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Induced resistance to control postharvest decay of fruit and vegetables

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Highlights

- Several treatments applied to fruit and vegetables can trigger host defenses
- Induced resistance can be systemic acquired resistance or induced systemic resistance
- Biocontrol agents, physical means, and natural compounds elicit host defense system
- Induced resistance is a strategy that contributes to control of postharvest decay
- Induced resistance can increase amounts of beneficial antioxidant compounds

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Abstract

More than one third of harvested fruit and vegetables are lost and do not reach the customers mainly due to postharvest decay. During the last decade, several postharvest fungicides have been excluded from the market, or their allowed residues have been significantly decreased. Therefore, there is growing interest in eco-friendly and safe alternatives to synthetic fungicides. Induced resistance has gained increasing attention as a sustainable strategy to manage postharvest decay of fruit and vegetables. Their natural resistance can be increased by various means, such as biocontrol agents or their secreted elicitors. Alternatively, physical means, such as UV-C, ozone, and heat treatment, can prime plant resistance through abiotic stress. Moreover, various defense-related phytohormones, biological elicitors, non-organic elicitors, and volatile organic compounds have been shown to induce plant resistance. During the last decades, new technologies have enabled the evaluation of gene expression, such as quantitative real time PCR and the most recent next-generation sequencing, and thus the quantification of physiological changes, which have revealed new knowledge about preharvest and postharvest induced resistance in response to various treatments. These techniques allow optimization of postharvest application of the control means, although these data cannot disregard the evaluation of *in vivo* effectiveness. The elicitation of host defenses prevents the appearance of resistant isolates of pathogens. Induced resistance can lead to increased levels of phenolic compounds in the plant tissues, which often have antioxidant properties that are highly beneficial to humans. Moreover, induced resistance preserves the natural microflora, which is rich in potential biocontrol agents, and which provides a combined approach in the control of postharvest decay that is sustainable and safe for both growers and consumers. This approach meets the requirements of integrated disease management on sustainable use of pesticides that in the EU is implemented through Directive 128/2009. This review summarizes recent achievements and knowledge of the elicitation of host defenses to control postharvest decay of fruit and vegetables, and provides an outlook on the new challenges in this fascinating subject.

Keywords: biostimulants, elicitors, induced systemic resistance, resistance inducers, systemic acquired resistance

1. Introduction

Recent investigations have shown that more than one third of harvested fruit and vegetables are lost (FAO, 2011; USDA, 2014; OECD, 2014). Most losses occur due to pathogen infections in the field or after harvest, which lead to postharvest decay, when fruit ripen and vegetables senesce. Moreover, during the last decade, several postharvest fungicides that often had wide spectra of targets have been withdrawn from the market, due to: (i) selection of resistant fungal isolates; (ii) toxicity to humans and the environment; (iii) increasing consumer concern toward risk of pesticide residues on products, with the consequent strict requirements from several major supply chains for the quantity and number of active ingredient(s) on foodstuffs, as percentages of maximum residue limit; and (iv) increasing costs of registration and re-registration (Romanazzi et al., 2016a). Therefore, there is growing interest in finding cheap, safe, and eco-friendly alternatives to synthetic fungicides for the control of postharvest decay of fresh produce. Induction of plant resistance by biological, chemical, or physical means is considered a sustainable strategy to manage postharvest decay of fruit and vegetables. This approach has gained increasing interest during recent years, in which we can see a high trend in papers dealing with induced resistance, from few ones recorded 30 years ago to more than 800 recorded yearly in 2013-2015 (Fig. 1), and due to new tools, further knowledge has been obtained on host responses to various methods of control (Hershkovitz et al., 2013; Gapper et al., 2014).

The beneficial effects of induced resistance in the postharvest environment were originally demonstrated about two decades ago. For example, the use of heat treatment to decrease chilling injury and disease incidence in fruit through the induction of host resistance has been extensively studied (Lurie and Pedreschi, 2014). Ultraviolet-C (UV-C) irradiation and exposure to sunlight have been shown to induce resistance to pathogens and chilling tolerance in many harvested commodities (Wilson et al., 1994; Ruan et al., 2015; Sivankalyani et al., 2016). More recently, different inducers, such as cell-wall components, plant extracts, compounds of biological origin, and synthetic chemicals, have been shown to trigger plant resistance to pathogen attack locally and systemically (Walters and Fountaine, 2009). Moreover, biological control agents can induce plant resistance to pathogens (Vallad and Goodman, 2004; Da Rocha and Hammerschmidt, 2005; Lyon, 2007). However, to correctly induce resistance in different plants, it is necessary to know and understand the host–microbe interactions, and the effects on postharvest physiology and handling of the different fruit and vegetables (Da Rocha and Hammerschmidt, 2005).

Here, we review the different biological, physical, and chemical inducers that have been shown to control postharvest diseases of fruit and vegetables, and highlight their proposed mechanisms of action.

2. Mechanisms involved in induced resistance

Various biotic inducers (e.g., fungi, bacteria, viruses, phytoplasma, insects) and abiotic stresses (e.g., chemical and physical inducers) can trigger resistance in plants, which is known as ‘induced resistance’ (Pieterse et al., 2012; Walters et al., 2013; Pieterse et al., 2014). These can produce rapid expression of defense responses (Conrath et al., 2002; Fu and Dong, 2013). Examples of treatments able to induce resistance in host tissues and of representative mechanisms involved are reported in Fig. 2. We can imagine induced resistance as produced by an array of treatments that elicit a cloud of defense responses. There are two types of induced resistance in plants: systemic acquired resistance (SAR) and induced systemic resistance (ISR). Both of these mechanisms can induce defenses that confer long-lasting protection against a broad spectrum of microorganisms, and are mediated by phytohormones, such as salicylic acid (SA), jasmonic acid (JA), and ethylene (ET). SAR requires the signal molecule SA and is associated with accumulation of pathogenesis-related (PR) proteins, which are believed to contribute to resistance (Durrant and Dong, 2004). Instead, the ISR pathway functions independently of SA, while it is dependent on JA and ET (van Wees et al., 1999).

This induced resistance does not directly activate plant defense responses, but activates the plant to a state of ‘alertness’, so that a future pathogen attack will be strongly and efficiently responded to. This phenomenon is also known as the ‘priming effect’ (Conrath et al., 2006; Jung et al., 2009), and one of the most known priming effects is root colonization by plant-growth-promoting rhizobacteria (PGPR), which induce plant development and ISR-mediated resistance (Vallad and Goodman, 2004; Verhage et al., 2010). While PGPR induces ISR, other inducers can activate SAR or both of these systems.

2.1. Systemic acquired resistance (SAR)

The mechanisms of SAR are based on SA-mediated defense. The transcription factor Nonexpressor of pathogenesis-related genes 1 (NPR1) is considered to be the master regulator of SA and SAR. Here, biotic, abiotic, chemical, and physical inducers can trigger defense responses locally, and can also induce the production of suggested mobile immune signals, including SA, methyl salicylic acid (MeSA), azelaic acid (AzA), glycerol 3-phosphate, and abietane-diterpenoid-dehydroabietinal (Park et al., 2007; Chaturvedi et al., 2012). One or more of these signals can lead to systemic defense ‘memory’ that can last for weeks to months, to protect the plant from future infection (Jung et al., 2009).

Cellular redox and reactive oxygen species (ROS) are modified during SAR. Both primary and secondary oxidative bursts are required for the onset of SAR (Alvarez et al., 1998). Furthermore, cellular redox has been shown to be modified during SAR, as initially oxidized, and then reduced. NPR1, a master regulator of SA and SAR, was shown to be reduced by thioredoxin. The NPR1 oligomer is disrupted and its monomers enter the nucleus, and activate SA-mediated defense (Tada et al., 2008). The transcription factor NPR1 and the activated SA-mediated defense response result in SAR and the activation of approximately 10% of the plant transcriptome (Fu and Dong, 2013). This defense response includes direct targets of the WRKY domain transcription factor family, and synthesis and secretion of various PR proteins, with activation of the mitogen-activated protein kinase (MAPK) cascade, the DNA repair machinery, histone modifications, and a whole arsenal of defense-related transcripts (Fu and Dong, 2013).

2.2. Induced systemic resistance (ISR)

Induced systemic resistance is known to reprogram plant-induced mechanisms based on JA and ET defenses that alleviate physiological, abiotic, and biotic stresses. Elicitors and effectors known as ‘microbe-associated molecular patterns’ can be identified by plant receptors (Bent and Mackey, 2007). This recognition has a key role in activation of innate immunity. The ISR responses to biotic or abiotic stresses are diverse and can elicit plant defense responses. These responses include: MAPK signaling, generation of ROS, the octadecanoic pathway (which synthesizes oxidized fatty acid signals known as oxylipins), the phenylpropanoid pathway (which is involved in terpenoid and phytoalexin biosynthesis), increased levels of phenolic compounds, lignification at the site of pathogen infection, and cell-wall metabolism (Shoresh et al., 2010; Lloyd et al., 2011). ISR activates hydrogen peroxide producers on the one hand, such as oxalate oxidase and glucose oxidase, and antioxidants on the other hand, such as peroxidase (POD) and superoxide dismutase (SOD) (Shoresh et al., 2010). The balance between these two determines the ROS levels, and high ROS levels can lead to lipid peroxidation (Mittler, 2002). Additionally, several key transcripts, such as lipoxygenase (LOX1), phenylalanine ammonia lyase (PAL), and heat-shock proteins (HSPs), have been shown to be induced during ISR (Bi et al., 2007; Shoresh et al., 2010).

2.3. Other mechanisms of induced resistance

While the main induced resistance mechanisms are SAR and ISR, some processes of induced resistance combine these two resistance mechanisms in various ways. For example, β -aminobutyric acid (BABA)-induced resistance involves both SA-dependent and abscisic acid (ABA)-dependent defense mechanisms (Buonaurio et al., 2009; Pieterse et al., 2009). The relative importance of these

phytohormone-dependent defenses varies according to the nature of the challenge pathogen. Indeed, BABA-induced resistance against *Botrytis cinerea* resembles SAR and requires SA accumulation (Zimmerli et al., 2000), while the ABA-dependent pathway, which is associated to callose deposition, is necessary against *Hyaloperonospora parasitica* and *Plectosphaerella cucumerina* (Zimmerli et al., 2000; Ton and Mauch-Mani, 2004).

3. Induced resistance by biocontrol agents

Numerous reports have indicated that biocontrol agents, such as antagonistic yeast, can increase fruit resistance against postharvest diseases (Ippolito et al., 2000; Tian et al., 2006; Janisiewicz et al., 2008; Droby et al., 2016; Spadaro and Droby, 2016) (Tab. 1). As defense responses in plants are complex and involve both biochemical and structural barriers, the mechanisms of biocontrol agents are usually multiple. One is the secretion of extracellular lytic enzymes, as for *Pichia membranifaciens* and *Cryptococcus albidus*, where these can attach and degrade the hyphae of *Monilinia fructicola*, *Penicillium expansum* and *Rhizopus stolonifer*, both in vitro and in vivo (Chan and Tian, 2005). The second is the accumulation of host PR proteins (Jijakli and Lepoivre, 1998). These are strongly induced in response to wounding or infection by pathogens, and they accumulate abundantly at the site of infection, to contribute to SAR (Ryals et al., 1996). Treatment with *Cryptococcus laurentii* noticeably stimulated expression of the β -1,3-glucanase (*Glu-1*) gene in jujube fruit (Tian et al., 2007), which suggested that *Glu-1* has a role in defense responses to fungal pathogens. *C. laurentii* and *P. membranifaciens* have been shown to reduce disease incidence in pears (Tian et al., 2006), peaches (Xu et al., 2008a), and table grapes (Meng and Tian, 2009), via enhancement of defense-related enzyme activities, such as chitinase (CHT), β -1,3-glucanase (GLU) and PAL. *Aureobasidium pullulans* induced the production of CHT, GLU and POD in apple tissues starting 24 h after treatment, which reached maximum levels 48 h and 96 h after treatment (Ippolito et al., 2000). Then, the induction of host antioxidant enzymes and specific proteins has a relevant role. *P. membranifaciens* can induce host hydrogen peroxide metabolism, to enhance the resistance of sweet cherry against blue mold by *P. expansum* (Chan and Tian, 2006). Additionally, four antagonistic yeasts (*P. membranifaciens*, *C. laurentii*, *Candida guilliermondii*, *Rhodotorula glutinis*) can stimulate catalase (CAT) and POD activity, and reduce the levels of protein carbonylation in response to ROS caused by *M. fructicola* in peach fruit (Xu et al., 2008b). These results suggest that yeast treatments can alleviate protein carbonylation and pathogen-induced oxidative damage, which implies that the antioxidant defense response is involved in the mechanisms of microbial biocontrol agents against fungal pathogens. Other yeasts, such as *P. membranifaciens*, *C. guilliermondii*, and *R. glutinis*, have

also been shown to stimulate the expression of both PR proteins and redox enzymes (e.g., CHT, GLU, CAT, POD) in peach fruit during all storage periods (Xu et al., 2008b). *Candida saitoana* induced postharvest systemic resistance in apple fruit against *B. cinerea*, with increased activities of CHT and GLU reported (El Ghaouth et al., 2003b). *Candida intermedia* induced strawberry fruit defense mechanisms, which included β -1,3-exoglucanase, and was effective in postharvest control of *B. cinerea* (Huang et al., 2011a). Higher PAL, CHT and GLU activities were observed when *Pichia guilliermondii* and *C. laurenti* were applied to harvested tomatoes (Zhao et al., 2008).

4. Induced resistance by physical means

Several physical means, such as ultraviolet-C (UV-C) light, heat, hypobaric and hyperbaric treatments are known to be effective in controlling postharvest decay of fruit and vegetables (Usall et al., 2016). The advantage of most of these relies on direct effects on the pathogen without leaving residues on the fruit (Sanzani et al., 2009a). Moreover, these can induce several changes in host tissues, including increased resistance to abiotic and biotic stress (Tab. 2).

4.1. UV-C irradiation

The eliciting effects of UV-C irradiation have been studied for different fruit and vegetables (Charles and Arul, 2007). The first studies on the effects of UV-C irradiation on host tissues were carried out with carrots, the phytoalexin content of which was increased by the treatment (Mercier et al., 1993). Strawberries exposed to UV-C at 0.50 kJ m⁻² and 1.00 kJ m⁻² increased their PAL activity 12 h after treatment (Nigro et al., 2000). Peach fruit treated with UV-C showed increases in PAL, CHT and GLU activities (El Ghaouth et al., 2003a). Tomatoes exposed to UV-C and later inoculated with *R. stolonifer* showed 40% reduction in polygalacturonase (PG) activity, as compared to the control 72 h after the challenge (Stevens et al., 2004). The application of UV-C to harvested table grape berries increased the content of trans-resveratrol and catechin in the skin (Cantos et al., 2000; Romanazzi et al., 2006). A higher production of both compounds occurred when the berries were treated with chitosan 48 h before harvest and later exposed to UV-C irradiation (Romanazzi et al., 2006).

4.2. Heat treatment

Temperature (both high and low) is one of the oldest means to control postharvest diseases of fruit and vegetables. In particular, the use of low temperature is the most diffuse means of control, and nowadays imperfect management of this cold chain can cause heavy losses of fresh produce (Romanazzi et al., 2016a). The physiological changes in host tissues induced by heat treatment were

well described by Lurie and Pedreschi (2014). In peaches exposed to hot water treatment, cell-wall genes involved in ripening, such as β -galactosidase, pectin lyase, PG, and pectin methyl esterase, showed general decreased expression, while ROS scavenging genes and PAL, CHT, and HSP70 showed increased expression (Spadoni et al., 2014). A microarray analysis run on apples treated at 45 °C for 45 min and inoculated with *P. expansum* showed up-regulation of HSP, HS cognate protein, and HS transcription factor genes, which were linked to resistance of the fruit to the heat stress (Spadoni et al., 2015). Thus, this short hot water treatment has been shown to induce resistance to chilling and pathogens in various fruit (Fallik, 2004; Lurie and Pedreschi, 2014).

4.3. Hypobaric and hyperbaric treatment

Hypobaric and hyperbaric treatments rely on the use of pressure different from atmospheric pressure over a short period of time. These treatments do not have direct effects on the pathogen, although they affect the host tissues (Romanazzi et al., 2008). For hyperbaric treatment the occurrence of physiological changes is presumed, but not yet clarified, while the variations in host tissues induced by hypobaric applications have been well known since the middle of the last century, when they were applied to fresh fruit to delay ripening (Burg and Burg, 1966). The reduced pressure affects ET metabolism, which reduces respiration, delays ripening, and makes the fruit less prone to decay (Lougheed et al., 1978). Short hypobaric treatment has been shown to be an effective means of control of postharvest decay of strawberries, sweet cherries, and table grapes (Romanazzi et al., 2001), and increased activities of PAL, CHT, and POD were observed in strawberry exposed to 0.5 atm for 4 h (Hashmi et al., 2013). In contrast, the mechanisms of action of hyperbaric treatments are still not clear. However, this treatment changed lycopene accumulation in tomatoes, as it reduced during storage and increased during ripening (Liplap et al., 2013).

5. Induced resistance by natural and synthetic chemicals

5.1. Phytohormones and chemical elicitors

Phytohormones are well-known in the control of defense responses to pathogens and in the modulation of plant induced resistance (Alkan and Fortes, 2015). A central role in the regulation of plant immune responses has been ascribed to the defense hormones SA, JA, ABA, and ET in the regulation of plant–pathogen interactions (Fujita et al., 2006; Spoel and Dong, 2008). Gibberellic acid, auxin indolacetic acid, brassinosteroids (BR), and cytokinines have recently emerged as important modulators of plant defenses against microorganisms, mostly based on vegetative tissue data and on the lifestyle of the infecting pathogen (Robert-Seilanianantz et al., 2011). The SA and JA signaling pathways are generally considered as antagonistic and are dependent on NPR1 (Spoel et al.,

2007; Spoel and Dong, 2008; Pieterse et al., 2012). This interplay between SA and JA was suggested to optimize the host response to the pathogen lifestyle (Glazebrook, 2005; Spoel and Dong, 2008; Pieterse et al., 2012; Siciliano et al., 2015). In vegetative tissues, it is commonly postulated that effective responses to biotrophic pathogens are typically mediated by SA and programmed cell death, and responses to necrotrophic pathogens, which benefit from host cell death, involve JA and ET signaling (Glazebrook, 2005; Spoel and Dong, 2008). Examples of changes in gene expression or enzyme activities in response to application of natural and synthetic chemicals are reported in Tab. 3.

5.1.1. Salicylic acid and its analogs

SA participates in the mechanisms of defense, plant development, fruit ripening, and responses to various abiotic factors. SA application to an active necrotrophic infection with *Colletotrichum gloeosporioides* led to programmed cell death and increased susceptibility (Alkan et al., 2012). Similarly, infection with *C. gloeosporioides* on a ripe *NahG* tomato fruit mutant that lacked SA responses showed increased tolerance to *C. gloeosporioides* (Alkan and Fortes, 2015). Also, preharvest and postharvest treatments with SA for latent infection of *C. gloeosporioides* effectively reduced the occurrence of anthracnose of mango (Zainuri et al., 2001). Application of SA at 0.14 mg/mL by preharvest spraying or soaking before storage induced resistance to gray mold in kiwi fruit (Poole et al., 1998). SA at 0.05 mM in combination with ultrasound induced higher disease resistance to blue mold in peach fruit (Yang et al., 2011). Treatment with 0.5 mM SA reduced incidence and severity of decay caused by *P. expansum* on sweet cherries (Chan and Tian, 2006).

Treatment with SA and its analogs induces the accumulation of ROS, which can kill pathogen cells (Baker and Orlandi, 1995; Mittler et al., 2011). ROS levels are determined by the activities of enzymes, such as NADPH oxidase, that are designated as ‘respiratory burst oxidase homologs’, and antioxidants, such as SOD, ascorbate peroxidase (APX), glutathione reductase (GR), and CAT (Sharma et al., 2012). The induced host resistance was closely related to the levels of hydrogen peroxide and the activities of antioxidant enzymes (Liu et al., 2005; Ren et al., 2012; Dickman and Fluhr, 2013; Ge et al., 2015). On the other hand, activation of NADPH oxidase promotes the SA defense response (Alkan et al., 2012).

SA increased the activities of CHT, PAL, GLU and GR, and reduced the activities of CAT and APX in pears (Cao et al., 2006). A combination of SA and an antagonistic yeast significantly increased the activities of polyphenol oxidase (PPO), PAL, and GLU activities in cherries (Qin et al., 2003). Additionally, in cherry, increases in the activities of CAT, POD, APX, and SOD were observed after application of MeSA (Valverde et al., 2015).

Proteome research has shown that antioxidant and PR proteins, as well as enzymes associated with sugar metabolism, are involved in resistance of peach and sweet cherry fruit treated with SA (Chan et al., 2007). Therefore, the induced resistance by SA in fruit and vegetables activates a global defense response, which includes activation of the phenylpropanoid pathway, induction of accumulation of ROS and antioxidants, and production of PR proteins (Liu et al., 2014; Wang et al., 2015b).

5.1.2. *Benzothiadiazole (BTH)*

Benzothiadiazole (benzo(1,2,3)-thiadiazole-7-carbothioic acid S-methyl ester, also known as BTH or ASM) is perhaps the most potent synthetic elicitor discovered to date (Terry and Joyce, 2004; Bi et al., 2007). BTH is a light-insensitive functional analog of SA that induces resistance against a broad range of pathogens through activation of SAR in plants. Preharvest and postharvest BTH treatments have effectively reduced latent infections and induced resistance to diseases in fruit and vegetables, including strawberries (Terry and Joyce, 2004; Mazaro et al., 2008; Cao et al., 2011; Feliziani et al., 2015), pears (Cao et al., 2006), peaches (Liu et al., 2005), melons (Ren et al., 2012; Liu et al., 2014; Li et al., 2015b), and potatoes (Bokshi et al., 2003).

Preharvest BTH treatment significantly reduced *Alternaria* rot and blue mold of pears during storage (Cao et al., 2005). The same authors reported that the activities of PR proteins such as POD, CHT, and GLU were significantly enhanced in pears treated with BTH. Similar results were also observed in potatoes (Bokshi et al., 2003), peaches (Liu et al., 2005), and melons (Bi et al., 2006a).

Postharvest application of BTH to strawberries induced gene expression and increased activity of a range of enzymes that included several that are linked to biotic stress resistance (Landi et al., 2014). BTH-induced disease resistance enhanced gene expression of PPO and POD, and up-regulation of these genes was related to accumulation of total phenolic compounds, in harvested mango fruit (Lin et al., 2011). Postharvest dipping with BTH at 100 mg/L reduced artificial and natural infections in melons, while concentrations greater than 300 mg/L failed to promote resistance and caused phytotoxicity (Bi et al., 2006a).

5.1.3. *2,6-Dichloronicotinic acid (INA)*

2,6-Dichloronicotinic acid (INA) is a synthetic compound that is a structural and functional analog of SA, and it has been reported to mediate resistance against a broad spectrum of pathogens and its induced resistance has been suggested to have long-lasting effects (Lucas, 1999).

Preharvest foliar spray of INA at 50 mg/L significantly reduced postharvest diseases of melons (Bokshi et al., 2006). The resistance against *C. gloeosporioides* in mango was noticeably enhanced

by preharvest treatment with INA (Santiago et al., 2006). INA at 0.5 g/L also effectively reduced the disease spot sizes on the peel of banana fruit when inoculated with *C. musae* (Huang et al., 2011b).

5.1.4. Jasmonic acid (JA) and methyl jasmonate (MeJA)

Application of JA and methyl jasmonate (MeJA) can control decay incidence of several fruit. Postharvest JA treatment at 0.01 mM reduced green mold of grapefruit and orange (Porat et al., 2002). The optimal concentration of JA or MeJA varies for different fruit, and for different diseases on the same fruit. The effective concentration of MeJA to control tomato anthracnose was 0.0448 mM (Tzortzakis, 2007), while it was 10 mM for the control of tomato gray mold (Zhu and Tian, 2012). When applied by infiltration, the concentration of MeJA was reduced to 0.1 mM (Yu et al., 2009).

JA stimulated production of signaling molecules related to resistance and accumulation of antimicrobial compounds, and strengthened the structural barriers that restrict pathogen infection (Tian et al., 2007). MeJA treatment promoted higher PAL activity and increased total phenolics, flavonoids, and anthocyanins (Wang et al., 2009b). MeJA treatment promoted early accumulation of hydrogen peroxide, and increased gene expression of Cu-Zn SOD, CAT, and APX, at the same time. MeJA treatment also enhanced the contents of ascorbic acid and glutathione, which can scavenge excess ROS to alleviate protein oxidation injury (Zhu and Tian, 2012). MeJA treatment induced resistance against *Penicillium citrinum* by priming defense responses, and up-regulated the hydrogen peroxide burst and enhanced translation levels of defense-related proteins and the contents of antimicrobial compounds in Chinese bayberries (Wang et al., 2014).

5.1.5. Other chemical elicitors

Brassinosteroids (BRs) are a group of phyto-steroidal hormones that have crucial roles in a wide spectrum of biochemical, physiological, growth, and developmental processes in plants. A remarkable feature of BRs is their potential to increase resistance to a wide spectrum of stress in plants (Krishna, 2003). BRs at 5 μ M effectively inhibited development of blue mold rot and enhanced the activities of defense-related enzymes in jujube fruit. BRs did not show in vitro antimicrobial activity against *P. expansum* (Zhu et al., 2010).

Although BABA is only rarely found naturally in plants, it has been shown to be a potent inducer of acquired resistance and has a broad spectrum of activity against many pathogens (Conrath et al., 2001). BABA induced resistance of mango fruit to postharvest anthracnose caused by *C. gloeosporioides*, and enhanced the activity of fruit defense mechanisms (Zhang et al., 2013). BABA caused short-lasting activation of CHT and POD after a first spray, and a boost after a second spray, although to a lower level than that caused by INA (Bokshi et al., 2006).

The application of oxalic acid has been shown to induce systemic resistance against postharvest diseases (Zheng et al., 2005). Oxalic acid induced defense-related enzymes and priming of defense responses, enhanced the contents of antimicrobial compounds and modulated antioxidant enzymes in muskmelon (Deng et al., 2015). Oxalic acid at 5 mM increased antioxidant levels and PPO activity, which was beneficial for delayed ripening and enhanced disease resistance in peach fruit during storage at low temperature (Zheng et al., 2005).

L-arginine induced disease resistance via its effects on nitric oxide (NO) biosynthesis and defensive enzyme activity in tomato fruit (Zheng et al., 2011).

Riboflavin at 1.0 mM inhibited development of *Alternaria* rot, enhanced the activities of defense-related enzymes, such as PAL, PPO and POD, and increased accumulation of flavonoids, phenolics, and lignin (Li et al., 2012b).

1-Methylcyclopropene induced resistance against postharvest decay in jujube fruit, and increased PAL, PPO, CAT, and SOD activities (Zhang et al., 2012).

5.2. Biological elicitors

5.2.1. Bacterial effectors

5.2.1.1. Harpin

Harpin is an acidic, heat-stable, glycine-rich, 44-kDa protein encoded by the *hrpN* gene, and it was first described in *Erwinia amylovora*, which causes fire blight of *Rosaceae*. Bacterial harpin has been shown to elicit the hypersensitive response and to induce SAR in plants (Baker and Orlandi, 1995). This elicitor has also been shown to induce resistance in some postharvest fruit and vegetables (Bi et al., 2007). Postharvest treatment with harpin at 0.04 mg/L to 0.16 mg/L inhibited incidence of blue mold in apple (De Capdeville et al., 2003). Field spraying with harpin at 50 mg/L reduced latent infections in muskmelons caused by *A. alternata* and *Fusarium* spp., with reductions proportional to the levels of applied harpin, up to 90 mg/L (Wang et al., 2011b). However, harpin higher than 90 mg/L failed to promote resistance in melons (Bi et al., 2005). Harpin reduced lesion diameter in both treated and untreated halves of the same melon, which indicated that SAR was induced by harpin (Bi et al., 2005). Some of the defense reactions in melons elicited by harpin were identified as alterations in the levels of preformed antifungal substances, such as phenols and flavonoids, accumulation of PR proteins, such as CHT and GLU, induction of enzyme activities, such as PAL and POD, modulation of metabolism of ROS, such as SOD, CAT, and hydrogen peroxide, and reinforcement of cell walls and lignin (Bi et al., 2005; Wang et al., 2011b; Zhu and Zhang, 2016).

5.2.1.2. Oligandrin

Oligandrin is an elicitor-like protein with molecular mass of >10 kDa that has been shown to be secreted by *Pythium oligandrum*. Oligandrin is known to induce resistance against a number of plant diseases. The treatment of tomato fruit with oligandrin at 10 µg/mL significantly reduced incidence and severity of gray mold, increased the activities of the defense-related enzymes, like PAL, PPO, and POD, and the mRNA levels of the genes encoding PR proteins, and activated the ET-dependent signaling pathway (Wang et al., 2011a).

5.2.2. Fungal effectors

5.2.2.1. Chitosan

Chitosan (poly-β-(1,4)-N-acetyl-d-glucosamine) is a natural biopolymer with wide antimicrobial properties that can elicit defense responses in fruit and vegetables (El Ghaouth et al., 1992; Romanazzi et al., 2002). This chemical and its derivatives, such as oligochitosan and glycol chitosan, can be used in solution, as powders, and as edible coatings (Romanazzi et al., 2016b). Preharvest and postharvest treatments with chitosan and its derivatives tend to suppress storage rots in many commodities, such as strawberry (El Ghaouth et al., 1992; Reddy et al., 2000), jujube (Yan et al., 2012), sweet cherries (Feliziani et al., 2013a), citrus fruit (Fajardo et al., 1998; Zeng et al., 2010), apples (Felipini and Di Piero, 2009), banana (Meng et al., 2012), table grapes (Romanazzi et al., 2002; Meng et al., 2008), and tomatoes (Liu et al., 2007; Badawy and Rabea, 2009), and for many other vegetables (Miranda-Castro, 2016).

Several studies have shown that chitosan has multiple mechanisms of action, with direct antimicrobial properties, film-forming activities, and induction of host defenses (Romanazzi et al., 2016b). There was a significant increase in CHT and GLU activities in banana and jujube treated with oligochitosan (Meng et al., 2012; Yan et al., 2012). Chitosan and oligochitosan treatments induced significant increases in the activities of PPO (Liu et al., 2007), POD (Liu et al., 2007; Yan et al., 2012), and PAL (Romanazzi et al., 2002; Meng et al., 2012; Landi et al., 2014) in several harvested products. Chitosan treatment enhanced the total content of phenolics, flavonoids, and other antifungal substances (El Ghaouth et al., 1992; Yan et al., 2012), and accumulated ROS through regulation of the activity of metabolic enzymes, such as SOD, CAT, and APX (Zeng et al., 2010; Yan et al., 2012; Landi et al., 2014). Spraying with chitosan increased the activities of PPO and PAL in table grapes, thus promoting protection from latent infection of *B. cinerea* (Romanazzi et al., 2002; Meng et al., 2008). Next-generation sequencing with chitosan-treated avocado has defined more genes as up-regulated than down-regulated (Gutiérrez-Martínez et al., 2016). CHT and ROS production in table grape berries varied according to the formulation of chitosan applied (Feliziani et al., 2013b). Structural defense responses, such as preservation of pectin binding sites and the intense and regular

cellulose distribution over host cell walls were observed in the first tissue layers beneath the ruptured cells in bell peppers treated with chitosan (El Ghaouth et al., 1997).

5.2.2.2. Burdock fructooligosaccharide

Burdock fructooligosaccharide (BFO) is a fructosan oligomer that effectively inhibited postharvest diseases and reduced incidence of gray mold in tomato. BFO increased mRNA levels of genes encoding PR proteins, such as PR-1a, PR-2a (extracellular GLU), PR-2b (intracellular GLU), PR-3a (extracellular CHT), and PR-3b (intracellular CHT), and induced accumulation of PAL mRNA in tomatoes (Wang et al., 2009a). BFO also effectively controlled postharvest diseases in grapes, apples, banana, kiwi fruit, citrus fruit, strawberries, and pears (Sun et al., 2013).

5.2.3. Other biological elicitors

Peach fruit pretreated with yeast saccharide activated CHT, GLU, PAL, and POD. Moreover, yeast saccharide triggered endogenous NO in peaches during storage (Yu et al., 2012). The flavonoid quercetin significantly reduced blue mold in apples, and genes differentially expressed in quercetin-treated apples revealed high similarities with different classes of PR proteins (i.e., RNase-like PR10, PR8), and with proteins expressed under stress conditions (Sanzani et al., 2009b; Sanzani et al., 2010). The resistance inducer protein hydrolysates were effective against green mold of citrus fruit and gray mold of table and wine grapes (Lachhab et al., 2015, 2016).

5.3. Inorganic elicitors

5.3.1. Silicon (Si)

Silicon (Si) is the second most abundant element in the Earth lithosphere, and it is as important as phosphorus and magnesium in the biota (Exley, 1998). Si is also considered to be biologically active and to trigger more rapid and extensive deployment of plant natural defenses. Guo et al. (2007) reported that Si oxide and sodium silicate suppressed pink rot in muskmelons. Sodium silicate at 100 mM reduced rots in melons caused by *A. alternata*, *F. semitectum*, and *T. roseum* (Bi et al., 2006b; Li et al., 2012a).

Sodium silicate has been shown to be effective for suppression of pathogen growth and for induction of resistance to postharvest diseases in fruit and vegetables. Si treatment did not affect the activities of POD, PPO, PAL, and GLU, or the content of total phenols and flavonoids in potato tuber, although these were significantly accumulated after a challenge with *F. sulphureum* (Li et al., 2009). These results indicated that a priming state was induced by Si in potato tubers (Conrath et al., 2001). In Hami melons, Si treatments caused activation of POD and CHT (Bi et al., 2006b). The effects of

Si on postharvest diseases of muskmelons were associated with elicitation of the antioxidant system (Li et al., 2012a).

5.3.2. Nitric oxide (NO)

Nitric oxide (NO) is an important bioactive molecule that serves as a signal in plants, in particular for maturation and senescence (Leshem et al., 1998). Lai et al. (2011) reported that NO treatment activated antioxidant enzymes and induced resistance against gray mold in tomato. NO increased accumulation of phytoalexin rishitin in potato tubers (Niritake et al., 1996), and induced resistance against dry rot in potato tubers (Hu et al., 2014a). Moreover, NO treatment modulated anthracnose levels and induced defense-related enzymes in mangoes (Hu et al., 2014b).

5.3.3. Sodium carbonate

Sodium carbonate and bicarbonate were effective in the control of green mold of citrus fruit (Youssef et al., 2014). These salts exerted direct antifungal effects on *Penicillium digitatum*, and increased the activities of resistance enzymes, such as GLU, POD, and PAL. Moreover, citrus peel extracts showed increased levels of sugars and phytoalexins, with sucrose and scoparone being the most represented.

6. Induced resistance by disinfecting agents

6.1. Ozone

Since its recognition in 1997 by the US Food and Drug Administration as a safe food disinfectant, both gaseous ozone (O₃) and ozonated water have gained particular attention for the control of postharvest diseases of fruit and vegetables. The most recent examples of its use are those reported for blueberries (Crowe et al., 2012), persimmons (Ikeura et al., 2013), papaya (Ali et al., 2014), and table grapes (Feliziani et al., 2014). Ozone is a strong oxidizing agent; however, as the O₃ disinfecting activity is limited to surface-contaminating microflora, and as various microbes show different susceptibilities (Pascual et al., 2007), further modes of action appear to be involved (Feliziani et al., 2016). Artes-Hernandez et al. (2007) reported that continuous flow of O₃ increased total flavonol and hydroxycinnamates contents in cold-stored 'Autumn Seedless' table grapes. Moreover, it has been reported that O₃ boosts maintained the total polyphenols, and greatly increased the phytoalexin resveratrol content in cold-stored 'Napoleon' table grapes (Artés-Hernández et al., 2003). Similarly, O₃ at 0.1 µmol/mol increased total phenolic content in red bell peppers (Glowacz et al., 2015). A proteomic analysis conducted on O₃-treated kiwi fruit identified 102 differentially expressed proteins that were mainly involved in energy, protein metabolism, defense, and cell structure (Minas et al., 2012). A set of candidate kiwifruit proteins was defined as sensitive to protein carbonylation, which

was induced by ripening and depressed by O₃. Moreover, O₃ treatment transiently increased expression of CHT and PAL in ‘Redglobe’ and ‘Sugraone’ table grapes (Duarte-Sierra et al., 2016) (Tab. 4).

6.2. Electrolyzed water

Electrolyzed water (EW) is obtained by adding a small amount of an electrolyte to tap water, which is traditionally sodium chloride, and passing an electrical current across an anode and a cathode. EW has high oxidation–reduction potential with strong direct effects against decay causing fungi (Guentzel et al., 2010). However, recently, it has shown resistance inducer potential. In particular, it caused 30% increase in the total phenolic content of broccoli (Navarro-Rico et al., 2014) and maintained fresh-cut cilantro firmness (Hao et al., 2015), thus improving their resistance to pathogen attack. More recently, other electrolytes have been tested to improve EW performance and to avoid production of chlorine by-products. Encouraging results were obtained using NaHCO₃ against postharvest rots of citrus fruit, which also induced host defense responses (Fallanaj et al., 2016). In particular, up-regulation of defense-related genes that encode CHT, POD, and PAL was observed at 6 h to 12 h post-treatment, with increased activity of the related enzymes and of GLU (Tab. 4). As this was observed at 12 h post-treatment, this suggested an early host response against *P. digitatum* by limiting tissue colonization.

6.3. Ethanol

The effects of ethanol dipping as a surface disinfectant that can reduce pathogen populations without impairing product quality are well known (Lichter et al., 2002; Mlikota Gabler et al., 2004; Lee et al., 2015). However, its mode of action appears to be much more complex. Ethanol induction of resistance to postharvest anthracnose in loquat fruit was demonstrated by Wang et al. (2015a). Ethanol at 300 µL/L inhibited anthracnose caused by *Colletotrichum acutatum*, and maintained overall quality. Moreover, it increased SOD activity, thus resulting in higher levels of hydrogen peroxide, which can activate disease resistance. Meanwhile, ethanol treatment significantly enhanced the activities of defense-related enzymes, including PAL, POD, PPO, CHT, and GLU (Tab. 4). Recently, ethanol treatments were shown to decelerate the ripening process and down-regulate expression of major lipoxygenase-encoding genes involved in melon fruit ripening, thus contributing to its increased resistance to biotic and abiotic stress (Zhang et al., 2015).

7. Induced resistance by microbial and plant volatile organic compounds (VOCs)

Biotic agents that contribute to plant disease management include PGPR, growth promoting fungi, and fungi that work as biocontrol agents (Lyon, 2007). In addition to the use of microorganisms and chemical compounds in plant protection, there is an important developing area of research in the control of postharvest pathogens in fruit and vegetables based on the use of volatile organic compounds (VOCs), which can be produced by microorganisms (MVOCs) or by plants (PVOCs) (Mari et al., 2016).

7.1. Induced resistance by microbial volatile organic compounds (MVOCs)

Microbial volatile organic compounds (MVOCs) are mixtures of carbon-based compounds that are highly volatile or are vapors (Morath et al., 2012). Fungi, yeasts, and bacteria produce MVOCs as primary and secondary metabolites (Jijakli and Lepoivre, 1998; Korpi et al., 2009). MVOCs have shown applicative potential as biofumigants and have motivated great interest, mainly because they occur naturally without chemical synthesis (Li et al., 2015a). MVOCs can be signaling substances for regulation and control of some physiological actions, which include induction of systemic resistance against pathogens (Ryu et al., 2004).

Some defense mechanisms activated by biocontrol bacteria are induced in fruit and leaves by MVOCs, including production of phytoalexins, PR proteins, such as CHT and GLU, and protein inhibitors (Conrath et al., 2006; Li et al., 2012c). *Bacillus* spp. VOCs (2-3-butanediol and lipopeptides) induced over-expression of the surfactin and fengicin genes, which caused metabolic changes in host tissues (Ryu et al., 2004; Ongena et al., 2007).

7.2. Induced resistance by plant volatile organic compounds (PVOCs)

The eliciting activities of PVOCs, such as MeSA, were reported in section 5.1.1. Citrus fruit produce VOCs that are actively involved in defense systems before pathogen attacks. The presence and variation of VOCs depend on the type of produce and of its development phase. VOCs occur naturally in plant systems and can be associated with the biochemistry of constitutive defense mechanisms (Wightwick et al., 2010). Structures in the petals of citrus flowers, called osmophores, can release more than 60 VOCs, such as phenols, terpenes, and lipophilic compounds, which have been recognized as antifungal agents (Caccioni et al., 1995; Lattanzio et al., 2006). It was reported that limonene and linalool have antifungal actions against *C. acutatum* and are associated with constitutive biochemical responses and can be used in the control of pathogens (Rodrigues Marques et al., 2015).

Essential oils produce their activity through vapor, and thyme and cinnamon essential oils increased PPO, PAL, CHT, GLU activities in peach fruit (Cindi et al., 2016). In tomato, it has been reported that VOCs are induced and modified in defense responses, and ripening processes, and by

wounding. In a microarray analysis of tomato fruit at different stages of ripening, more than 4,000 differentially expressed genes have been reported. Genes related to defense mechanisms were expressed in the early stages of ripening, and genes related to VOCs changed during late maturation. Overall, more than 40 VOCs were detected, and their profiles were characterized along the fruit ripening stages (Baldassarre et al., 2015).

8. Induced resistance perspectives

A large amount of data has been generated during the last few years that are related to the triggering of host defenses during postharvest (see as examples, Tables 1, 2, 3 and 4, and Fig. 1). The elicitation of host defenses has a central role nowadays in integrated disease management strategies, and this is welcome for the reasons summarized in Tab. 5. The effects on plants can last for weeks or months. Induced resistance has a wider spectrum of targets than synthetic fungicides. Induced resistance does not lead to the appearance of resistant strains, due to involvement of various modes of action. Induced resistance is usually applied before the appearance of the symptoms, so there are no side effects on nontarget organisms and on humans, in terms of farmers, people working in the packinghouses, retailers, or consumers. Moreover, induced resistance can lead to increased levels of phenolic compounds in the plant tissues, which often have antioxidant properties that are highly beneficial to humans. However, we cannot forget the weaker points linked to the application of strategies based on induced resistance, such as possible inconsistent results or difficulties in their implementation in packinghouse practices. Recently, a high number of biostimulants have appeared on the market, and are considered as part of the effective treatments. Usually these biostimulants have combinations of direct activities on pathogens and indirect activities on the host. Increasing interest in this novel approach that is based on the triggering of host defenses satisfies consumer demands and the guidelines on sustainable approaches to plant protection, which in European Union is implemented through Directive 128/2009 on sustainable use of fungicides. New tools, such as the -omics sciences, allow better understanding of the changes in host physiology and provide information on gene functions. This information will provide optimization of the application of alternative treatments to control postharvest decay.

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1148

1149 **Figure captions**

1150

1151 **Figure 1.** Number of articles available through Scopus over the last 30 years using the search
1152 keywords of “induced resistance postharvest” (accessed on June 16, 2016).

1153

1154 **Figure 2.** Treatments that can induce resistance in fresh fruit and vegetables, and the mechanisms
1155 or enzymes involved. Images from <http://cliparts.co/free-fruit-pictures>.

1156 INA, 2,6-Dichloronicotinic acid; HWT, Hot water treatment; GRAS, generally recognized as safe;
1157 JA, jasmonic acid; SA, salicylic acid; MeJa, methyl jasmonate; BFO, Burdock
1158 fructooligosaccharide; BTH, benzothiadiazole; EOW, Electrolyzed oxidizing water; NO, nitric
1159 oxide; UV-C, ultraviolet C irradiation; BCA, biocontrol agents; MVOCs, microbial volatile organic
1160 compounds; Si, silicon; PVOCs, plant volatile organic compounds; EtOH, ethanol; SC, sodium
1161 carbonate; SAR, systemic acquired resistance; PG, polygalacturonase; BABA, β -aminobutyric acid;
1162 GLU, β -1,3-glucanase; PR, pathogenesis related proteins; MAMP, microbe-associated molecular
1163 pattern; LOX, lipoxygenase; ROS, reactive oxygen species; CHT, chitinase; CAT, catalase; ABA,
1164 abscisic acid; PPO, polyphenol oxidase; PAL, phenylalanine ammonia lyase; ISR, induced systemic
1165 resistance; PAMP, pathogen-associated molecular pattern; SOD, superoxide dismutase; NPR1,
1166 nonexpressor of pathogenesis-related genes 1; hsp, heat shock protein.

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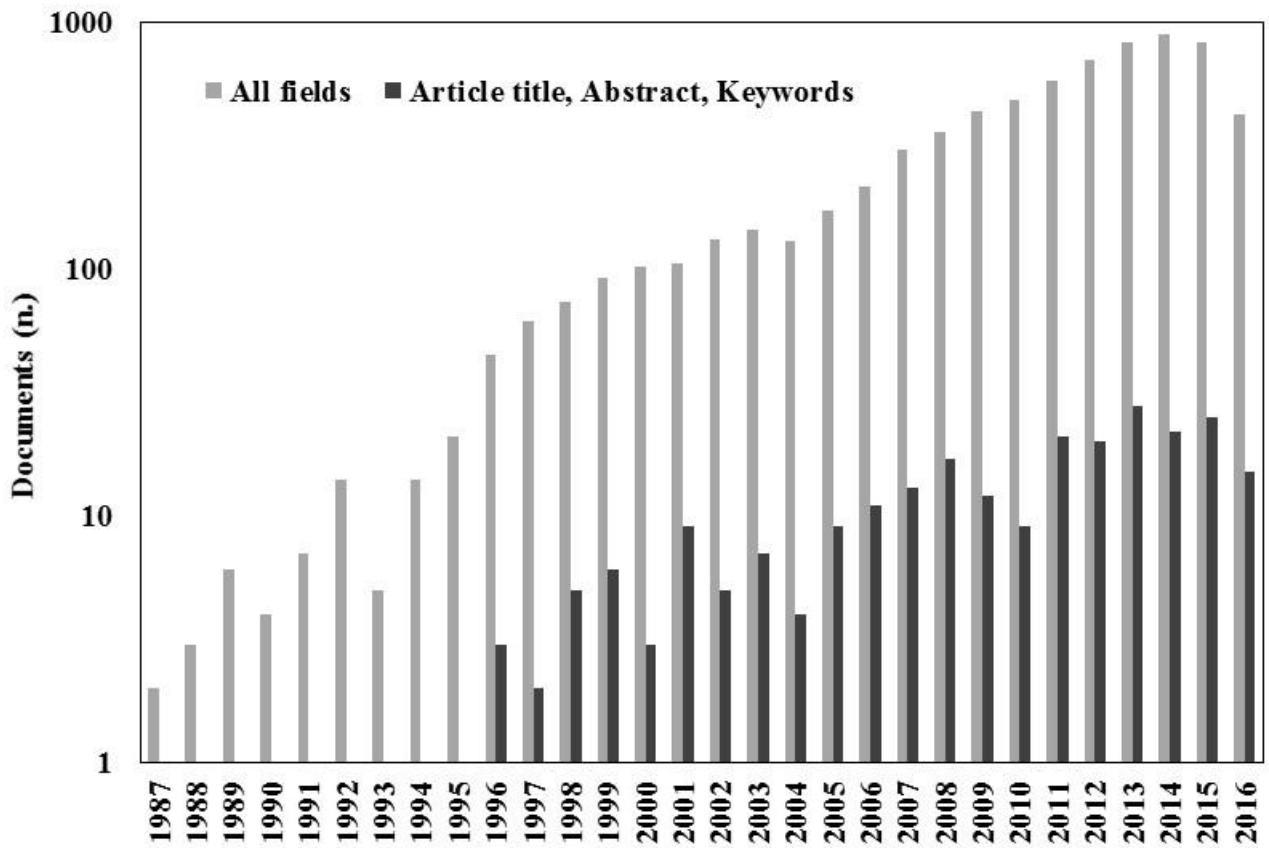


Fig. 1

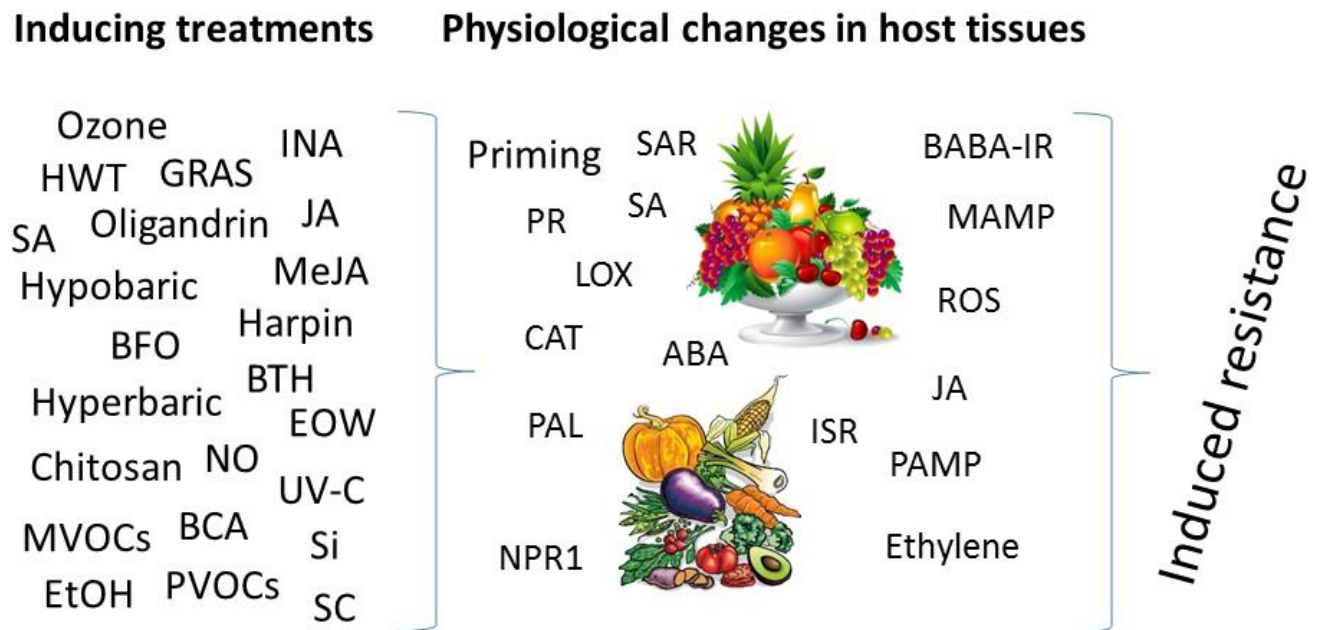


Fig. 2

Table 1. Examples of differential gene expression or enzyme activities in response to the application of biocontrol agents.

| Treatment | Genes and/or enzymes | | | | | | | | | | Reference |
|---|----------------------|------|-----|--------|----|--------|--------|-----------|----------|------|--------------------------|
| | PAL* | CHT | SOD | CAT | SA | LOX | GLU | SURFACTIN | FENGICIN | POD | |
| <i>Bacillus subtilis</i> | | | | | | +/++** | | +/++++ | +/++++ | | Ongena et al., 2007 |
| <i>Candida saitoana</i> | | +/++ | | | | | + | | | | El Ghaouth et al., 2003b |
| <i>Cryptococcus laurentii</i> | | | | | | | + | | | | Tian et al., 2007 |
| <i>Pichia membranifaciens</i> , <i>Candida guilliermondii</i> <i>Rhodotorula glutinis</i> | | + | | +/++++ | | | +/++++ | | | +/++ | Xu et al., 2008b |
| <i>Pichia guilliermondii</i> | + | + | + | -/+ | | | + | | | -/+ | Zhao et al., 2008 |
| <i>Aureobasidium pullulans</i> | | + | | | | | + | | | + | Ippolito et al., 2000 |

*PAL, phenylalanine ammonia lyase; CHT, chitinase; SOD, superoxide dismutase; CAT, catalase; SA, salicylic; LOX, lipoxygenase; GLU, β -1,3-glucanase; POD, peroxidase

**+, overexpressed up to 3-fold; ++, overexpressed from 4-fold to 10-fold; +++, overexpressed more than 10-fold; -, down-regulated up to 3-fold

Table 2. Examples of differential gene expression or enzyme activities in response to application of physical means.

| Treatment | Genes and/or enzymes | | | | | | Reference |
|---------------------|----------------------|--------|-----|-----|--------------------------|----------|---|
| | PAL* | CHT | GLU | POD | <i>Trans-resveratrol</i> | Catechin | |
| UV-C irradiation | + | ++/+++ | + | | ++/+++ | ++ | Cantos et al., 2000; Nigro et al., 2000; El Ghaouth et al., 2003a; Romanazzi et al., 2006 |
| Heat treatment | +/- | - | | | | | Spadoni et al., 2014, 2015 |
| Hypobaric treatment | + | + | | + | | | Hashmi et al., 2013 |

*PAL, phenylalanine ammonia lyase; CHT, chitinase; GLU, β -1,3-glucanase; POD, peroxidase

**+, overexpressed up to 3-fold; ++, overexpressed from 4-fold to 10-fold; +++, overexpressed more than 10-fold; -, down-regulated up to 3-fold

Table 3 Examples of differential gene expression or enzyme activities in response to application of natural and synthetic chemicals.

| Treatment | Genes and/or enzymes | | | | | | | | Reference |
|----------------------------|----------------------|-----|-----|------|------|-------|-------|-----|--|
| | SOD | CAT | POD | APX | CHT | PAL | GLU | PPO | |
| Salicylic acid | | -** | ++ | - | + | ++ | + | | Cao et al., 2006 |
| Methyl salicylic acid | + | ++ | ++ | + | | | | | Valverde et al., 2015 |
| Benzothiadiazole | + | + | ++ | | + | + | ++ | | Cao et al., 2005 |
| β -aminobutyric acid | | | + | | + | | | | Bokshi et al., 2006 |
| Riboflavin | | | ++ | | | + | | + | Li et al., 2012b |
| 1-Methylcyclopropene | + | + | | | | + | | + | Zhang et al., 2012 |
| Harpin | | | +++ | | +++ | | | | Bi et al., 2005 |
| Oligandrin | | | + | | | + | | + | Wang et al., 2011a |
| Chitosan | | | + | +/++ | +/++ | +/+++ | +/+++ | + | Romanazzi et al., 2002; Liu et al., 2007; Meng et al., 2012; Yan et al., 2012; Feliziani et al., 2013b; Landi et al., 2014 |
| Yeast saccharide | | | ++ | | + | +++ | +++ | | Yu et al., 2012 |
| Silicon | | | +++ | | +++ | | | | Bi et al., 2006b |
| Sodium carbonate | | | ++ | | - | ++ | ++ | | Youssef et al., 2014 |

SOD, superoxide dismutase; CAT, catalase; POD, peroxidase; APX, ascorbate peroxidase; CHT, chitinase; PAL, phenylalanine ammonia lyase; GLU, β -1,3-glucanase; PPO, polyphenol oxidase

**+, overexpressed up to 3-fold; ++, overexpressed from 4-fold to 10-fold; +++, overexpressed more than 10-fold; -, down-regulated up to 3-fold

Table 4. Examples of differential gene expression or enzyme activities in response to application of disinfecting agents.

| Treatment | Genes and/or enzymes | | | | | | Reference |
|--------------------|----------------------|-----|-----|-----|-----|-----|----------------------------|
| | PPO* | GLU | PAL | LOX | POD | CHT | |
| Ozone | | | *** | | | + | Duarte-Sierra et al., 2016 |
| Electrolyzed water | | | + | | ++ | +++ | Fallanaj et al., 2016 |
| Ethanol | | | | - | | | Zhang et al., 2015 |

*PPO, polyphenol oxidase; GLU, β -1,3-glucanase; PAL, phenylalanine ammonia lyase; LOX, lipoxygenase; POD, peroxidase; CHT, chitinase

***+, overexpressed up to 3-fold; ++, overexpressed from 4-fold to 10-fold; +++, overexpressed more than 10-fold; -, down-regulated up to 3-fold

Table 5. Aspects related to the induction of resistance to postharvest diseases of fruit and vegetables

| Negative sides | Positive sides |
|--|---|
| Complete effect is not always reproducible | Long-lasting effects |
| Does not provide a complete control of decay | Broad range of targets |
| Not easy to implement as part of farmer and packinghouse practices | Do not cause appearance of resistant isolates of the pathogen |
| Investigation methods are not standardized | Increasing number of biostimulants on the market |
| | Low side effects |
| | Reduction of pesticide use |
| | Promoted by EU Directive n. 128/2009 |
| | «Sustainable Use of Pesticides» and following National Action Plans |
| | Increased amounts of beneficial antioxidant compounds |