanced stage malignant pleural mesothelioma: a retrospective study. J Thorac Oncol. 2015;10(3):492-9.

- 182. Churg A, Sheffield BS, Galateau-Salle F. New markers for separating benign from malignant mesothelial proliferations: are we there yet? *Arch Pathol Lab Med*. 2016;140:318-321.
- 183. Sheffield BS, Hwang HC, Lee AF, Thompson K, Rodriguez S, Tse CH, Gown AM, Churg A. BAP1 immunohistochemistry and p16 FISH to separate benign from malignant mesothelial proliferations. Am J Surg Pathol. 2015;39(7):977-82.
- 184. Churg A, Hwang H, Tan L, Qing G, Taher A, Tong A, Bilawich AM, Dacic S. Malignant mesothelioma in situ. Histopathology. 2018;72(6):1033-1038.2
- 185. McGregor SM, Dunning R, Hyjek E, Vigneswaran W, Husain AN, Krausz T. BAP1 facilitates diagnostic objectivity, classification, and prognostication in malignant pleural mesothelioma. *Hum Pathol.* 2015;46:1670-1678.
- 186. Carbone M, Shimizu D, Napolitano A, Tanji M, Pass HI, Yang H, Pastorino S. Positive nuclear BAP1 immunostaining helps differentiate non-small cell lung carcinomas from malignant mesothelioma. Oncotarget. 2016;7(37):59314-59321.
- 187. Yoshimura M, Kinoshita Y, Hamasaki M, Matsumoto S, Hida T, Oda Y, Nabeshima K. Diagnostic application of BAP1 immunohistochemistry to differentiate pleural mesothelioma from metastatic pleural tumours. Histopathology. 2017;71(6):1011-1014. 188. Cigognetti M, Lonardi S, Fisogni S, Balzarini P, Pellegrini V, Tironi A, Bercich L, Bugatti M, Rossi G, Murer B, Barbareschi M, Giuliani S, Cavazza A, Marchetti G, Vermi W, Facchetti F. BAP1 (BRCA1-associated protein 1) is a highly specific marker for differentiating mesothelioma from reactive mesothelial proliferations. Mod Pathol.

2015;28(8):1043-57.

- 189. Andrici J, Parkhill TR, Jung J, Wardell KL, Verdonk B, Singh A, Sioson L, Clarkson A, Watson N, Sheen A, Farzin M, Toon CW, Gill AJ. Loss of expression of BAP1 is very rare in non-small cell lung carcinoma. Pathology. 2016;48(4):336-40.
- 190. Chapel DB, Husain AN, Krausz T, McGregor SM. PAX8 Expression in a Subset of Malignant Peritoneal Mesotheliomas and Benign Mesothelium has Diagnostic Implications in the Differential Diagnosis of Ovarian Serous Carcinoma. Am J Surg Pathol. 2017;41(12):1675-1682.
- 191. Andrici J, Jung J, Sheen A, D'Urso L, Sioson L, Pickett J, Parkhill TR, Verdonk B, Wardell KL, Singh A, Clarkson A, Watson N, Toon CW, Gill AJ. Loss of BAP1 expression is very rare in peritoneal and gynecologic serous adenocarcinomas and can be useful in the differential diagnosis with abdominal mesothelioma. Hum Pathol. 2016;51:9-15.
- 192. Kapur P, Christie A, Raman JD, Then MT, Nuhn P, Buchner A, Bastian P, Seitz C, Shariat SF, Bensalah K, Rioux-Leclercq N, Xie XJ, Lotan Y, Margulis V, Brugarolas J. BAP1 immunohistochemistry predicts outcomes in a multi-institutional cohort with clear cell renal cell carcinoma. J Urol. 2014 Mar;191(3):603-10.
- 193. Shinozaki-Ushiku A, Ushiku T, Morita S, Anraku M, Nakajima J, Fukayama M. Diagnostic utility of BAP1 and EZH2 expression in malignant mesothelioma. Histopathology. 2017 Apr;70(5):722-733.
- 194. Bahnasy AA, El-Din RS, Sabri NA, Abdel-Rahman CA, Bastawisy AE. BAP1 gene mutations in Egyptian patients with advanced sporadic malignant pleural mesothelioma (MPM): relation with clinical outcomes and survival. Cancer Genet. 2018 Dec;228-229:83-92.

- 195. Chou A, Toon CW, Clarkson A, Sheen A, Sioson L, Gill AJ. The epithelioid BAP1-negative and p16-positive phenotype predicts prolonged survival in pleural mesothelioma. Histopathology. 2018 Feb;72(3):509-515.
- 196. Farzin M, Toon CW, Clarkson A, Sioson L, Watson N, Andrici J, Gill AJ. Loss of expression of BAP1 predicts longer survival in mesothelioma. Pathology. 2015 Jun;47(4):302-7.
- 197. McGregor SM, McElherne J, Minor A, Keller-Ramey J, Dunning R, Husain AN, Vigneswaran W, Fitzpatrick C, Krausz T. BAP1 immunohistochemistry has limited prognostic utility as a complement of CDKN2A (p16) fluorescence in situ hybridization in malignant pleural mesothelioma. Hum Pathol. 2017 Feb;60:86-94.
- 198. Forest F, Patoir A, Dal Col P, Sulaiman A, Camy F, Laville D, Bayle-Bleuez S, Fournel P, Habougit C. Nuclear grading, BAP1, mesothelin and PD-L1 expression in malignant pleural mesothelioma: prognostic implications. Pathology. 2018 Oct;50(6):635-641.
- 199. Arzt L, Quehenberger F, Halbwedl I, Mairinger T, Popper HH. BAP1 protein is a progression factor in malignant pleural mesothelioma. Pathol Oncol Res. 2014 Jan;20(1):145-51.
- 200. E, Huilgol K, Moffat D, Henderson DW, Klebe S. Malignant Mesothelioma, BAP1 Immunohistochemistry, and VEGFA: Does BAP1 Have Potential for Early Diagnosis and Assessment of Prognosis? Dis Markers. 2017;2017:1310478.
- 201. Baumann F, Flores E, Napolitano A, Kanodia S, Taioli E, Pass H, Yang H, Carbone M. Mesothelioma patients with germline BAP1 mutations have 7-fold improved long-term survival. Carcinogenesis. 2015 Jan;36(1):76-81.

- 202. Betti M, Casalone E, Ferrante D, Romanelli A, Grosso F, Guarrera S, Righi L, Vatrano S, Pelosi G, Libener R, Mirabelli D, Boldorini R, Casadio C, Papotti M, Matullo G, Magnani C, Dianzani I. Inference on germline BAP1 mutations and asbestos exposure from the analysis of familial and sporadic mesothelioma in a high-risk area. Genes Chromosomes Cancer. 2015 Jan;54(1):51-62.
- 203. Svoronos AA, Engelman DM, Slack FJ. OncomiR or Tumor Suppressor? The Duplicity of MicroRNAs in Cancer. Cancer Res 2016;76:3666-70.
- 204. Yu M, Liang H, Fu Z, Wang X, Liao Z, Zhou Y, Liu Y, Wang Y, Hong Y, Zhou X, Yan X, Yu M, Ma M, Zhang W, Guo B, Zhang J, Zen K, Zhang CY, Wang T, Zhang Q, Chen X. BAP1 suppresses lung cancer progression and is inhibited by miR-31. Oncotarget. 2016 Mar 22;7(12):13742-53.
- 205. Wang N, Li Y, Zhou J. miR-31 Functions as an Oncomir Which Promotes Epithelial-Mesenchymal Transition via Regulating BAP1 in Cervical Cancer. Biomed Res Int 2017;2017:6361420. doi:10.1155/2017/6361420.
- 206. Sarcognato S, Gringeri E, Fassan M, Di Giunta M, Maffeis V, Guzzardo V, Cillo U, Guido M. Prognostic role of BAP-1 and PBRM-1 expression in intrahepatic cholangiocarcinoma. Virchows Arch. 2019 Jan;474(1):29-37.
- 207. Brierley JD, Gospodarowicz MK, Wittekind C. TNM Classification of Malignant Tumours, 8th edition, Union Int. Cancer Control, 2016. 208. Tsao AS, Garland L, Redman M, Kernstine K, Gandara D, Marom EM. A practical guide of the Southwest Oncology Group to measure malignant pleural mesothelioma tumors by RECIST and modified RECIST criteria. J Thorac Oncol. 2011;6(3):598-601. 209. Galateau-Salle F, Churg A, Roggli V, Travis WD. World Health Organization Committee for Tumors of the Pleura. The 2015 World Health Organization Classification

- of Tumors of the Pleura: Advances since the 2004 Classification. J Thorac Oncol 2016;11:142–154.
- 210. Bott M, Brevet M, Taylor BS, Shimizu S, Ito T, Wang L, Creaney J, Lake RA, Zakowski MF, Reva B, Sander C, Delsite R, Powell S, Zhou Q, Shen R, Olshen A, Rusch V, Ladanyi M. The nuclear deubiquitinase BAP1 is commonly inactivated by somatic mutations and 3p21.1 losses in malignant pleural mesothelioma. Nat Genet. 2011;43(7):668-72.
- 211. de Reyniès A, Jaurand MC, Renier A, Couchy G, Hysi I, Elarouci N, Galateau-Sallé F, Copin MC, Hofman P, Cazes A, Andujar P, Imbeaud S, Petel F, Pairon JC, Le Pimpec-Barthes F, Zucman-Rossi J, Jean D. Molecular classification of malignant pleural mesothelioma: identification of a poor prognosis subgroup linked to the epithelial-to-mesenchymal transition. Clin Cancer Res. 2014;20(5):1323-34.
- 212. De Rienzo A, Archer MA, Yeap BY, Dao N, Sciaranghella D, Sideris AC, Zheng Y, Holman AG, Wang YE, Dal Cin PS, Fletcher JA, Rubio R, Croft L, Quackenbush J, Sugarbaker PE, Munir KJ, Battilana JR, Gustafson CE, Chirieac LR, Ching SM, Wong J, Tay LC, Rudd S, Hercus R, Sugarbaker DJ, Richards WG, Bueno R. Gender-Specific Molecular and Clinical Features Underlie Malignant Pleural Mesothelioma. Cancer Res. 2016;76(2):319-28.
- 213. Boffetta P, Righi L, Ciocan C, Pelucchi C, La Vecchia C, Romano C, Papotti M, Pira E. Validation of the diagnosis of mesothelioma and BAP1 protein expression in a cohort of asbestos textile workers from Northern Italy. Ann Oncol. 2018;29(2):484-489. 214. Kumar V, Patel S, Tcyganov E, Gabrilovich DI. The Nature of Myeloid-Derived Suppressor Cells in the Tumor Microenvironment. Trends Immunol. 2016;37(3):208-220.